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1 *Type of the Paper (Review)*

2 **pH dependent antimicrobial peptides and proteins,**
3 **their mechanisms of action and potential as**
4 **therapeutic agents**

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12

13 **Abstract:** Antimicrobial peptides (AMPs) are potent antibiotics of the innate immune system that
14 have been extensively investigated as a potential solution to the global problem of infectious
15 diseases caused by pathogenic microbes. A group of AMPs that are increasingly being reported are
16 those that utilise pH dependent antimicrobial mechanisms and here, we review research into this
17 area. This review shows that these antimicrobial molecules are produced by a diverse spectrum of
18 creatures, including vertebrates and invertebrates, and are primarily cationic although a number of
19 anionic examples are known. Some of these molecules exhibit high pH optima for their
20 antimicrobial activity but in most cases, these AMPs show activity against microbes that present
21 low pH optima, which reflects the acidic pH generally found at their sites of action, particularly the
22 skin. The modes of action used by these molecules are based on a number of major structure /
23 function relationships, which include metal ion binding, changes to net charge and conformational
24 plasticity, and primarily involve the protonation of histidine, aspartic acid and glutamic acid
25 residues at low pH. The pH dependent activity of pore forming antimicrobial proteins involves
26 mechanisms that generally differ fundamentally to those used by pH dependent AMPs, which can
27 be described by the carpet, toroidal pore and barrel-stave pore models of membrane interaction. A
28 number of pH dependent AMPs and antimicrobial proteins have been developed for medical
29 purposes and have successfully completed clinical trials, including kappacins, LL-37, histatins and
30 lactoferrin, along with a number of their derivatives. Major examples of the therapeutic application
31 of these antimicrobial molecules include wound healing as well as the treatment of multiple cancers
32 and infections due to viruses, bacteria and fungi. In general, these applications involve topical
33 administration, such as the use of mouth washes, cream formulations and hydrogel delivery
34 systems. Nonetheless, many pH dependent AMPs and antimicrobial proteins have yet to be fully
35 characterized and these molecules, as a whole, represent an untapped source of novel biologically
36 active agents that could aid fulfillment of the urgent need for alternatives to conventional antibiotics,
37 helping to avert a return to the pre-antibiotic era.
38

39 **Keywords:** antimicrobial peptides and proteins; pH dependent antimicrobial activity; invertebrates;
40 vertebrates;

41 **PACS:** J0101
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43 **1. Introduction**

44 A multiplicity of synergistic factors, including diminished pharmaceutical investment, clinical
 45 over-prescription and misuse by the food industry has led to the increasing occurrence of microbial
 46 pathogens with multiple drug resistance (MDR) and rendered infectious diseases the leading cause
 47 of global mortality [1]. This bleak situation led the World Health Organization (WHO) to recently
 48 predict that the uncurbed rise of MDR pathogens could see conditions in the 21st Century return to
 49 those of the pre-antibiotic era when no antimicrobials were available for the treatment of many
 50 common diseases [2]. In response, a major analysis by the WHO and a report from the O'Neill review,
 51 sponsored by the UK Government, have concluded that the problem of antimicrobial drug resistance
 52 can only be fully addressed by a coordinated global approach that operates through a number of
 53 major interventions (Table 1) [3]. In particular, intervention six (Table 1) proposed the urgent
 54 development of novel products and strategies that could provide alternatives to conventional
 55 antibiotics, which has generated intensive research into antimicrobial design [4]. Examples of this
 56 research range from revisiting old anti-infective strategies, such as phage therapy, which was popular
 57 in Eastern European countries in the early 20th Century [5], to recently reported antimicrobial
 58 strategies, such as the development of compounds whose antibiotic activity can be regulated by light
 59 and sound [4,6-8]. One particularly promising approach proposed by O'Neill [3] was the therapeutic
 60 development of antimicrobial peptides (AMPs), which are potent antibiotics of the innate immune
 61 system [9,10]. The activity of these peptides against microbes involves relatively non-specific modes
 62 of action at multiple sites with the result that microbial resistance to AMPs has a low incidence and
 63 is generally due to inherent rather than adaptive mechanisms [11]. Based on these observations, the
 64 generally held view is that microbial resistance to AMPs is unlikely to approach those of conventional
 65 antibiotics, endowing these peptides with a major medical advantage [10], and currently, a number
 66 of AMPs are in clinical trials (Table 2).
 67

68 **Table 1:** Major areas of intervention to combat antimicrobial drug resistance

1.	A global public awareness campaign.
2.	Improve sanitation and hygiene to prevent the spread of infection.
3.	Reduce the unnecessary use of antimicrobials in agriculture and their dissemination in the environment.
4.	Improve the global surveillance of drug resistance and antimicrobial consumption in humans and animals.
5.	Promote new rapid diagnostics to reduce use of unnecessary antimicrobials.
6.	Promote the development and use of vaccines and alternatives
7.	Improve the number, pay and recognition of people working in infectious disease.
8.	A global innovation fund for early stage and non-commercial research and development.
9.	Better incentives to promote investment for new drugs.

