

Anisotropic pH-Responsive Hydrogels Containing Soft or Hard Rod-Like Particles Assembled Using Low Shear

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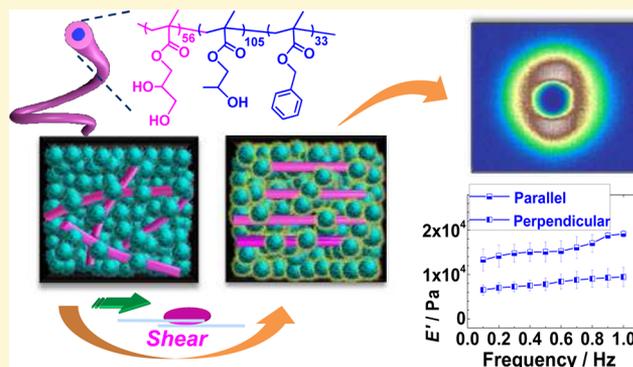
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Supporting Information

ABSTRACT: A simple and versatile low-shear approach for assembling hydrogels containing aligned rod-like particles (RLPs) that are birefringent and exhibit pH-triggered anisotropic swelling is developed. Anisotropic composite hydrogels are prepared by applying low shear (0.1 s^{-1}) to mixtures of pH-responsive nanogels (NGs) and RLPs. The NGs, which contained high methacrylic acid contents, acted as both shear transfer vehicles and macro-cross-linkers for anisotropic gel formation. Three model RLP systems are investigated: (i) soft triblock copolymer worms, (ii) stiff self-assembled β -sheet peptide fibers, and (iii) ultrahigh modulus nanocrystalline cellulose fibers. RLP alignment was confirmed using polarized light imaging, atomic force microscopy, and small-angle X-ray scattering as well as modulus and anisotropic swelling experiments. Unexpectedly, the composite gel containing the soft copolymer worms showed the most pronounced anisotropy swelling. The copolymer worms enabled higher RLP loadings than was possible for the stiffer RLPs. For fixed RLP loading, the extent of anisotropic swelling increased with intra-RLP bonding strength. The facile and versatile approach to anisotropic gel construction demonstrated herein is expected to enable new applications for strain sensing or biomaterials for soft tissue repair.



INTRODUCTION

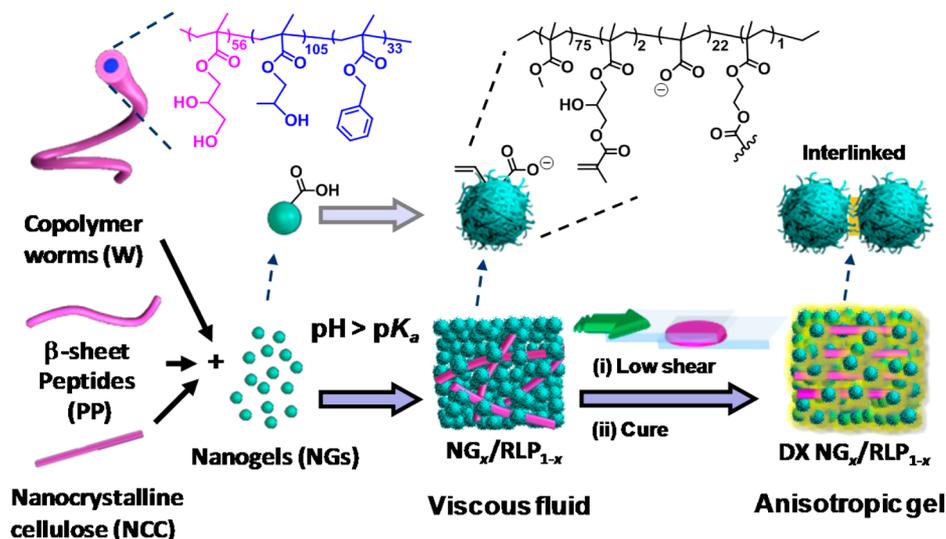
A major challenge for hydrogel research is the design of hydrogels with controlled morphology over a range of length scales. While many new hydrogels have been reported with excellent mechanical properties,^{1–10} there have been relatively few reports involving anisotropic composite gels.^{11,12} Such gels are gaining increasing attention in view of their potential applications in photonics,¹³ as well as biomaterials that mimic the extra-cellular matrix and direct tissue growth.¹⁴ Cells can sense and respond to nanoscale textures¹⁵ which provides the opportunity to guide repair of tissues such as artificial muscles,¹⁶ tendons,^{17,18} heart tissues,¹⁹ and neural path-

ways^{14,15,18,20–22} and within the spinal cord.²³ Hydrogels containing aligned rod-like particles (RLPs) are a potential route to anisotropic gels for such applications. Unfortunately, a facile and versatile method for permanently aligning RLPs within hydrogels is currently lacking. Usually, RLP alignment is achieved using high shear,^{24,25} cold drawing,²⁶ or solvent evaporation,²⁷ which constrain experimental versatility and the potential to include cells. High shear rates are known to

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Scheme 1. Preparation of Aligned Doubly-Crosslinked NG/RLP Gel Composites (DX NG_x/RLP_{1-x})^a

^aThe RLPs were copolymer worms comprised of poly(glycerol monomethacrylate)-*b*-poly(2-hydroxypropyl methacrylate)-*b*-poly(benzyl methacrylate), β -sheet peptide fibers, or nanocrystalline cellulose fibers. The NGs comprised poly(methyl methacrylate-*co*-methacrylic acid-*co*-ethyleneglycol dimethacrylate) and were vinyl functionalized. A viscous fluid formed when the pH of mixed NG_x/RLP_{1-x} (x = dry NG weight fraction) dispersions was increased. The latter was sheared, and the NGs were covalently interlinked to form an *anisotropic* composite gel.

decrease cell viability.²⁸ Other methods for obtaining anisotropic gels include freeze-casting,²⁹ imprinting,¹⁷ concentration via dialysis,¹⁸ magnetic fields,³⁰ or self-assembly.^{14,21,31,32} These methods either require conditions that are not cyto-compatible or involve specialized materials and equipment. Alignment methods involving low shear rates provide enhanced application potential with cells. Here, we investigate a new approach for anisotropic composite gel formation that is versatile and scalable and uses inexpensive laboratory equipment. We hypothesized that low shear could be used to align RLPs if they were dispersed in a mixture of soft (swollen) copolymer nanoparticles and that this alignment could be “locked in” if the nanoparticles could be covalently interlinked to form a network. Herein, we investigate anisotropic gels containing RLPs assembled using nanogels (NGs). NGs are cross-linked pH-responsive copolymer nanoparticles with a size less than 100 nm that swell when the pH approaches the pK_a of the constituent polymer.³³ These nanosized particles are smaller than their larger microgel cousins.^{34–37} NGs have a range of delivery and diagnostics applications.³⁸

Three RLP systems were used in this study: soft triblock copolymer worms (W), stiff β -sheet peptide fibers (PP), and ultrahigh modulus nanocrystalline cellulose (NCC). The copolymer worms were the most flexible, whereas NCC fibers were highly rigid and the PP fibers possessed intermediate stiffness. The shear-induced alignment of *small molecule* surfactant worms in the presence and absence of spheres has been studied.^{39–41} Those worms were “living” systems because the characteristic time for worm breaking (τ_{break}) was comparable to the experimental shear time (τ_{shear}). Mixtures of such surfactant worms with monomers commonly used to prepare hydrogels are unstable.⁴¹ Furthermore, orienting such worms within hydrogels is not currently possible to our knowledge. We overcame this obstacle by mixing non-living RLPs with NG macro-cross-linkers to construct anisotropic composite gels. The RLPs used here were less susceptible to breaking and the NGs acted as nanometer-scale gel building

blocks. Triblock copolymer worms were the primary RLP system studied (Scheme 1). PP fibers and NCC were used to study the effects of intra-RLP bonding on anisotropic gel swelling and demonstrate versatility of our new, low-shear approach for preparing anisotropic gels.

To align RLPs under shear, the local shear rate must be high compared to the RLPs rotational relaxation time.⁴² The latter is determined by the RLP dimensions as well as colloidal and hydrodynamic interactions and the viscosity of the sheared fluid. Key parameters controlling the hydrodynamic interactions of mixed dispersions of rods and spheres are the ratio of the rod length (L) to the sphere diameter (D_{NG}), i.e., $\xi = (L/D_{\text{NG}})$. The ratio of the RLP diameter (D_{RLP}) to the sphere diameter ($q = (D_{\text{RLP}}/D_{\text{NG}})$) is also important. If $\xi \gg 1$, then q determines the nature of the hydrodynamic interactions.³⁹ We selected RLPs with large ξ values and used NGs to achieve q values close to unity. This combination provided efficient stress transfer from the NGs to the RLPs and enabled alignment at low shear. Furthermore, because the NGs scattered light weakly, they provided a transparent matrix which enabled RLP alignment to be probed using polarized light optical microscopy.

The three RLP systems examined (Scheme 1) have different types of intra-RLP bonding and hence τ_{break} values. Block copolymer worms are soft, flexible RLPs that have attracted considerable interest.^{43–48} They can be conveniently synthesized by polymerization-induced self-assembly (PISA)^{45,49} and are biocompatible.⁴⁶ The worms used for this study comprised a poly(glycerol monomethacrylate)-*b*-poly(2-hydroxypropyl methacrylate)-*b*-poly(benzyl methacrylate) (PGMA₅₆-PHPMA₁₀₅-PBzMA₃₃) triblock copolymer synthesized directly in water using PISA.⁵⁰ Concentrated worm gels have relatively low moduli⁴⁶ as a result of their relatively small τ_{break} values. Herein, we report for the first time their incorporation into composite gels as well as their alignment. In contrast to worms, β -sheet peptide fibers self-assemble owing to the attractive intermolecular H-bonding between the backbone amides of PP chains and π - π interactions of aromatic residues.⁵¹ PP gels

have a range of applications which include substrates for cell culture.^{51–53} They have also been used to form composites with temperature-responsive microgels.⁵⁴ PP fiber dispersions form relatively brittle gels at low concentration.^{55,56} Accordingly, PP physical gels have higher τ_{break} values than worms. NCC fibers, which are obtained from a renewable, biocompatible,⁵⁷ and abundant material, exhibit ultrahigh moduli^{58,59} (i.e., 150 GPa⁶⁰), have been used to reinforce conventional gels,⁶¹ and can form hierarchical structures.⁶² NCC fibers do not break down under normal rheological experimental time frames and so were selected in this study as an RLP system with an effectively infinite τ_{break} value. These RLPs enabled us to study the effect of τ_{break} value on gel properties.

We recently established a method for preparing *isotropic* hydrogels using pH-responsive vinyl-functionalized NGs as macro-cross-linkers.³³ Concentrated NG dispersions were prepared using a convenient scalable synthesis and formed space-filling physical gels upon raising the solution pH. These gels formed due to NG particle swelling and were covalently interlinked via peripheral vinyl groups to form pH-responsive hydrogels, which are termed doubly cross-linked NGs (DX NGs).³³ In the present study, the NGs played two crucial roles in the formation of anisotropic DX NG/RLP gels (Scheme 1): they transferred shear stress to the RLPs and also enabled the otherwise transient anisotropy generated under shear to be permanently locked in place.

In this paper, we first characterize the NG and copolymer worm binary mixtures. Shear-induced alignment of the worms and the formation of anisotropic DX NG/worm gels is then demonstrated using a new rheo-optical technique, small-angle X-ray scattering (SAXS), and anisotropic swelling studies. The DX NG/worm gels are shown to exhibit low cytotoxicity which is encouraging for future soft tissue repair applications. The versatility of our approach is then demonstrated using PPs and NCCs. Dynamic mechanical analysis data for the DX NG/NCC system demonstrate the presence of anisotropic modulus values. The aligned composite gels are studied using polarized light microscopy as well as pH-triggered swelling, and the relationship between intrarod bonding and anisotropic swelling is discussed. The new and versatile anisotropic gel assembly established here may lead to optical sensors using polarized light due to the strong correlation between birefringence and RLP orientation^{63,64} or next-generation gels for directing tissue growth *in vivo*.

EXPERIMENTAL SECTION

Materials. Methyl methacrylate (MMA, 98.5%), methacrylic acid (MAA, 99%), ethylene glycol dimethacrylate (EGDMA, 98%), benzyl methacrylate (BzMA, 96%), glycidyl methacrylate (GM, 97%), ammonium persulfate (APS, 98%), sodium dodecyl sulfate (SDS, 98.5%), *N,N,N',N'*-tetramethylethylenediamine (TEMED, 99%), and microcrystalline cellulose (20 μm) were purchased from Aldrich and used as received. Doubly distilled deionized water was used for all experiments. Glycerol monomethacrylate (GMA; 99.8%) was donated by GEO Specialty Chemicals (Hythe, UK) and used without further purification. 2-Hydroxypropyl methacrylate (HPMA, 97%) and 4,4'-azobis(4-cyanopentanoic acid) (ACVA; V-501; 99%) were purchased from Alfa Aesar (Heysham, UK) and used as received. 2-Cyano-2-propyl dithiobenzoate (CPDB, 80% from ¹H NMR spectroscopy) was purchased from Strem Chemicals (Cambridge, UK). Tris(2-carboxyethyl)phosphine (TCEP hydrochloride, 99%) was purchased from Amresco (Solon, Ohio, USA). CD₃OD and dimethyl sulfoxide-*d*₆ (DMSO) were purchased from Goss Scientific (Nantwich, UK). All other solvents were purchased from Fisher Scientific (Loughborough, UK) and used as received.

Synthesis of Vinyl-Functionalized Nanogels (NGs). The NG particles were prepared via monomer-starved emulsion polymerization using a method described elsewhere³³ (see Scheme S1a). Briefly, SDS (1.2 g, 4.0 mmol) was dissolved in water (240 g), and the solution was purged with nitrogen before addition of APS (0.20 g, 0.90 mmol) in water (2.0 mL). A monomer feed comprising MMA (42.0 g, 0.42 mol), MAA (10.0 g, 0.12 mol), and EGDMA (1.1 g, 5.0 mmol) was added at a rate of 0.30 mL min⁻¹. The product was extensively dialyzed against water. For vinyl functionalization, GM (4.50 g, 0.39 mol) was added to the NG dispersion (120 g, 5.0 wt %) and the pH was adjusted to 5.1. The dispersion was stirred for 8 h at 40 °C. The dispersion was repeatedly washed with *n*-hexane, and residual solvent was removed by rotary evaporation.

Synthesis of PGMA₅₆. A protocol for the synthesis of PGMA₅₆ macro-CTA (chain transfer agent) is given in Scheme S1b. To a round-bottomed flask containing CPDB RAFT agent (6.03 g, 0.020 mol), GMA monomer (203 g, 1.27 mol) and anhydrous ethanol (156 g, 3.38 mol) were added to afford a target degree of polymerization (DP) of 63. ACVA initiator (1.14 g, 4.07 mmol, CTA/ACVA molar ratio = 5.0) was added, and the resulting pink solution was purged with N₂ for 20 min, before the sealed flask was immersed into an oil bath set at 70 °C. After 140 min (69% conversion as judged by ¹H NMR), the polymerization was quenched by immersion of the flask in an ice bath followed by exposure to air. The crude polymer was purified by precipitation into excess CH₂Cl₂ and washed three times in this solvent before being dried under high vacuum at 40 °C. ¹H NMR analysis indicated a DP of 56 (see Figure S1). DMF GPC analysis (Figure S2) indicated M_n and M_w/M_n values of 1.43×10^4 g mol⁻¹ and 1.14, respectively.

Synthesis of PGMA₅₆-PHPMA₁₀₅-PBzMA₃₃ Triblock Copolymer. The method used is depicted in Scheme S1b. PGMA₅₆ macro-CTA (4.28 g, 0.28 mmol) was added to a 100 mL round-bottomed flask, followed by HPMA monomer (0.25 g, 0.03 mol) and water (39.0 g, to make a 15% w/w solution). ACVA was then added (0.02 g, 0.07 mmol, CTA/ACVA molar ratio = 4.0), and the solution was purged with N₂ for 30 min. The sealed flask was immersed in an oil bath set at 70 °C. The solution was magnetically stirred for 2.5 h to ensure complete monomer conversion (>99% as judged by ¹H NMR in CD₃OD), and the polymerization was quenched by cooling the flask to 20 °C and exposure to air. BzMA monomer (1.64 g, 9.33 mmol) was then immediately added along with additional ACVA (20.0 mg, 0.07 mmol) before the reaction mixture was degassed with N₂ for a further 30 min. The flask was then replaced in the oil bath at 70 °C for a further 5 h, and the second-stage BzMA polymerization was quenched by exposure to air while at 20 °C. A sample of the copolymer dispersion was freeze-dried and dissolved in DMSO for ¹H NMR analysis (see Figure S1), which indicated that essentially all of the BzMA monomer had been incorporated into the copolymer. DMF GPC analysis indicated $M_n = 30\,600$ g mol⁻¹ and $M_w/M_n = 1.13$ for the PGMA₅₆-PHPMA₁₀₅ diblock copolymer and $M_n = 33\,200$ g mol⁻¹ and $M_w/M_n = 1.13$ for PGMA₅₆-PHPMA₁₀₅-PBzMA₃₃ triblock copolymer (see Figure S2).

Preparation of Self-Assembled Peptide Fibers (PP). The peptide studied was FEFEFKFE (F for phenylalanine, E for glutamate, and K for lysine residues), which was purchased from BIOMATIK Corporation (Delaware, USA) with more than 95% purity. Its identity and purity were confirmed in-house by ESI-MS and reverse phase HPLC, respectively. To prepare fibers, the peptide (19.4 mg, 7.0 μmol) was dissolved in HPLC-grade water (700 μL) by sonication for 30 min at 80 kHz. The peptide solution (pH 1.8) was titrated using 0.5 M NaOH to trigger gelation. A hydrogel was obtained at pH 5.3. The gel volume was adjusted to 1.0 mL using water to obtain a 14 mM (~1.9% w/v) hydrogel concentration. The gel was allowed to equilibrate overnight at 4 °C and used within 24 h.

Preparation of Nanocrystalline Cellulose. NCC was prepared by acid treatment of microcrystalline cellulose using a literature protocol.⁶⁵ Briefly, microcrystalline cellulose (5.0 g) was mixed with water (18 g), and the resulting suspension was placed in an ice bath. Sulfuric acid (32 g of 98% solution) was added slowly, and the mixture stirred for 150 min at 45 °C. [Caution: Concentrated acids are corrosive.]

The product was purified by repeated centrifugation and redispersion in water.