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Table 1 was derived from [3]

Table 2: Major examples of AMPs in clinical trials or in development

Antimicrobial peptides	Indication	Phase	Company
Pexiganan (MSI-78), an analogue of magainin.	Topical cream for the treatment of diabetic foot infections and ulcers.	3	Dipexium Pharma /MacroChem / Genaera
Iseganan (IB-367), a derivative of protegrin 1.	Mouthwash for the treatment of chemotherapy induced oral mucositis.	3	Ardea Biosciences / national Cancer Institute.
	Mouthwash for the treatment of ventilator-associated pneumonia.	3	IntraBiotics Pharmaceuticals.
PAC-113 (P-113) a synthetic derivative of histatin 3 and histatin 5.	Oral gel for the treatment of candidiasis		Pacgen Biopharmaceuticals
Omiganan (MBI 226, MX-226, CSL-001), an analogue of indolicidin.	Topical cream for the treatment of skin antisepsis, prevention of catheter infections / Rosacea.	3	Mallinckrodt / Cutanea Life Sciences, Inc.
	Topical cream for the treatment of usual type vulvar intraepithelial neoplasia / moderate to severe inflammatory acne vulgaris / mild to moderate atopic dermatitis.	3	Cutanea Life Sciences, Inc.
OP-145, a derivative of LL-37.	Ear drops for treatment of chronic bacterial middle-ear infection.	2	OctoPlus
hLF1-11, a derivative of lactoferrin.	Intravenous administration for treatment of neutropenic stem cell transplantation patients. Prevention of bacteraemia and fungal infections.	1/2	AM Pharma.
Brilacidin, (PMX-30063), a defensin mimetic.	Intravenous administration for treatment of acute bacterial skin and skin structure Infection caused by Gram-positive bacteria, including methicillin-resistant <i>Staphylococcus aureus</i> (MRSA).	3	Cellceutix.
	Oral rinse for the treatment of ulcerative mucositis associated with chemo / radiation therapy of cancer.	2	Cellceutix.
Arenicins, naturally occurring AMPs.	For the treatment of infections due to MDR Gram-positive bacteria.	Preclinical	Adenium Biotech
Novexatin (NP213), a synthetic AMP.	Brush on treatment for fungal infections of the toenail.	1/2	NovaBiotics
C16G2, a synthetic specifically targeted AMP.	Mouthwash for the treatment of tooth decay caused by <i>Streptococcus mutans</i>	2	C3 Jian, Inc.
Lytixar (LTX-109), a peptidomimetic.	Topical antibiotic for the treatment of nasal carriers of MRSA.	1/2	Lytix Biopharma.
	Topical cream for the treatment of infections due to Gram-positive bacteria.	2	Lytix Biopharma.

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Table 2 was derived from [12-15]

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In order to develop AMPs as medically relevant anti-infective agents, there have been numerous investigations into their antimicrobial mechanisms, which to date has shown that membrane interaction is a requirement for virtually all of these mechanisms [9,10,16,17]. These investigations have also shown that there are a number of major drivers in the membrane interactions of AMPs of which the most important are charge, hydrophobicity and amphiphilicity [9,18,19]. The vast majority of AMPs are cationic to help facilitate the targeting of microbes through direct electrostatic interaction with anionic components of their membranes [19,20]. Nearly all AMPs are also amphiphilic, which generates hydrophobic surfaces that are able to drive the partitioning of these peptides into microbial membranes and hydrophilic surfaces that are able to stabilize these hydrophobic interactions via electrostatic associations with the head group regions of these membranes [21,22]. Based on these investigations, a variety of models have been proposed to describe the antimicrobial action of AMPs with those most frequently reported appearing to be variants of the barrel stave pore and carpet type mechanisms, which involve membrane disruption via discrete channel formation and non-specific solubilisation respectively [23].

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There have been many advances in understanding the mode of action used by AMPs but although a number of earlier studies showed that pH can modulate the antimicrobial activity of these peptides, no major review of this area of research appears to have been presented in the literature [24-28]. However, it is now becoming increasingly clear that pH is a major driver in the membrane interactions and biological activity of not only many AMPs but also a number of antimicrobial proteins produced by eukaryotes (Table 3). To update on these antimicrobial molecules, here, we present an overview of recent progress in the understanding of their modes of action along with the development of their therapeutic and biotechnological potential.

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2. An overview of pH dependent peptides and proteins with antimicrobial activity

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In the 1980s and 1990s, a series of seminal studies, including work on the African clawed frog, *Xenopus laevis*, and a number of mammals, led to what many take to be the first major description of eukaryotic AMPs such as magainins, defensins and SAAPs [29]. However, in 1956, phagocytin from humans, rabbits, horses and guinea pigs was reported to exhibit non-membranolytic activity against a range of Gram-positive and Gram-negative bacteria that was enhanced by low pH [24,25]. The peptide was not characterised or further investigated and today, it is not even known as to whether phagocytin was rediscovered later and given an alternative name [30]. However, it would appear to be a matter of historical fact that what was most likely the first AMP to be reported from eukaryotes showed a pH dependent mode of action [24,25]. Since these earlier studies, it is now known that these peptides are produced by virtually all multicellular organisms [31,32] and that an increasing number of these molecules possess pH-dependent activity (Table 3).

Table 3. AMPs with pH dependent activity.

Vertebrates	AMPs	Host organism	Key references
Fish	Gaduscidin-1 and gaduscidin-2	<i>Gadus morhua</i>	[33,34]
Amphibians	Chensinin-1	<i>Rana chensinensis</i>	[35,36]
	Esculentin-2EM	<i>Glandirana emeljanovi</i>	This work
	Dermaseptin PD-3-7	<i>Pachymedusa dactylos</i>	[37]
Humans	Phagocytin		[24,25]
	Psoriasin		[38-40]
	β -microseminoprotein		[41].
	LL-37		[42]
	Hep-25 and hep-20		[43-48]
	Histatins		[49,50]
	Lactoferrin		[51]
	DCD-1(L)		[52-54]
	Kappacin A and kappacin B		[55,56]
Rabbits	Phagocytin		[24,25]
	Platelet microbiocidal proteins		[57]
	NP1 and NP2		[58,59]
Horses	Phagocytin		[24,25]
Guinea pigs	Phagocytin		[24,25]
Mice	CRAMP		[42]
Cattle	Lactoferricin B		[60,61].
Invertebrates	AMPs	Host organism	Key references
Marine	Myticin C	<i>Mytilus galloprovincialis</i>	[62,63]
	KPS-1	<i>Atrina pectinate</i>	[64]
	Ci-PAP-A22 and Ci-MAM-A24	<i>Ciona intestinalis</i>	[65-68]
	Clavaspilin and clavanins	<i>Styela clava</i>	[69-80]
	Styelins	<i>Styela clava</i>	[72,73,81]
Terrestrial	Hebraein	<i>Amblyomma hebraeum</i>	[82]
	Amoebapores	<i>Entamoeba histolytica</i>	[83-87]
	Acanthaporin	<i>Acanthamoeba culbertsoni</i>	[88]
	Caenopores	<i>Caenorhabditis elegans</i>	[89-93]