Preparation of Anisotropic Doubly Cross-Linked Gel Composites. The NG and worm (W) proportions in the gel composites are reported as DX NG_x/W_{1-x}, where *x* is the dry weight fraction of NG used to prepare each sample. The following gives the preparation protocol for DX NG_{0.5}/W_{0.5}, which was prepared at a particle concentration of 12 wt % at pH 7.5 by vortex-mixing a NG dispersion (0.70 g, 14.7 wt %) with an aqueous worm dispersion (0.60 g, 17.1 wt %) and water (0.285 g). An APS solution (60 μL, 78 mM) was added followed by addition of an alkaline TEMED solution (70 μL). The latter was prepared by mixing 4.0 M NaOH with TEMED at a volume ratio of 24:1. The pH increase produced a physical gel containing singly cross-linked NGs, denoted NG_{0.5}/W_{0.5}. For DX NG/worm composites with different *x* values, the masses of NG and worms were adjusted and the amounts of APS and NaOH/TEMED changed in proportion to the NG content. Unless otherwise stated, the total copolymer content (worms and/or NG) was 12.0 wt %. Anisotropic DX NG/worm composites were prepared by aligning worms during in situ cross-linking of the physical gels. Rotational shear or linear shear was used with either planar or cylindrical geometries. For the former, an Anton Paar Physica MCR-301 rheometer equipped with a SIPLI device was employed (see below). A shear rate of 0.1 s⁻¹ and a gap of 200 μm were used. The temperature employed was 45 °C.

In the case of linear shear, the NG_x/W_{1-x} precursor physical gel was placed between two microscope slides fixed to the clamps of a World Precision Instruments (AL-1000) syringe pump (see Figure S3). The gap between the two microscope slides was 500 μm, and the shear rate was 0.1 s⁻¹. A heated air flow was carefully directed over the top of the uppermost microscope slide to give a surface temperature of ~45 °C, and care was taken to minimize gel dehydration. Using the same protocol, anisotropic DX NG_x/PP_{1-x} gel composites were prepared by mixing 0.15 g of PP fibers (2 wt %) with NG (440 μL, 22 wt %). Then, APS (35 μL, 78 mM) was added, followed by NaOH/TEMED solution (30 μL). DX NG_{0.92}/NCC_{0.08} was prepared by mixing NCC (0.50 g of 2 wt %) with NG (440 μL, 22 wt %), followed by addition of APS (35 μL, 78 mM) and NaOH/TEMED (30 μL) solution. The mixtures were vortexed for 2 min prior to application of shear.

For shear alignment using a cylindrical geometry, strings of DX NG_{0.5}/W_{0.5} were prepared by injecting the NG_{0.5}/W_{0.5} precursor through a syringe needle (Gauge 18) onto a glass slide using a shear rate of 18 s⁻¹. Such strings were cured in a sealed Petri dish at 37 °C for 1 h.

Isotropic gel composites were prepared by placing the physical gel in an O-ring sealed between two microscope glass slides. The gels were cured at 37 °C overnight. Several control DX NG gels were also prepared using the same approach without the addition of copolymer worms (see Figure S5e,f).

Physical Measurements. Molecular weight data were obtained using a DMF GPC setup using poly(methyl methacrylate) calibration standards, as described earlier.⁶⁶ ¹H NMR spectroscopy was performed using a 400 MHz Bruker Avance-500 spectrometer. Potentiometric titration data were obtained in the presence of 0.05 M NaCl using a Mettler Toledo titrator. Dynamic light scattering (DLS) and electrophoretic mobility data were obtained using a Malvern Zetasizer NanoZS instrument and buffer solutions. A Hitachi U-1800 spectrophotometer was used for UV–visible spectroscopy measurements. An Olympus BX41 microscope was used to obtain optical microscopy images using transmitted light. For polarized light optical microscopy (POM), the light was passed through a polarizer and an analyzer, with the latter fixed at 90° with respect to the former. SEM and TEM images were obtained using Philips FEGSEM at 6 kV and Phillips CM100 at 100 kV instruments, respectively. AFM studies of the DX NG/worm composites were performed using a Bruker Catalyst (mounted on a Nikon Eclipse Ti inverted light microscope) with a Nanoscope V controller. Imaging was performed in ScanAsyst mode in liquid using ScanAsyst Fluid cantilevers (nominal spring constant, 0.7 nm/V; nominal probe radius, 20 nm (Bruker AXS, SAS)). For pure PP and NCC fibers, ScanAsyst mode in air was used.

Images were second-order flattened using the Bruker Nanoscope Analysis software.

SAXS data were obtained using a modified Bruker AXS Nanostar instrument equipped with a microfocus Xenocs Genix 3D X-ray source (Cu Kα radiation), a collimator comprised of two sets of motorized scatterless slits, a camera length of 1.46 m, and a 2D HiSTAR multiwire gas detector (pixel size, 0.105 mm²). 2D SAXS patterns were collected from sample films (500 μm thickness) sealed between two mica disks (25 μm thickness each). The scattering data were reduced to 1D plots by Nika SAS macros for Igor Pro.⁶⁷

Shear-induced polarized light imaging (SIPLI) measurements were conducted using a prototype rheometer for mechano-optical rheology. A mechanical Anton Paar Physica MCR 301 rheometer was coupled with an optical SIPLI device based on principles of reflection polariscopy.⁶⁸ The optical part of the instrument enabled a polarized light image of the sample loaded in the rheometer to be obtained during shear flow. Samples were placed between two parallel horizontal plates of the rheometer (one of the plates was transparent for light), and a shear pulse was applied by rotating the top plate.

Conventional dynamic rheology measurements were performed using a TA Instruments AR G2 rheometer. A 20 mm diameter plate geometry was used, and the gap was 2500 μm. DMA measurements were performed using a TA-Q800 instrument equipped with the shear sandwich (shear) geometry at 25 °C. For these measurements, DX NG_{0.92}/NCC_{0.08} samples were prepared by vortexing NG (700 μL, 22 wt %) and NCC (400 μL, 3.6 wt %) followed by addition of APS (55 μL, 78 mM) and NaOH/TEMED (47 μL), mixing, and curing as described above.

Anisotropic Swelling Experiments. Anisotropic gel swelling was studied by allowing samples to reach swelling equilibrium in buffer solutions over a period of ~18 h. The dimensions for each swollen sample (*W* and *L*) were measured at pH 8.0, while dimensions corresponding to the collapsed state (*W*₀ and *L*₀) were measured at pH 5.0. A sketch of the geometries used is shown in Figure 3e. The anisotropic swelling parameter (*A*) was calculated⁶⁹ using

$$A = \frac{\alpha_W}{\alpha_L} - 1 \quad (1)$$

where α_W ($=W/W_0$) and α_L ($=L/L_0$) are the linear swelling ratios measured perpendicular and parallel to the shear direction, respectively.

Cytotoxicity Studies. The cytotoxicity of DX NG_{0.5}/W_{0.5} was examined in the presence of human nucleus pulposus (NP) cells. The latter were seeded onto 24-well plates at a density of 5×10^4 per well and cultured in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum (FBS, Gibco), antibiotic/antimycotic, and L-ascorbic acid 2-phosphate (10 μM) (Sigma-Aldrich, UK) in a humidified atmosphere containing 5% CO₂ at 37 °C for 24 h. The DX NG_{0.5}/W_{0.5} samples (20 mg) were washed in phosphate buffered saline (PBS) for 2 days prior the biocompatibility test, sterilized with 70% ethanol, and rinsed with sterile PBS. The gel samples were put into 0.4 μm cell-culture inserts (BD Biosciences, UK) that were placed in the well plate. Cell viability was tested by the MTT assay (Aldrich, UK) after 1, 4, and 7 days. An empty insert was used as the control. For each time point, three data points were collected using a BMG Labtech FLUOstar plate reader.

RESULTS AND DISCUSSION

Properties of Nanogels, Worms, and Mixed Dispersions. All of the materials used to construct the anisotropic composite gels in this study were prepared using scalable methods. The NGs and copolymer worms were synthesized by emulsion polymerization and RAFT solution polymerization, respectively (Scheme S1). The worms were characterized using ¹H NMR spectroscopy and GPC (Figures S1 and S2), and their composition is shown in Scheme 1. The NGs contained 22 mol % MAA as well as vinyl groups from GM (2 mol %) and had an

apparent pK_a of 7.1.³³ Their composition is also shown in Scheme 1.

The NG and worm morphologies were assessed using TEM (Figure 1a,b). The NGs were spheres with a number-average

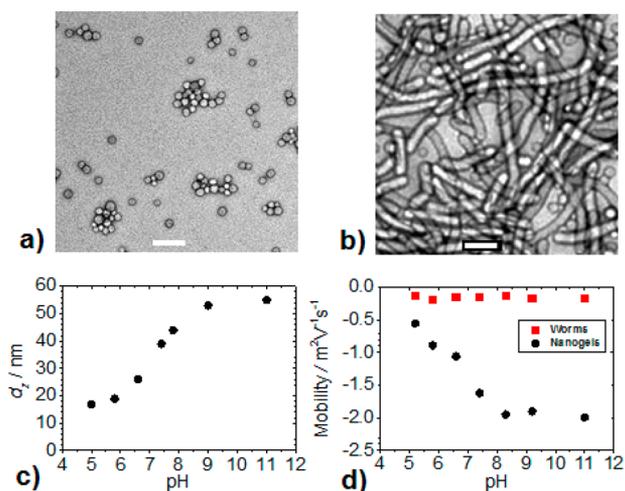


Figure 1. TEM images obtained for (a) NGs and (b) copolymer worms, respectively. The scale bars are 100 nm. (c) Variation of z -average diameter with pH for the NG particles. (d) Electrophoretic mobility vs pH for the NGs and copolymer worms.

diameter of 17 ± 3 nm. The anisotropic worms (Figure 1b) had a mean diameter of 35 ± 5 nm and a polydisperse length distribution with a number-average worm length of 310 ± 280 nm. High worm length polydispersities are typical for such syntheses and arise from the stochastic nature of worm formation via multiple 1D fusion of precursor spheres.⁴⁸ The NGs were pH-responsive, and the z -average DLS diameter (d_z) strongly increased from pH 5 to pH 11 (Figure 1c). The NGs had a d_z of 17 nm in the collapsed state (pH 5) but swelled up to 52 nm at pH 9. The d_z value was ~ 40 nm at pH ~ 7.5 and hence $q \sim 1$, as intended. The latter pH was used to prepare the composite gels.

One prerequisite for achieving shear-induced worm alignment in NG/RLP binary mixtures is that these two species do not attract each other because NG/RLP aggregate formation would otherwise oppose ordering. Electrophoretic mobility vs pH studies (Figure 1d) confirmed that the NGs were negatively charged and became more strongly anionic as the pH approached 7.5. The copolymer worms remained weakly anionic at all pH values studied. The anionic character observed for the NGs and worms was due to neutralization of MAA repeat units and also residual anionic carboxylate groups from the ACVA initiator,⁶⁶ respectively. Thus, there were no attractive electrostatic interactions between the NGs and worms.

Dilute binary NG/worm dispersions (NG_x/W_{1-x} , where x is the dry weight fraction of NG) were investigated using turbidity and DLS measurements to confirm that both components remained colloidally stable when mixed. Digital photographs of the sample vials (Figure S4a) did not show any evidence of visible aggregates. The optical density measured at 400 nm of the binary mixtures (Figure S4b) followed a linear relationship with x , indicating attractive interactions were not present between the NGs and worms. The absence of attractive interactions was also shown by DLS data (Figure S4c), which reported bimodal size distributions due to each (isolated)

component. In summary, the NGs and worms were both well dispersed in the mixed dispersions.

Low Shear Alignment and Assembly of Anisotropic DX NG/Worm Gels. A preliminary investigation was conducted of the ability of binary NG/worm dispersions to form isotropic covalent gels and to determine the compositional limits of DX NG_x/W_{1-x} systems (see Supporting Discussion and Figure S5). Briefly, isotropic gels could be formed provided x was greater than or equal to 0.35, which was identified from rheology data as the gel-to-fluid boundary. Whereas the DX NGs were covalently interlinked, the worms relied upon hydrophobic interactions and contributed less to the composite gel modulus. The τ_{break} value for the worms was identified as 1.0 from dynamic rheology data (Figure S6). MTT data obtained for nucleus pulposus cells in the presence of DX $NG_{0.5}/W_{0.5}$ composite gel were indistinguishable from a gel-free control (Figure S7) which indicated that significant release of toxic components did not occur over a period of 7 days.

Shear-induced worm alignment was investigated using a modified rheometer incorporating SIPLI, which enables in situ monitoring of the sample under shear (see Experimental Section).⁷⁰ Figure 2 shows polarized light images obtained for

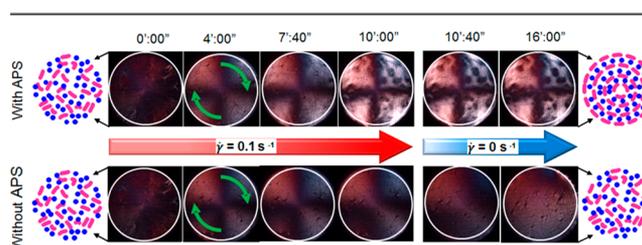


Figure 2. Rotational shear (γ) was applied to $NG_{0.5}/W_{0.5}$ binary mixtures while being viewed with polarized light using SIPLI. The samples were heated to 45°C , and the shear was terminated after 10 min. The top row shows the sample that underwent double cross-linking (and formed DX $NG_{0.5}/W_{0.5}$) during shear due to APS. The bottom row shows the control sample that did not contain APS. The green arrows indicate the direction of shear, and the cartoons depict the proposed initial and final composite gel morphologies.

two $NG_{0.5}/W_{0.5}$ physical gels. A mixed dispersion (top row) contained added APS initiator. The other control dispersion (bottom row) did not contain APS. Both binary gels were heated to 45°C , and characteristic Maltese cross patterns were observed under shear, indicating uniaxial alignment.^{70–72} After 10 min, the shear was terminated. The now doubly cross-linked APS-loaded gel retained the Maltese cross. In contrast, the Maltese cross rapidly disappeared for the control sample after shear was terminated. The NGs aligned the worms when sheared in both cases, but covalent interlinking of NG particles was required to “lock in” this alignment once shear was terminated. The NGs enabled the worms to be aligned at very low shear rates ($\sim 0.1 \text{ s}^{-1}$ at the sample edge). These results contrast to other studies which have required much higher shear rates (i.e., 200 s^{-1}) to align anisotropic particles in gels.^{25,73} In the present study, the ability to align the worms at low shear is attributed to the formation of a space-filling physical gel due to pH-triggered NG particle swelling and the mutual repulsion between the two colloidal components with comparable diameters (i.e., $q \sim 1$).

Building on the above results, we developed a simple linear shear method for aligning worms within DX NG/worm gels using inexpensive laboratory equipment (see Figure S3). $NG_x/$

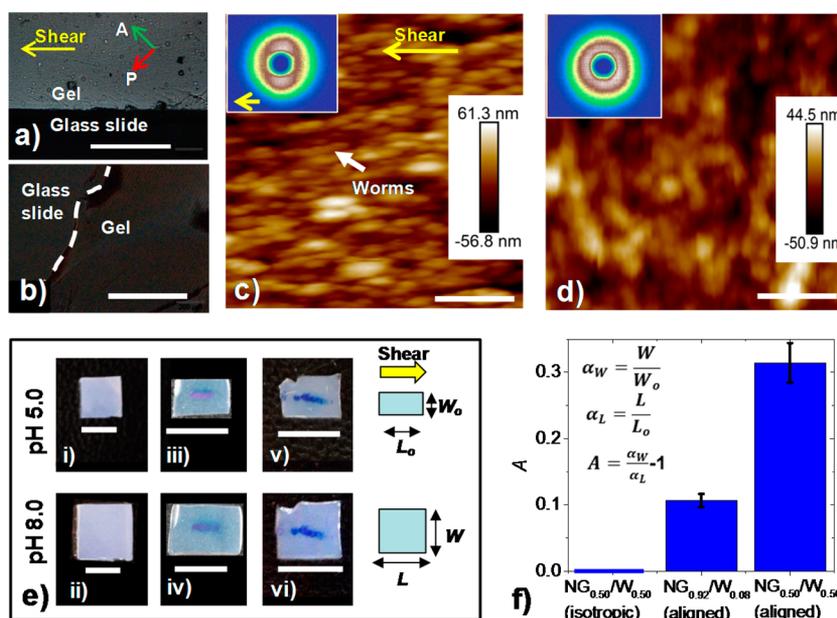


Figure 3. POM images of DX $\text{NG}_{0.5}/\text{W}_{0.5}$ gels prepared using (a) linear shear and (b) without shear. A and P in (a) are the directions of the analyzer and polarizer. (c, d) AFM images obtained for *anisotropic* and *isotropic* DX $\text{NG}_{0.5}/\text{W}_{0.5}$ composites in water, respectively. 2D SAXS patterns for each film are shown as insets. The shear direction for the inset in (c) is indicated. (e) Isotropic DX $\text{NG}_{0.5}/\text{W}_{0.5}$ (i, ii), anisotropic DX $\text{NG}_{0.92}/\text{W}_{0.08}$ films (iii, iv), and anisotropic DX $\text{NG}_{0.5}/\text{W}_{0.5}$ (v, vi) films at pH 5.0 or 8.0. Blue marker pen lines were used to record the shear directions. Scale bars: (a, b) 400 μm ; (c, d) 200 nm; (e) 6.0 mm. The cartoon depicts the anisotropic swelling. (f) Anisotropic swelling parameters (A) from the images shown in (e).

W_{1-x} physical gels were subjected to planar shear (0.1 s^{-1}) and simultaneously cured at $\sim 45 \text{ }^\circ\text{C}$. A polarized light optical microscopy (POM) image (Figure 3a) shows a bright area due to shear-aligned worms within the DX $\text{NG}_{0.5}/\text{W}_{0.5}$ gel. A control (*isotropic*) DX $\text{NG}_{0.5}/\text{W}_{0.5}$ gel that had not been subjected to shear is also shown (Figure 3b) for comparison. In that case, bright areas were not observed in *any* orientation, indicating that no alignment had occurred. The anisotropic DX $\text{NG}_{0.5}/\text{W}_{0.5}$ gels were examined using AFM (Figure 3c). Worm alignment in the direction of shear and an anisotropic morphology are evident. As a control, a nonaligned DX $\text{NG}_{0.5}/\text{W}_{0.5}$ control sample was examined using AFM (Figure 3d). The film showed randomly dispersed nano-objects and was isotropic. These data confirm that shear-induced alignment was achieved for planar DX NG/worm films using low shear.