111 2.1. Fish

112 Gaduscidin-1 (gad-1) and gaduscidin-2 (gad-2) were AMPs identified in the Atlantic cod, *Gadus*
113 *morhua* and shown to be highly, constitutively expressed in immune-relevant tissues [94,95]. Low pH
114 was found to enhance the activity of both peptides against *Escherichia coli*, which appeared to involve
115 membrane interaction, but interestingly, although gad-1 and gad-2 were predominantly α -helical at
116 neutral pH, acid conditions led to a large decrease in the levels of α -helicity possessed by these
117 peptides [33]. These results contrast with most α -helical AMPs where an enhanced capacity for
118 membranolytic and antimicrobial activity is generally associated with increased levels of this
119 secondary structure [96,97]. It was proposed that gad-1 and gad-2 each possessed a structural
120 plasticity, which facilitated an appropriate balance between amphiphilic and mixed hydrophobic /
121 hydrophilic structural features that promoted maximal levels of membrane interaction and
122 antibacterial activity [33]. Gad-1 and gad-2 were found to be histidine rich AMPs and the enhanced
123 capacity of these peptides for membranolytic and antimicrobial activity at low pH appeared to

124 involve these residues [33]. It is well established that histidine (pKa 6.5) is uncharged at physiological
125 pH but fully positively charged at low pH, thereby enhancing the potential of AMPs for interaction
126 with anionic membranes under acid conditions [50,98]. However, there also appeared to be a complex
127 relationship between the level of histidine residues possessed by these AMPs and their pH dependent
128 capacity for membrane interaction [33,34]. In response, molecular dynamic simulation studies were
129 undertaken and predicted that the number of sequential histidine pairs contained by gad-1 and gad-
130 2 were important to their ability for membrane disruption [34]. These AMPs possess one and two of
131 these histidine pairs respectively [94] and the N-terminal regions of both peptides, which included
132 this motif, were preferentially located proximal to membrane channels with which gad-1 and gad-2
133 were associated [34]. Based on the topology of the peptide-lipid interactions mediating the formation
134 of these channels, it was suggested that the antimicrobial action of gad-1 and gad-2 may involve the
135 use of a disordered toroidal pore type mechanism of membrane disruption [34,99,100].

136 2.2. Amphibians

137 Chensinin-1 is a histidine rich peptide produced by the frog, *Rana chensinensis* and recent
138 studies showed that low pH enhanced the positive charge of the peptide [35] and thereby, its ability
139 to kill Gram-positive bacteria, such as *Bacillus cereus* [36]. This antibacterial activity appeared to
140 involve the adoption of an extended structure, similar to that of other AMPs that are rich in specific
141 residues [101,102], which induced lysis of the *B. cereus* membrane [36]. Interestingly, the peptide
142 showed no activity against Gram-negative bacteria [36,103], which appeared to involve high affinity
143 binding between chensinin-1 and lipopolysaccharide (LPS) in the outer membrane of these bacteria
144 [103]. Maximin H5 from the toad, *Bombina maxima*, was also recently found to be ineffective against
145 Gram-negative bacteria due to high affinity binding to phosphatidylethanolamine (PE) in the
146 cytoplasmic membrane (CM) of these organisms [104,105]. Similar PE mediated mechanisms have
147 been proposed to mediate the resistance of microbes to other AMPs [104,105], supporting the growing
148 view that receptors could play a variety of roles in the biological activities of these peptides [23,98,106-
149 108]. Esculentin-2EM (E2EM, previously gaegurin 4) is an α -helical peptide isolated from the frog,
150 *Glandirana emeljanovi* (formerly *Rana rugosa*) KIM [109,110], that is able to kill protozoa, fungi, Gram-
151 positive bacteria and Gram-negative bacteria [110-112]. E2EM possesses a C-terminal cyclic region
152 stabilized by a disulphide bond (Rana box) that is conserved across many ranid AMPs and helps
153 stabilise pore formation by the peptide thereby promoting its antimicrobial action (Figure
154 1)[109,110,112-115]. Several models have been proposed to represent pore formation by E2EM and
155 the best supported by experimental evidence appear to be the toroidal pore and barrel stave
156 mechanisms (Figure 1). Here we present data showing that the linear reduced form of E2EM (E2EM-
157 lin) possesses antimicrobial activity consistent with recent studies showing that the reduction of
158 cysteine-stabilized AMPs to generate peptides with novel mechanisms of antimicrobial activity may
159 form part of some innate immune systems [116,117]. Our results showed that E2EM-lin was active
160 against both Gram-positive and Gram-negative bacteria and appeared to exhibit pH dependent
161 antimicrobial activity, which parallels the molluscan cysteine stabilized AMPs, myticins, whose
162 reduced forms were described above to show pH dependent antibacterial and antiviral action [62,63].
163 It was found that under the low pH conditions associated with the skin of frogs [118], E2EM-lin had
164 a general ability to induce the lysis of both anionic and zwitterionic membranes, which was enhanced
165 at higher pH (Table 4). These data clearly suggest that the C-terminal disulphide bond in the E2EM
166 Rana box region does not play a major role in its ability for membrane pore formation (Figure 1),
167 which supports earlier work [114,115]. In the case of DMPC, which is a key component of membranes
168 within Gram-positive bacteria [119], E2EM-lin induced relatively low levels of membrane lysis at acid
169 pH (< 25%). However, a shift to alkaline pH led to a large increase in the lytic activity of the peptide
170 to circa 95%, which was accompanied by a correspondingly large increase in its α -helical content of
171 circa 25% to give levels approaching 75% (Table 4). Previous studies have shown that E2EM-lin and
172 E2EM adopt highly similar α -helical structures in membranes and undergo oligomerization to form

173 pores [115,120]. Based on these data, we speculate that high pH may enhance the ability of E2EM-lin
 174 to lyse DMPG membranes by increasing the potential of the peptide for pore formation through an
 175 increased capacity for self-association. The segments of E2EM-lin involved in pore formation are
 176 strongly amphiphilic α -helices with wide hydrophobic faces that would be maximized by alkaline
 177 pH, promoting the potential for the mutual interactions involved in the formation of multimeric
 178 species (Figure 1). In the case of DMPE, which is often taken to represent the membranes of Gram-
 179 negative bacteria [119], E2EM-lin induced high levels of membrane lysis at acid pH (60%). A move to
 180 alkaline pH led to a relatively low increase in the lytic activity of the peptide, which was around 25%
 181 and was accompanied by a decrease in its α -helical content of 20% to give levels of circa 30% (Table
 182 4). These data would seem to indicate that E2EM-lin uses a different mechanism of lysis in the case
 183 of DMPE membranes and it has previously been suggested that the peptide may adopt a number of
 184 lipid interactive forms [115]. The functional significance of our data is not fully clear but given the
 185 very high levels of membrane lysis induced by E2EM-lin at higher pH for each lipid investigated,
 186 biological relevance is suggested, which forms the basis of ongoing investigations.
 187

188 **Table 4.** The α -helical content and lysis levels of E2EM-lin

Lipid	pH	Lysis (%)	α -helicity (%)
DMPS	6	17	30
	8	63	49
DMPG	6	23	51
	8	94	73
DMPC	6	52	45
	8	73	15
DMPE	6	60	49
	8	83	31

189 The levels of lysis exhibited by E2EM-lin were determined using a calcein release assay and the levels of α -
 190 helicity shown by the peptide were measured using CD spectroscopy, all as previously described [105].

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Figure 1. Models for the membrane pore formation by E2EM