SAXS is an ideal method for probing anisotropy because it is statistically robust and reports averaged structural information via scattering. 2D SAXS patterns were obtained for DX $\text{NG}_{0.5}/\text{W}_{0.5}$ films where the incident beam was perpendicular to the film plane. An anisotropic film (inset of Figure 3c) and a control isotropic film (inset of Figure 3d) were examined. Only the former gave a 2D pattern that was clearly elongated (Figure 3c), confirming worm alignment. The 2D SAXS patterns were reduced to 1D profiles both parallel and perpendicular to the direction of shear (see Figure S8). The SAXS data for both the aligned and isotropic films have intensity minima at $q_{\text{min}} \sim 0.028 \text{ \AA}^{-1}$ associated with the first minimum of the worm form factor and suggest that the radius of the worm core cross-section R_w was about 13.5 nm (using $q_{\text{min}}R_w = 3.83^{74}$). A higher scattering intensity was observed for the low q region for the aligned film which indicates the preferred orientation of the worms along the shear direction (Figure S8c). In contrast, there was no difference between the scattering profiles within the film plane for the isotropic, nonaligned film (Figure S8d).

Because our composites consisted of aligned (anisotropic) worms that contained a hydrophobic core dispersed within a gel matrix (of isotropic interlinked NGs), we investigated whether the composite gels exhibited anisotropic swelling. The anisotropic swelling parameter (A) was calculated using eq 1 (see also cartoon in Figure 3e and equations in Figure 3f). pH-triggered swelling experiments for the nonaligned DX $\text{NG}_{0.5}/\text{W}_{0.5}$ system confirmed they underwent *isotropic* swelling (Figure 3e(i,ii), $\alpha_L = \alpha_W$ and $A = 0$). Interestingly, the aligned DX $\text{NG}_{0.92}/\text{W}_{0.08}$ (Figure 3e(iii,iv)) and DX $\text{NG}_{0.5}/\text{W}_{0.5}$ gels (Figure 3e(v,vi)) both exhibited *anisotropic* swelling. The A values for the latter two systems were 0.11 ± 0.01 and 0.31 ± 0.03 , respectively. These A values are shown in Figure 3f (values for α_L , α_W , and A for all of the gels studied in this work are shown in Figure S9). The anisotropic gels swelled *least* in the direction parallel to worm alignment (lower α_L) because the worm cores were connected and extended within this plane (which contrasts to the perpendicular plane). The hydrophobic worm cores behaved as physical cross-links and opposed swelling. The extent of DX $\text{NG}_x/\text{W}_{1-x}$ anisotropic swelling increased with increasing worm content (i.e., decreasing x) because the contribution from the physical cross-links of the worm cores increased most in the parallel plane. These results demonstrate the ability to tune anisotropic swelling for these soft aligned gels by varying x .

Gel strings containing anisotropic RLPs are attracting considerable interest for biomaterials applications.¹⁴ As a proof-of-concept study to demonstrate versatility, we used a simple syringe to prepare anisotropic DX $\text{NG}_{0.50}/\text{W}_{0.5}$ strings. A concentrated $\text{NG}_{0.5}/\text{W}_{0.5}$ physical gel was injected through a syringe needle (inner diameter = 0.84 mm). Such extrusion induced worm alignment via shear from the inner syringe needle walls. The physical gel was subsequently converted into the corresponding DX $\text{NG}_{0.50}/\text{W}_{0.5}$ string by heating at $37 \text{ }^\circ\text{C}$. Worm alignment within the DX $\text{NG}_{0.50}/\text{W}_{0.5}$ composite gel

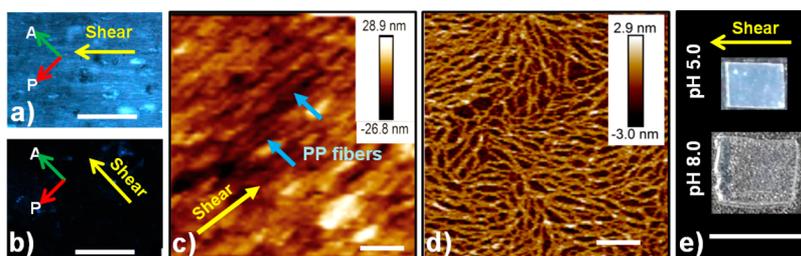


Figure 4. (a, b) POM images for aligned DX NG_{0.92}/PP_{0.08} composite gels. (b) The film was dark when the shear direction was parallel to the analyzer. (c, d) AFM images for aligned DX NG_{0.92}/PP_{0.08} and deposited PP fibers, respectively. PP fibers are highlighted in (c). (e) Images for pH-dependent swelling of DX NG_{0.92}/PP_{0.08}. The textured appearance is due to the background. Scale bars: (a, b) 400 μm ; (c, d) 200 nm; (e) 6.0 mm.

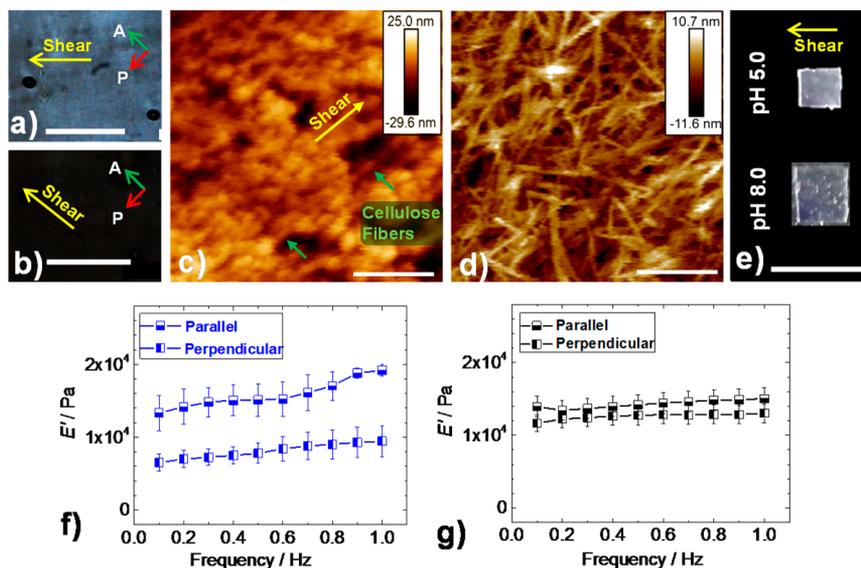


Figure 5. (a, b) POM images for a DX NG_{0.92}/NCC_{0.08} composite gel oriented at different angles with respect to the polarizer and analyzer. (c, d) AFM images for an anisotropic DX NG_{0.92}/NCC_{0.08} composite gel and the deposited pristine NCC fibers, respectively. (e) Anisotropic swelling images for a DX NG_{0.92}/NCC_{0.08} film. Scale bars: (a, b) 400 μm ; (c, d) 200 nm; (e) 6.0 mm. Storage modulus as a function of frequency for (f) anisotropic DX NG_{0.92}/NCC_{0.08} film and (g) control NCC-free DX NG ($x = 1.0$) measured parallel or perpendicular to the shear direction used during film formation.

string was confirmed by POM (Figure S10). Formation of an anisotropic composite gel string using such a simple protocol is rare in the literature.¹⁴

Anisotropic DX NG/RLP Gels Containing Peptides or Nanocrystalline Cellulose. To investigate the effects of intra-RLP bond strength and also to establish versatility of our approach, we prepared (linear) shear-aligned DX NGs gel films containing PP fibers. The PPs had a mean diameter of 16 ± 2 nm as determined by AFM. Frequency-sweep rheology data for a pure PP gel dispersion was measured (Figure S11), and G' and G'' did not intersect in the frequency range from 0.1 to 70 Hz. Consequently, we infer that τ_{break} was at least 10 s for this system. This relatively high value is most likely due to the stronger intra-RLP interactions. Figure 4a,b shows POM images obtained for DX NG_{0.92}/PP_{0.08}. The birefringence evident for this system is similar to that shown in Figure 3a and indicates alignment of the PP fibers. This birefringence disappeared when the sample was rotated through 45° (Figure 4b). AFM studies were conducted (Figure 4c) and confirmed that the peptide fibers were indeed aligned in the shear direction. In contrast, the AFM image for the parent peptide (Figure 4d) showed interconnected, randomly oriented fibers. The DX NG_{0.92}/PP_{0.08} gel films were examined for pH-triggered anisotropic swelling (Figure 4e). The A value of 0.14

± 0.02 was significantly greater than the value measured for DX NG_{0.92}/W_{0.08}. This result indicates that the intra-RLP bond strength dictates the extent of anisotropic swelling.

A RLP system derived from abundant biomass (cellulose) was also examined. The NCCs had a mean diameter and length of 10 ± 1 nm and 161 ± 30 nm, respectively, as determined from AFM. NCC fibers possess an ultrahigh modulus,⁶⁰ have strong covalent and hydrogen bonds within the RLP interior, and exhibit essentially infinite τ_{break} values. These RLPs were mixed with NGs for the preparation of anisotropic DX NG_{0.92}/NCC_{0.08} films. Birefringence was observed using POM (Figure 5a,b), which indicated NCC alignment. AFM data were also obtained for the sheared composite (Figure 5c) and provided evidence for NCC fiber alignment when compared to randomly dispersed NCCs present for a nonaligned sample of deposited pure NCCs (Figure 5d).