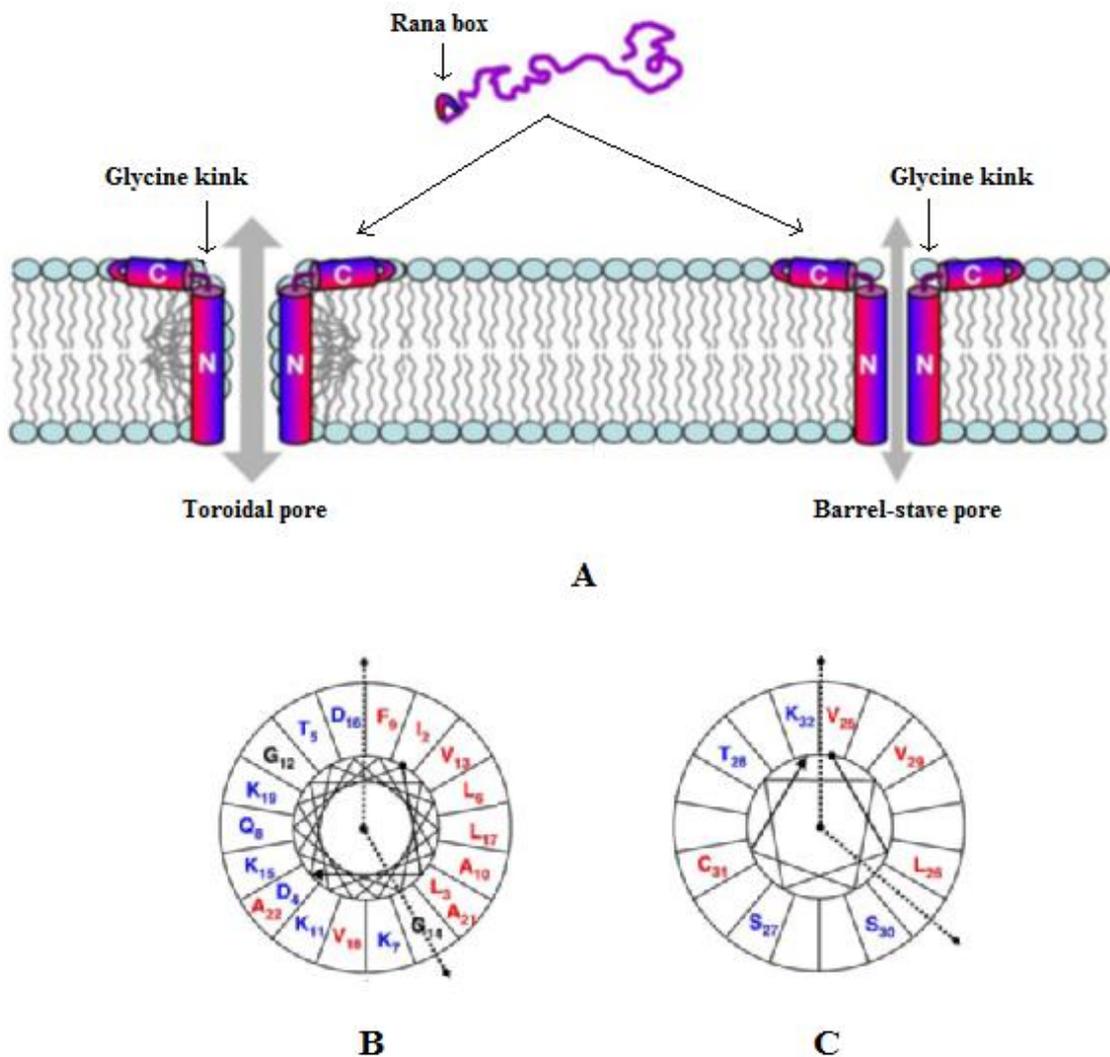


Figure 1 was revised from [115] and Figure 1A shows models for pore formation by E2EM, which are the toroidal pore and barrel stave mechanisms (Table 1) and are the best supported experimentally. In both models, the N-terminal 23 residues of the peptide spans the bilayer and a glycine kink orientates the 7 residue, C-terminal Rana box region of E2EM to lie parallel to the membrane surface. In this orientation, the Rana box region of the peptide, which is a cystein stabilized macrocyclic structure, interacts with the lipid head-group region of the membrane and stabilises pore formation by E2EM [115]. The major difference between these models is that in the toroidal pore mechanism, the membrane leaflets deform to allow the lipid head-group region to remain in contact with the hydrophilic face of the E2EM membrane spanning region, which is not observed in the barrel stave mechanism [23]. For clarity, two monomers of E2EM are shown in the schematic pore above but oligomers formed by between five and ten peptide molecules have been proposed [115,120]. Similar models of membrane interaction appear to apply to the linear reduced form of the peptide [115], which is represented in our studies as E2EM-lin. Figures 1B and 1C show two-dimensional axial projections [121] for the membrane spanning region and Rana box domain of E2EM, respectively, that are involved in pore formation by the peptide. In both cases, these segments for amphipilic α -helices with wide hydrophobic faces that our data suggest would be maximised by alkaline pH, thereby promoting the potential for the mutual interaction of E2EM monomers and the formation of multimeric species involved in pore formation.

218 In addition to cationic AMPs, anionic AMPs with pH dependent activity have also been reported
219 for amphibians, such as dermaseptin PD-3-7, which was isolated from the frog, *Pachymedusa*
220 *dacnicolor* [122]. In aqueous solution, the peptide showed an inherent propensity to adopt an extended
221 conformation and self-assemble into amyloid fibrils in a reversible pH-controlled manner [37]. At
222 low pH, dermaseptin PD-3-7 existed as amyloid-like β -sheet aggregates but at higher pH underwent
223 morphological changes, which led to the formation of metastable amorphous aggregates in a manner
224 that appeared to be mediated by deprotonation of the aspartic acid residues (pKa 3.9) and C-terminal
225 carboxyl groups possessed by the peptide. These amorphous aggregates induced damage to cells of
226 the insect, *Spodoptera frugiperda*, by an unidentified mechanism but showed no activity against *E. coli*
227 and *Bacillus subtilis* [37]. Based on these observations, it was suggested that amyloid formation by
228 dermaseptin PD-3-7 may act as a storage facility for the peptide similar to the depository function
229 proposed for the amyloidogenesis of pituitary peptide hormones [123]. Triggered by an increase in
230 pH, this storage facility would release a pre-formed, cytotoxic agent that contributed to the natural
231 defence strategy of the host amphibian [37]. However, it is worthy of note that the peptide was only
232 tested for activity against a small number of bacteria [37] and it is generally accepted that AMPs are
233 promiscuous in their antimicrobial mechanisms [29,124] with amyloid-mediated antibacterial
234 mechanisms increasingly being reported [23,125]. Interestingly, more recent studies on dermaseptin
235 PD-3-7 have shown that stereochemical modification of the peptide's second residue to form the
236 diastereomer [d-Leu2] strongly influenced the pH-triggered, morphological changes involved in
237 amyloid formation by dermaseptin PD-3-7, inducing a fundamental change in its superstructural
238 organization that was related to differences between the conformational propensities of these epimers
239 [126]. It was proposed by these latter authors that epimers of PD-3-7 may play a role as anionic AMPs,
240 or defence molecules, in the innate immune system of *P. dacnicolor* [126] and a similar proposal has
241 been made for the production of epimeric AMPs by other frogs and toads [9].
242