Anisotropic pH-dependent swelling (Figure 5e) occurred for the DX NG_{0.92}/NCC_{0.08} composite, and an A value of 0.15 ± 0.02 was measured. Interestingly, this value is indistinguishable from that obtained for DX NG_{0.92}/PP_{0.08} (Figure S9b shows all A values). As mentioned above, anisotropic swelling was caused by restriction of swelling parallel to the direction of shear imposed by intra-RLP bonding and was strongest for the RLPs with high τ_{break} values. It appears that once the intra-RLP bond

strength exceeds a critical value (which corresponds to a τ_{break} value greater than 10 s) the extent of anisotropic swelling reaches a maximum for a given x value. Following the example of the DX NG_{*x*}/W_{1-*x*} gels (Figure 3f), further increases for A should be possible by increasing x . However, a limitation with stiff RLPs is that their dispersion becomes increasingly difficult with increasing concentration because of entanglements. Attempts to prepare DX NG_{*x*}/W_{1-*x*} gels with x values of 0.10 or greater were unsuccessful because of NCC entanglements which prevented good dispersion being achieved prior to composite formation. This constraint was not present for the soft worms, and they were able to be loaded into the gels at higher concentrations (e.g., 50% dry weight), while retaining good dispersion, to give larger anisotropic swelling (Figure 3f). Thus, unexpectedly, the use of soft worm RLPs enabled the preparation of films with the most pronounced anisotropic swelling and highest A value compared to that which could be achieved using hard RLPs.

Dynamic mechanical analysis (DMA) was used to assess the anisotropic mechanical properties of the DX NG_{0.92}/NCC_{0.08} composite. For this purpose, we used a DMA equipped with a shear sandwich clamp which applied dynamic planar shear stress. The DMA data (Figure 5f) show that the frequency-dependent storage modulus (E') values for aligned DX NG_{0.92}/NCC_{0.08} measured parallel to the shear direction ranged from 1.3×10^4 to 1.9×10^4 Pa and were about a factor of 2 higher than the values of those measured perpendicular to the shear direction (which ranged from 6.5×10^3 to 9.5×10^3 Pa). This implies a doubling of the number of elastically effective chains for the gel in the parallel plane and is due to the contribution of the aligned ultrahigh modulus NCCs. In contrast, the mechanical properties of a control pure DX NG sample ($x = 1.0$) subject to planar shear did not show any dependence on the direction of applied shear (Figure 5g). The data confirm that these new anisotropic gels also have strongly anisotropic mechanical properties.

Table 1 summarizes the dimensions of the RLPs and NGs used in this study. For all RLP/NG mixtures explored, q was

Table 1. Summary of Structural Parameters for the NGs and RLPs

system	τ_{break}^a /s	diameter/nm	length/nm	ξ^b	q^c
NG		40			
worm	1.0	35	307	18.0	0.90
PP	>10	16	— ^d		0.40
NCC	∞	10	161	9.5	0.25

^aCharacteristic RLP breaking time. ^b $\xi = L_{\text{RLP}}/D_{\text{NG}}$, where L_{RLP} and D_{RLP} are the length and diameter of the RLPs, respectively. The values shown were obtained from TEM or AFM (see text). D_{NG} is the diameter of the NGs. The d_z value measured at pH 7.5 was used for D_{NG} . ^c $q = D_{\text{RLP}}/D_{\text{NG}}$. ^dIt was not possible to determine the length of the peptide fibers due to their branched nature.

comparable to unity, whereas ξ greatly exceeded unity. It was not possible to determine the precise value of ξ for NG_{*x*}/PP_{1-*x*} mixtures because the peptides had a pronounced tendency to form branches (Figure 4d). We propose that the use of NGs and RLPs with q reasonably close to unity was a key reason for achieving low shear alignment because of efficient stress transfer from the abundant NGs to the RLPs. Because all three systems exhibited shear-induced alignment, suitable design criteria for future DX NG_{*x*}/RLP_{1-*x*} formulations can

be suggested as $\xi \geq 9.5$ and q values of approximately 0.2 to 1.0, respectively. Furthermore, the τ_{break} values should be at least 1 s. These parameters should enable different RLPs to be utilized, ranging from water-dispersible multiwalled carbon nanotubes⁷⁵ to the tobacco mosaic virus.⁷⁶ Furthermore, NGs are very well-suited for systematic variation in size and/or internal modulus, which should further increase versatility.

An interesting question concerning our new approach for preparing anisotropic hydrogels is whether traditional linear cross-linkable polymer molecules could be used in place of the NGs. Our NGs were designed to efficiently transfer stress to the RLPs due to their q values (which were near to 1) and swollen nature. Linear polymer molecules typically have coil diameters that are at least an order of magnitude smaller than the RLP diameters, and hence, the q values would be very large. The latter would contribute to inefficient stress transfer. Linear polymer chains would also deform greatly under shear, dissipating energy, and would transfer stress less efficiently compared to our NGs. In contrast, our internally cross-linked NGs have an inherent elastic component which distributes stress to neighboring RLPs. While aligned RLP composites could be prepared, in principle, using traditional linear cross-linkable polymer chains, it is very likely that much higher shear rates would be needed compared to the low shear rates used in this study with NGs. The ability to use low shear rates is potentially beneficial for future work involving cells.

CONCLUSIONS

This paper reports a new low-shear approach for the preparation of anisotropic gel composites. Key to the success of our approach was the NGs which enabled low-shear alignment of three types of RLPs with varying intra-RLP stiffness (triblock copolymer worms, PPs, or NCC fibers). The worms were aligned using three different shear regimes: rotational shear, linear shear, and also via extrusion using a syringe. All three types of DX NG/RLP systems exhibited shear-induced birefringence and pH-triggered anisotropic swelling, which demonstrates that our new approach to forming anisotropic composite gels is versatile. The ability of NG particles to lock in the alignment of RLPs at low shear was critical for success, should also apply to other RLPs, and would in principle also be compatible with cells. Remarkably, the softest RLPs (worms) enabled preparation of aligned films with the strongest anisotropic swelling. This unexpected result was due to the ability to disperse the worms within the gel composites at high concentrations. It is highly likely that increases in the extent of anisotropic swelling can be achieved in the future through optimization of the NG and RLP properties. We stress that the NGs were prepared using a scalable, simple emulsion polymerization approach³³ and that alignment was achieved using inexpensive laboratory equipment. The DX NG/worm gel binary mixtures were not cytotoxic, and anisotropic swelling could be induced at around physiological pH; thus, these formulations are potentially useful new biomaterials. This suggestion also applies to the anisotropic gels containing PPs and NCCs because these RLPs are known to be biocompatible.^{51,77} Possible applications include strengthening and guiding regeneration of load-bearing soft tissues (e.g., intervertebral discs⁷⁸). In this context, the observation of worm alignment simply via injection is promising (Figure S10). Furthermore, all of the DX NG/RLP composite gels exhibited strain-induced birefringence

when viewed under polarized light, which suggests potential for strain sensor applications.^{63,79}

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.chemmater.7b00110.