243 2.3. Humans and other mammals

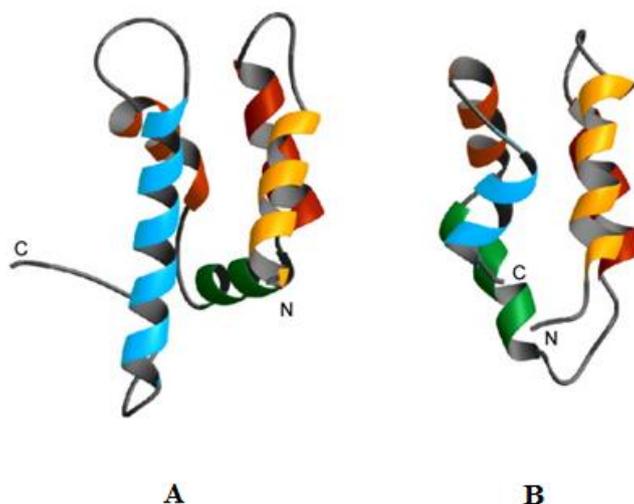
244 Psoriasin (S100A7) is a human, cysteine stabilized α -helical protein [127] of the S100 family of
245 signalling proteins [128-130] that is known to have a role in the antimicrobial defence of the skin,
246 including serving as a multifunctional modulator of neutrophil activation [131,132]. The protein was
247 investigated for antimicrobial activity and shown to kill *E. coli* using pH independent mechanisms
248 that were primarily due to the depletion of Zn^{2+} [38,39] and more recently, the protein was identified
249 in vaginal fluid, appearing to help protect the female genital tract from infection [40]. Psoriasin was
250 also found to kill *Bacillus megaterium* but via the use of two different modes of action, which involved
251 Zn^{2+} depletion at neutral pH but membrane pore formation and oligomerisation of the protein at
252 low pH. This pore forming mechanism was not further investigated but evidence suggested that it
253 was likely to show some similarities to a barrel-stave pore type mode of action [38,39]. It is interesting
254 to note that psoriasin exhibits a number of structural and functional similarities to amoebapore A
255 (Figure 2) [39,133], which is a pH dependent antimicrobial protein from the protozoa, *Entamoeba*
256 *histolytica* that is discussed below [85]. In particular, psoriasin possesses a histidine residue in its C-
257 terminal region [127] similar to amoebapore A [86] and based on these similarities, it can be
258 speculated that the enhanced action of psoriasin against *B. megaterium* at low pH may involve a
259 histidine mediated increased ability for pore formation and oligomerisation. β -microseminoprotein
260 (MSP), also named as PSP-94, is a human protein that is believed to have a protective role in prostate
261 carcinogenesis due to its ability to suppress the growth of tumours although more recent studies have
262 suggested that MSP may protect against prostate cancer by inhibiting fungal infection in this genital
263 region [134,135]. This suggestion was primarily based on recent work, which showed that the acid
264 conditions of the vagina promoted the ability of MSP in post coital seminal plasma to kill *Candidia*
265 *albicans*. This antifungal activity appeared to involve lysis of the organism's membranes and to be
266 mediated by a C-terminal fragment of MSP, which included a glutamic acid residue involved in the

267 ability of the protein to form coordinate bonds with Ca²⁺. It appeared that MSP coordination of Ca²⁺
268 at neutral pH inhibited the antifungal activity of the protein but at low pH, electrostatic interaction
269 between the ion and the C-terminal glutamic acid of MSP (pKa 4.1) decreased, facilitating the ability
270 of the protein to kill *C. albicans*. Porcine MSP appeared to use a similar pH dependent antifungal
271 mechanism, suggesting that it may be a widespread innate immune factor active against *C. albicans*
272 and possibly helping to explain the low sexual transmission rate of vulvovaginal candidiasis in
273 humans [41]. *C. albicans* was also reported to be susceptible to the pH dependent activity of LL-37
274 and its derivatives, KS-30, and RK-31, along with CRAMP [42], which is a murine homologue of LL-
275 37 [136]. It appeared that KS-30, and RK-31 were produced by the proteolytic cleavage of LL-37 in the
276 low pH environment of human sweat and that these pH conditions enhanced the ability all the
277 peptides tested to kill *C. albicans* via permeabilisation of the fungal membrane. The use of animal
278 models showed that although LL-37 and its derivatives were induced in murine skin in response to
279 *C. albicans* infection, this induction did not confer subcutaneous resistance to the organism [42]. Based
280 on these results, it was suggested that these peptides may be of primary importance in forming a
281 barrier against fungal infections on the skin surface [42], given that the dysregulated production of
282 LL-37 and its derivatives has been strongly associated with skin disease due to fungi and other
283 microbes [137-139]. Hepcidin, (hep-25) is a human β -sheet hormone that has a well-established role
284 in iron homeostasis [140] and has been shown to exhibit pH dependent antimicrobial activity [43]. In
285 particular, acid conditions been shown to enhance the activity of hep-25 and several of its isoforms,
286 such as hep-20, against the fungal pathogen, *Candida glabrata* [44], along with a range of Gram-
287 negative bacteria, such as *Pseudomonas aeruginosa*, and Gram-positive bacteria, including
288 *Enterococcus faecium* [45,46]. Studies on *E. coli* suggested that the antibacterial action of these
289 peptides involved membranolytic mechanisms which were enhanced under acid conditions due to
290 the presence of histidine residues within their primary structure [47]. Based on these observations, it
291 was suggested that with acidic pH, the increase in positive charge of histidine residues in hep-25 and
292 hep-20 would promote their ability to target and lyse bacterial membranes, resulting in the death of
293 the host organism [43]. Hep-20 has also been shown to be effective against drug resistant *C. glabrata*
294 under the low pH and physiological conditions associated with the vagina, which led to the
295 suggestion that the peptide may form the basis of novel therapeutics for the control of vaginal
296 infections due to the organism [48]. Another group of human AMPs rich in histidine residues are the
297 histatins (hst), which are salivary peptides with antiviral, antibacterial and antifungal activity [141-
298 146] and protecting the oral cavity from fungal pathogens appears to be to the primary role of these
299 peptides [147,148]. The pH of the oral cavity is mainly governed by that of saliva, which has a range
300 that generally varies between pH 5 and pH 8 [149,150] although significantly lower pH values can
301 occur on the surface of teeth due to the metabolic activities of cariogenic microorganisms [151], which
302 is able to promote the growth of fungi and form mixed species biofilms [152,153]. Low pH appears
303 able to enhance the antifungal activity of hst-1, hst-3 [154] and hst-5 [49], which, for example,
304 enhances the positive charge carried by hst-5 and facilitates its translocation into fungal cells to attack
305 intracellular targets [49]. However, a detailed description of the antifungal activity of these peptides
306 is lacking and a number of mechanisms have been proposed [141,143-146], such as complex formation
307 with iron to interfere with the cellular metabolism of the metal in fungi, such as *C. albicans* [155].
308 Studies on human lactoferrin, which is a multifunctional iron-binding protein [156] found that low
309 pH enhanced the ability of sub-lethal levels of the protein to kill *C. albicans*, through multiple
310 mechanisms, including dissipation of the proton motive force (PMF) across the CM of the organism
311 [51]. This pH effect was attributed to increased electrostatic interactions between the peptide and
312 anionic components of the *C. albicans* membrane, thereby enhancing the ability of lactoferrin to
313 partition into fungal CM and generate lesions associated with PMF dissipation and the peptide's
314 antifungal action [51]. Sub lethal levels of lactoferrin have also been shown to be effective against
315 biofilms of *P. aeruginosa* [157] and the protein was found able to synergise the activity of other
316 antimicrobials against biofilms formed from *P. aeruginosa* and methicillin resistant *Staphylococcus*
317 *aeus* (MRSA) [158,159]. It was proposed that this anti-biofilm activity was due to the iron-binding

318 properties of the protein [157] but this does not appear to be the only mechanism involved in this
 319 activity [160] and acidic pH is strongly associated with these sessile microbial communities [161].
 320 Lactoferricin B, which is a potent AMP derived from bovine lactoferrin [162], has also been shown to
 321 possess pH dependent antimicrobial activity, killing bacteria and *C. albicans* at low pH via
 322 mechanisms that appeared to involve membranolysis [60,61]. Low pH was also found to enhance the
 323 activity of platelet microbiocidal proteins, which were isolated from leporine platelets and showed
 324 activity against *Staphylococcus aureus*, *E. coli*, and *C. albicans* [57]. These results strongly supported the
 325 mounting evidence that platelets serve important multiple roles in host defence against infection,
 326 including the localized release of AMPs and other antimicrobial factors in response to microbial
 327 colonization and other stimuli [163]. The pH dependent antimicrobial mechanisms of platelet
 328 microbiocidal proteins were not further characterised but other studies showed that the antifungal
 329 and antibacterial activity of a number of these AMPs involved dissipation of the PMF across the
 330 cytoplasmic membrane of target organisms, which was able to synergise the activity of conventional
 331 antibiotics [164]. These results parallel those described above for human lactoferrin, allowing the
 332 speculation that low pH enhances the ability of some leporine platelet microbiocidal proteins to
 333 interact with membranes of *C. albicans* and generate lesions associated with PMF dissipation and the
 334 peptide's antifungal action. Other leporine AMPs shown to possess pH dependent activity are the
 335 defensins, NP1 and NP2, which were found to permeabilise the outer membrane of *P. aeruginosa* most
 336 efficiently at low pH although these peptides were ineffective against the organism under these pH
 337 conditions. These AMPs are present in leporine macrophages and it was suggested that this pH
 338 dependent membranolytic activity may synergise the antibacterial action of other defence molecules
 339 under the acid conditions associated with phagocytosis [58,59].

340

341 **Figure 2.** Similarities between the structures of psoriasin and amoebapore A



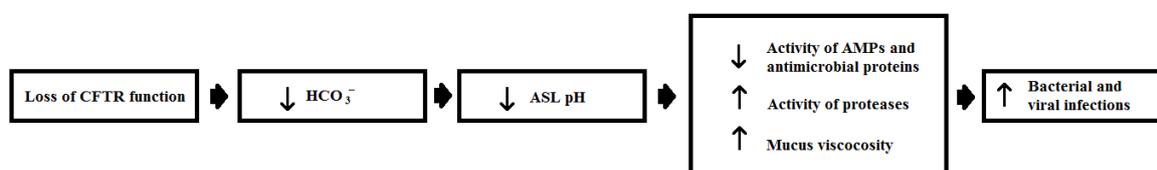
342

343 Figure 2 was revised from [39] and shows human psoriasin (A) and amoebapore A from the protozoa,
 344 *Entamoeba histolytica* (B). It can be clearly seen that these peptides show structural similarities and
 345 both have been shown to possess pH dependent mechanisms of antimicrobial activity that is
 346 enhanced by acid conditions [38-40,83-85,87]. In particular, psoriasin possesses a histidine residue in
 347 its C-terminal region [127] similarly to amoebapore A [86] and based on these similarities, it can be
 348 speculated that the enhanced antibacterial action of psoriasin at low pH may involve a histidine
 349 mediated increased ability for pore formation and oligomerisation.