Synthesis scheme for NGs and copolymer worms, ¹H NMR spectra and GPC for the copolymers and precursors, schematic representation of the shear-aligned gel film preparation method, characterization data for mixtures of NGs and copolymer worms, discussion of assembly and properties of isotropic DX NG/worm gels including SEM and dynamic rheology data, frequency-sweep data for concentrated worm dispersion, MTT data for DX NG_{0.5}/W_{0.5} gel, SAXS data for DX NG_{0.5}/W_{0.5} gels, swelling ratios and A values for all gels studied, photographs of DX NG_{0.5}/W_{0.5} string after injection through a syringe needle, frequency-sweep rheology data for a peptide gel (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Gong, J. P.; Katsuyama, Y.; Kurokawa, T.; Osada, Y. Double-Network Hydrogels with Extremely High Mechanical Strength. *Adv. Mater.* **2003**, *15*, 1155–1158.
- (2) Haque, M. A.; Kurokawa, T.; Gong, J. P. Super tough double network hydrogels and their application as biomaterials. *Polymer* **2012**, *53*, 1805–1822.
- (3) Haraguchi, K.; Takehisa, T. Nanocomposite Hydrogels: A Unique Organic–Inorganic Network Structure with Extraordinary Mechanical, Optical, and Swelling/De-swelling Properties. *Adv. Mater.* **2002**, *14*, 1120–1124.
- (4) Sun, J.-Y.; Zhao, X.; Illeperuma, W. R. K.; Chaudhuri, O.; Oh, K. H.; Mooney, D. J.; Vlassak, J. J.; Suo, Z. Highly stretchable and tough hydrogels. *Nature* **2012**, *489*, 133–136.
- (5) Yasuda, K.; Ping Gong, J.; Katsuyama, Y.; Nakayama, A.; Tanabe, Y.; Kondo, E.; Ueno, M.; Osada, Y. Biomechanical properties of high-toughness double network hydrogels. *Biomaterials* **2005**, *26*, 4468–4475.
- (6) Jeon, I.; Cui, J.; Illeperuma, W. R. K.; Aizenberg, J.; Vlassak, J. J. Extremely Stretchable and Fast Self-Healing Hydrogels. *Adv. Mater.* **2016**, *28*, 4678–4683.
- (7) Taylor, D. L.; in het Panhuis, M. Self-Healing Hydrogels. *Adv. Mater.* **2016**, *28*, 9060–9093.
- (8) Rodell, C. B.; Dusaj, N. N.; Highley, C. B.; Burdick, J. A. Injectable and Cytocompatible Tough Double-Network Hydrogels through Tandem Supramolecular and Covalent Crosslinking. *Adv. Mater.* **2016**, *28*, 8419–8424.
- (9) Luo, F.; Sun, T. L.; Nakajima, T.; King, D. R.; Kurokawa, T.; Zhao, Y.; Ihsan, A. B.; Li, X.; Guo, H.; Gong, J. P. Strong and Tough Polyion-Complex Hydrogels from Oppositely Charged Polyelectrolytes: A Comparative Study with Polyampholyte Hydrogels. *Macromolecules* **2016**, *49*, 2750–2760.
- (10) Feldner, T.; Häring, M.; Saha, S.; Esquena, J.; Banerjee, R.; Díaz, D. D. Supramolecular Metallogel That Imparts Self-Healing Properties to Other Gel Networks. *Chem. Mater.* **2016**, *28*, 3210–3217.
- (11) Moutos, F. T.; Freed, L. E.; Guilak, F. A biomimetic three-dimensional woven composite scaffold for functional tissue engineering of cartilage. *Nat. Mater.* **2007**, *6*, 162–167.
- (12) Wingfield, C.; Baski, A.; Bertino, M. F.; Leventis, N.; Mohite, D. P.; Lu, H. Fabrication of Sol–Gel Materials with Anisotropic Physical Properties by Photo-Cross-Linking. *Chem. Mater.* **2009**, *21*, 2108–2114.
- (13) Yue, Y.; Kurokawa, T.; Haque, M. A.; Nakajima, T.; Nonoyama, T.; Li, X.; Kajiwara, I.; Gong, J. P. Mechano-actuated ultrafast full-colour switching in layered photonic hydrogels. *Nat. Commun.* **2014**, *5*, 4659.
- (14) Zhang, S.; Greenfield, M. A.; Mata, A.; Palmer, L. C.; Bitton, R.; Mantei, J. R.; Aparicio, C.; de la Cruz, M. O.; Stupp, S. I. A self-assembly pathway to aligned monodomain gels. *Nat. Mater.* **2010**, *9*, 594–601.
- (15) Merzlyak, A.; Indrakanti, S.; Lee, S.-W. Genetically Engineered Nanofiber-Like Viruses For Tissue Regenerating Materials. *Nano Lett.* **2009**, *9*, 846–852.
- (16) Takashima, Y.; Hatanaka, S.; Otsubo, M.; Nakahata, M.; Kakuta, T.; Hashidzume, A.; Yamaguchi, H.; Harada, A. Expansion–contraction of photoresponsive artificial muscle regulated by host–guest interactions. *Nat. Commun.* **2012**, *3*, 1270.
- (17) Pei, X.; Zan, T.; Li, H.; Chen, Y.; Shi, L.; Zhang, Z. Pure Anisotropic Hydrogel with an Inherent Chiral Internal Structure Based on the Chiral Nematic Liquid Crystal Phase of Rodlike Viruses. *ACS Macro Lett.* **2015**, *4*, 1215–1219.
- (18) Wu, Z. L.; Kurokawa, T.; Sawada, D.; Hu, J.; Furukawa, H.; Gong, J. P. Anisotropic Hydrogel from Complexation-Driven Reorientation of Semirigid Polyanion at Ca²⁺ Diffusion Flux Front. *Macromolecules* **2011**, *44*, 3535–3541.
- (19) Chung, C.-Y.; Bien, H.; Entcheva, E. The Role of Cardiac Tissue Alignment in Modulating Electrical Function. *J. Cardio. Electrophys.* **2007**, *18*, 1323–1329.
- (20) Byun, M.; Santangelo, C. D.; Hayward, R. C. Swelling-driven rolling and anisotropic expansion of striped gel sheets. *Soft Matter* **2013**, *9*, 8264–8273.
- (21) Huang, Z.; Lee, H.; Lee, E.; Kang, S.-K.; Nam, J.-M.; Lee, M. Responsive nematic gels from the self-assembly of aqueous nanofibres. *Nat. Commun.* **2011**, *2*, 459.
- (22) Shigekura, Y.; Chen, Y. M.; Furukawa, H.; Kaneko, T.; Kaneko, D.; Osada, Y.; Gong, J. P. Anisotropic Polyion-Complex Gels from Template Polymerization. *Adv. Mater.* **2005**, *17*, 2695–2699.
- (23) Davies, S. J. A.; Goucher, D. R.; Doller, C.; Silver, J. Robust Regeneration of Adult Sensory Axons in Degenerating White Matter of the Adult Rat Spinal Cord. *J. Neurosci.* **1999**, *19*, 5810–5822.
- (24) Draper, E. R.; Mykhaylyk, O. O.; Adams, D. J. Aligning self-assembled gelators by drying under shear. *Chem. Commun.* **2016**, *52*, 6934–6937.
- (25) Haque, M. A.; Kamita, G.; Kurokawa, T.; Tsujii, K.; Gong, J. P. Unidirectional Alignment of Lamellar Bilayer in Hydrogel: One-Dimensional Swelling, Anisotropic Modulus, and Stress/Strain Tunable Structural Color. *Adv. Mater.* **2010**, *22*, 5110–5114.
- (26) Sehaqui, H.; Ezekiel Mushi, N.; Morimune, S.; Salajkova, M.; Nishino, T.; Berglund, L. A. Cellulose Nanofiber Orientation in Nanopaper and Nanocomposites by Cold Drawing. *ACS Appl. Mater. Interfaces* **2012**, *4*, 1043–1049.
- (27) Wang, P.-X.; Hamad, W. Y.; MacLachlan, M. J. Structure and transformation of tactoids in cellulose nanocrystal suspensions. *Nat. Commun.* **2016**, *7*, 11515.

- (28) Kong, H. J.; Smith, M. K.; Mooney, D. J. Designing alginate hydrogels to maintain viability of immobilized cells. *Biomaterials* **2003**, *24*, 4023–4029.
- (29) Chau, M.; De France, K. J.; Kopera, B.; Machado, V. R.; Rosenfeldt, S.; Reyes, L.; Chan, K. J. W.; Förster, S.; Cranston, E. D.; Hoare, T.; Kumacheva, E. Composite Hydrogels with Tunable Anisotropic Morphologies and Mechanical Properties. *Chem. Mater.* **2016**, *28*, 3406–3415.
- (30) Inomata, K.; Iguchi, Y.; Mizutani, K.; Sugimoto, H.; Nakanishi, E. Anisotropic Swelling Behavior Induced by Helix–Coil Transition in Liquid Crystalline Polypeptide Gels. *ACS Macro Lett.* **2012**, *1*, 807–810.
- (31) van Kuringen, H. P. C.; Leijten, Z. J. W. A.; Gelebart, A. H.; Mulder, D. J.; Portale, G.; Broer, D. J.; Schenning, A. P. H. J. Photoresponsive Nanoporous Smectic Liquid Crystalline Polymer Networks: Changing the Number of Binding Sites and Pore Dimensions in Polymer Adsorbents by Light. *Macromolecules* **2015**, *48*, 4073–4080.
- (32) Kempe, M. D.; Scruggs, N. R.; Verduzco, R.; Lal, J.; Kornfield, J. A. Self-assembled liquid-crystalline gels designed from the bottom up. *Nat. Mater.* **2004**, *3*, 177–182.
- (33) Milani, A. H.; Saunders, J. M.; Nguyen, N. T.; Ratcliffe, L. P. D.; Adlam, D. J.; Freemont, A. J.; Hoyland, J. A.; Armes, S. P.; Saunders, B. R. Synthesis of polyacid nanogels: pH-responsive sub-100 nm particles for functionalisation and fluorescent hydrogel assembly. *Soft Matter* **2017**, *13*, 1554.
- (34) Tan, B. H.; Pelton, R. H.; Tam, K. C. Microstructure and rheological properties of thermo-responsive poly(N-isopropylacrylamide) microgels. *Polymer* **2010**, *51*, 3238–3243.
- (35) Richtering, W. Responsive Emulsions Stabilized by Stimuli-Sensitive Microgels: Emulsions with Special Non-Pickering Properties. *Langmuir* **2012**, *28*, 17218–17229.
- (36) Hendrickson, G. R.; Lyon, L. A. Microgel Translocation through Pores under Confinement. *Angew. Chem., Int. Ed.* **2010**, *49*, 2193–2197.
- (37) Saunders, B. R.; Laajam, N.; Daly, E.; Teow, S.; Hu, X.; Stepto, R. Microgels: From responsive polymer colloids to biomaterials. *Adv. Colloid Interface Sci.* **2009**, *147–148*, 251–262.
- (38) Wu, H.-Q.; Wang, C.-C. Biodegradable Smart Nanogels: A New Platform for Targeting Drug Delivery and Biomedical Diagnostics. *Langmuir* **2016**, *32*, 6211–6225.
- (39) Guu, D.; Dhont, J. K. G.; Lettinga, M. P. Dispersions and mixtures of particles with complex architectures in shear flow. *Eur. Phys. J.: Spec. Top.* **2013**, *222*, 2739–2755.
- (40) Croce, V.; Cosgrove, T.; Dreiss, C. A.; King, S.; Maitland, G.; Hughes, T. Giant Micellar Worms under Shear: A Rheological Study Using SANS. *Langmuir* **2005**, *21*, 6762–6768.
- (41) Frounfelker, B. D.; Kalur, G. C.; Cipriano, B. H.; Danino, D.; Raghavan, S. R. Persistence of Birefringence in Sheared Solutions of Wormlike Micelles. *Langmuir* **2009**, *25*, 167–172.
- (42) Winkler, R. G.; Mussawisade, K.; Ripoll, M.; Gompper, G. Rod-like colloids and polymers in shear flow: a multi-particle-collision dynamics study. *J. Phys.: Condens. Matter* **2004**, *16*, S3941–S3954.
- (43) Sugihara, S.; Blanazs, A.; Armes, S. P.; Ryan, A. J.; Lewis, A. L. Aqueous Dispersion Polymerization: A New Paradigm for in Situ Block Copolymer Self-Assembly in Concentrated Solution. *J. Am. Chem. Soc.* **2011**, *133*, 15707–15713.
- (44) Fielding, L. A.; Lane, J. A.; Derry, M. J.; Mykhaylyk, O. O.; Armes, S. P. Thermo-responsive Diblock Copolymer Worm Gels in Non-polar Solvents. *J. Am. Chem. Soc.* **2014**, *136*, 5790–5798.
- (45) Warren, N. J.; Armes, S. P. Polymerization-Induced Self-Assembly of Block Copolymer Nano-objects via RAFT Aqueous Dispersion Polymerization. *J. Am. Chem. Soc.* **2014**, *136*, 10174–10185.
- (46) Verber, R.; Blanazs, A.; Armes, S. P. Rheological studies of thermo-responsive diblock copolymer worm gels. *Soft Matter* **2012**, *8*, 9915–9922.
- (47) Jones, E. R.; Semsarilar, M.; Blanazs, A.; Armes, S. P. Efficient Synthesis of Amine-Functional Diblock Copolymer Nanoparticles via RAFT Dispersion Polymerization of Benzyl Methacrylate in Alcoholic Media. *Macromolecules* **2012**, *45*, 5091–5098.
- (48) Canning, S. L.; Smith, G. N.; Armes, S. P. A Critical Appraisal of RAFT-Mediated Polymerization-Induced Self-Assembly. *Macromolecules* **2016**, *49*, 1985–2001.
- (49) Charleux, B.; Delaittre, G.; Rieger, J.; D'Agosto, F. Polymerization-Induced Self-Assembly: From Soluble Macromolecules to Block Copolymer Nano-Objects in One Step. *Macromolecules* **2012**, *45*, 6753–6765.
- (50) Mable, C. J.; Thompson, K. L.; Derry, M. J.; Mykhaylyk, O. O.; Binks, B. P.; Armes, S. P. ABC Triblock Copolymer Worms: Synthesis, Characterization, and Evaluation as Pickering Emulsifiers for Millimeter-Sized Droplets. *Macromolecules* **2016**, *49*, 7897–7907.
- (51) Jayawarna, V.; Ali, M.; Jowitt, T. A.; Miller, A. F.; Saiani, A.; Gough, J. E.; Ulijn, R. V. Nanostructured Hydrogels for Three-Dimensional Cell Culture Through Self-Assembly of Fluorenylmethoxycarbonyl–Dipeptides. *Adv. Mater.* **2006**, *18*, 611–614.
- (52) Banwell, E. F.; Abelardo, E. S.; Adams, D. J.; Birchall, M. A.; Corrigan, A.; Donald, A. M.; Kirkland, M.; Serpell, L. C.; Butler, M. F.; Woolfson, D. N. Rational design and application of responsive [alpha]-helical peptide hydrogels. *Nat. Mater.* **2009**, *8*, 596–600.
- (53) Shirbin, S. J.; Karimi, F.; Chan, N. J.-A.; Heath, D. E.; Qiao, G. G. Macroporous Hydrogels Composed Entirely of Synthetic Polypeptides: Biocompatible and Enzyme Biodegradable 3D Cellular Scaffolds. *Biomacromolecules* **2016**, *17*, 2981–2991.
- (54) Clarke, K. C.; Lyon, L. A. Microgel Surface Modification with Self-Assembling Peptides. *Macromolecules* **2016**, *49*, 5366–5373.
- (55) King, P. J. S.; Lizio, M. G.; Booth, A.; Collins, R. F.; Gough, J. E.; Miller, A. F.; Webb, S. J. A modular self-assembly approach to functionalised beta-sheet peptide hydrogel biomaterials. *Soft Matter* **2016**, *12*, 1915–1923.
- (56) Bowerman, C. J.; Nilsson, B. L. Review self-assembly of amphipathic β -sheet peptides: Insights and applications. *Biopolymers* **2012**, *98*, 169–184.
- (57) Thérien-Aubin, H.; Wang, Y.; Nothdurft, K.; Prince, E.; Cho, S.; Kumacheva, E. Temperature-Responsive Nanofibrillar Hydrogels for Cell Encapsulation. *Biomacromolecules* **2016**, *17*, 3244–3251.
- (58) Gregorczyk, K. E.; Pickup, D. F.; Sanz, M. G.; Irakulis, I. A.; Rogero, C.; Knez, M. Tuning the Tensile Strength of Cellulose through Vapor-Phase Metalation. *Chem. Mater.* **2015**, *27*, 181–188.
- (59) Eichhorn, S. J. Cellulose nanowhiskers: promising materials for advanced applications. *Soft Matter* **2011**, *7*, 303–315.
- (60) Azizi Samir, M. A. S.; Alloin, F.; Dufresne, A. Review of Recent Research into Cellulosic Whiskers, Their Properties and Their Application in Nanocomposite Field. *Biomacromolecules* **2005**, *6*, 612–626.
- (61) Yang, J.; Han, C.-R.; Duan, J.-F.; Xu, F.; Sun, R.-C. Mechanical and Viscoelastic Properties of Cellulose Nanocrystals Reinforced Poly(ethylene glycol) Nanocomposite Hydrogels. *ACS Appl. Mater. Interfaces* **2013**, *5*, 3199–3207.
- (62) Parker, R. M.; Frka-Petesic, B.; Guidetti, G.; Kamita, G.; Consani, G.; Abell, C.; Vignolini, S. Hierarchical Self-Assembly of Cellulose Nanocrystals in a Confined Geometry. *ACS Nano* **2016**, *10*, 8443–8449.
- (63) Maeda, A.; Inoue, T.; Yamaguchi, M. Dynamical rigidity of cellulose derivatives in melts. *Polym. J. (Tokyo, Jpn.)* **2014**, *46*, 149–154.
- (64) Pellens, L.; Vermant, J.; Mewis, J. Deviations from the Stress–Optical Rule in Telechelic Associative Polymer Solutions. *Macromolecules* **2005**, *38*, 1911–1918.
- (65) Bondeson, D.; Mathew, A.; Oksman, K. Optimization of the isolation of nanocrystals from microcrystalline cellulose by acid hydrolysis. *Cellulose* **2006**, *13*, 171–180.
- (66) Lovett, J. R.; Ratcliffe, L. P. D.; Warren, N. J.; Armes, S. P.; Smallridge, M. J.; Cracknell, R. B.; Saunders, B. R. A Robust Cross-Linking Strategy for Block Copolymer Worms Prepared via Polymerization-Induced Self-Assembly. *Macromolecules* **2016**, *49*, 2928–2941.
- (67) Ilavsky, J. Nika: software for two-dimensional data reduction. *J. Appl. Crystallogr.* **2012**, *45*, 324–328.

(68) Mykhaylyk, O. O.; Warren, N. J.; Parnell, A. J.; Pfeifer, G.; Laeuger, J. Applications of shear-induced polarized light imaging (SIPLI) technique for mechano-optical rheology of polymers and soft matter materials. *J. Polym. Sci., Part B: Polym. Phys.* **2016**, *54*, 2151–2170.

(69) Wang, C.; Hu, Z.; Chen, Y.; Li, Y. Swelling Behavior of Polymer Gels with Built-In Anisotropy near the Volume-Phase Transition Point. *Macromolecules* **1999**, *32*, 1822–1827.

(70) Mykhaylyk, O. O.; Warren, N. J.; Parnell, A. J.; Pfeifer, G.; Laeuger, J. Applications of shear-induced polarized light imaging (SIPLI) technique for mechano-optical rheology of polymers and soft matter materials. *J. Polym. Sci., Part B: Polym. Phys.* **2016**, *54*, 2151–2170.

(71) Mykhaylyk, O. O. Time-resolved polarized light imaging of sheared materials: application to polymer crystallization. *Soft Matter* **2010**, *6*, 4430–4440.

(72) Mykhaylyk, O. O.; Parnell, A. J.; Pryke, A.; Fairclough, J. P. A. Direct Imaging of the Orientational Dynamics of Block Copolymer Lamellar Phase Subjected to Shear Flow. *Macromolecules* **2012**, *45*, 5260–5272.

(73) Haque, M. A.; Gong, J. P. Multi-functions of hydrogel with bilayer-based lamellar structure. *React. Funct. Polym.* **2013**, *73*, 929–935.

(74) Roe, R. J. *Methods of X-ray and Neutron Scattering in Polymer Science*; Oxford University Press: New York, 2000.

(75) Kennedy, A. J.; Hull, M. S.; Steevens, J. A.; Dontsova, K. M.; Chappell, M. A.; Gunter, J. C.; Weiss, C. A. Factors influencing the partitioning and toxicity of nanotubes in the aquatic environment. *Environ. Toxicol. Chem.* **2008**, *27*, 1932–1941.

(76) Maier, E. E.; Krause, R.; Deggelmann, M.; Hagenbuechle, M.; Weber, R.; Fraden, S. Liquidlike order of charged rodlike particle solutions. *Macromolecules* **1992**, *25*, 1125–1133.

(77) Zoppe, J. O.; Peresin, M. S.; Habibi, Y.; Venditti, R. A.; Rojas, O. J. Reinforcing Poly(ϵ -caprolactone) Nanofibers with Cellulose Nanocrystals. *ACS Appl. Mater. Interfaces* **2009**, *1*, 1996–2004.

(78) Milani, A. H.; Freemont, A. J.; Hoyland, J. A.; Adlam, D. J.; Saunders, B. R. Injectable Doubly Cross-Linked Microgels for Improving the Mechanical Properties of Degenerated Intervertebral Discs. *Biomacromolecules* **2012**, *13*, 2793–2801.

(79) Shikinaka, K.; Koizumi, Y.; Shigehara, K. Mechanical/optical behaviors of imogolite hydrogels depending on their compositions and oriented structures. *J. Appl. Polym. Sci.* **2014**, *132*, 41691.