350 A particularly important case of human AMPs with activity that can be influenced by pH is that
351 found in the airway surface liquid (ASL) of individuals with cystic fibrosis (CF) [165,166], which is a
352 lethal genetic disorder characterized by viscous mucus and bacterial colonization of the airways
353 [167]. In the mammalian respiratory system, the ASL represents a first line of pulmonary defence by
354 forming the interface between the environment and the host organism and helping to protect against
355 the action of inhaled and aspirated bacteria by producing a variety of antimicrobial molecules
356 [168,169]. These ASL molecules include AMPs, such as LL-37, HNP-1, HBD-1 and lactoferrin, along
357 with antimicrobial proteins, such as lysozyme, surfactant protein A and surfactant protein D
358 [165,166]. Many of these antimicrobial molecules also contribute to the pulmonary innate immune
359 system by adorning lattices of extracellular DNA, chromatin, enzymes and other proteins to form
360 neutrophil extracellular traps (NETs). These DNA complexes are released in response to the presence
361 of microbial pathogens and provide a mechanism for the localised concentration of effector
362 molecules. NETs have been reported able to eradicate microbial pathogens using a variety of
363 mechanisms, including the action of antimicrobial attachments and proteolytic degradation, as well
364 as neutralizing their activity by forming a physical barrier that prevents the dissemination of these
365 pathogens [166,170-172]. More recent studies have shown that the Human Short Palate Lung Nasal
366 Epithelial Clone 1 (SPLUNC1), which is a protein expressed in the upper airways of the lung, plays
367 multiple roles in pulmonary innate immunity. These roles include: the direct inhibition of bacterial
368 growth, the prevention of microbial biofilm formation and the regulation of other AMPs and
369 antimicrobial proteins, such as LL-37, HBD-2 and lysozyme [173-176]. However, in CF, mutations in
370 the cystic fibrosis transmembrane conductance regulator (CFTR) gene leads to reduced HCO_3^-
371 secretion and produces an abnormally acidic pH in ASL [177,178], which studies on humans and
372 animal models have suggested can negatively affect the efficacy of ASL antimicrobial molecules and
373 predispose CF airways to microbial infection (Figure 3) [165,179]. For example, recent studies on a
374 porcine CF model showed that the low pH of the ASL inhibited the activity of LL-37, lactoferrin and
375 other AMPs when directed against *S. aureus* and *P. aeruginosa* [180,181], which are known, major CF
376 pathogens [182,183]. These acid conditions also reduced the ability of ASL AMPs to synergize their
377 activities when in combination with each other and with antimicrobial proteins, such as lysozyme
378 [180,181]. It has been further proposed that conditions of low pH in CF airways could reduce the
379 efficacy of AMPs and antimicrobial proteins that adorn NETs [170] along with the antimicrobial and
380 other biological activities of SPLUNC1 [179,184]. The mechanisms by which the low pH impairs the
381 activity of AMPs and antimicrobial proteins in CF airways are currently unclear but it has been
382 suggested that these mechanisms include a variety of contributions. For example, it has been
383 proposed that low pH in CF airways may mediate the degradation of AMPs *via* the activation of host
384 proteases, such as cysteine cathepsins, and microbial enzymes, such as aureolysin of *S. aureus* and
385 elastase of *P. aeruginosa*, and the immobilization of AMPs through binding to mucins, which are large,
386 anionic glycoproteins and the primary component of mucus. It has been further proposed that low
387 pH in CF airways may induce conformational changes in AMPs that reduce the ability of these
388 peptides to bind microbial membranes and cell wall components, such as lipid II
389 [165,166,170,180,181,185]. In addition to reducing the activity of AMPs, low pH appears able to
390 negatively impact on other defence factors of the ASL; for example, by increasing the rheological
391 properties of secreted mucins, decreasing ciliary beat frequency, impairing phagocyte function and
392 depleting ASL volume [165,166,179,185]. Based on these observations, it has been proposed that
393 connections between the loss of CFTR, reduced ASL pH, and impaired CF host defense function
394 could provide a paradigm for the identification of new therapeutic targets and strategies to reduce
395 the morbidity associated with CF lung disease [165].

396

397

398 **Figure 3.** The pH of airway surface liquid and the pathogenesis of cystic fibrosis

399

400 Figure 3 was revised from [165] and shows a scheme for how changes in ASL pH may influence the
 401 pathogenesis of CF. In CF, the loss of CFTR function results in decreased HCO_3^- conductance across airway
 402 epithelial cells and leads to low pH in the ASL. Under these pH conditions, ASL AMPs, such as LL-37,
 403 HNP-1, HBD-1 and lactoferrin, and antimicrobial proteins, such as lysozyme, surfactant protein A and
 404 surfactant protein D, have reduced activity. Lower pH also leads to the increased viscosity of mucins,
 405 decreased ciliary beat frequency, impaired phagocyte function and depleted ASL volume. These effects
 406 lead to a decrease in the antimicrobial efficacy of the ASL and subsequently contribute to increased
 407 respiratory infections in the CF airway, caused by both viral and bacterial pathogens [165,166,179,185].

408

409 A number of anionic AMPs with pH dependent activity have been identified in humans,
 410 including DCD-1(L), which is proteolytically cleaved from dermcidin, which is also an anionic
 411 peptide and found in human sweat [9,52,186,187]. DCD-1(L) is characterised by its broad range
 412 antimicrobial activity, killing fungi, such as *C. albicans*, as well as Gram-positive bacteria, including
 413 MRSA, Gram-negative bacteria, such as *Salmonella typhimurium*, and acid fast bacteria including
 414 rifampin- and isoniazid-resistant *Mycobacterium tuberculosis* [188-193]. The peptide appears to exhibit
 415 pH dependent antibacterial action [188] whereby low pH induces the peptide to adopt α -helical
 416 structure on the bacterial membrane surface, leading to ion channel formation *via* Zn^{2+} stabilized
 417 DCD-1(L) oligomers and death of the host organism through membrane disruption [52,53]. A more
 418 recent study gives general support to this model and suggested that under acid conditions, the
 419 negative charge on DCD-1(L) becomes neutral, which facilitates membrane partitioning and Zn^{2+}
 420 dependent membrane channel formation *via* either a barrel-stave pore or a toroidal pore type
 421 mechanism [54]. Another example of AAMPs with pH-dependent activity are kappacin A and
 422 kappacin B, which are classed as food peptides and appear to be cleaved from κ -casein in bovine milk
 423 by digestion with chymosin in the human stomach [194]. These peptides exhibit potent activity
 424 against a range of Gram-positive and Gram-negative bacteria, including *Streptococcus mutans*,
 425 *Porphyromonas gingivalis* and *Actinomyces lundii* [55,56], which make a major contribution to
 426 supragingival dental plaques [195,196]. Characterisation studies showed that the antibacterial
 427 mechanisms used by kappacins involved the pH-dependent lysis of microbial membranes, which
 428 was enhanced by acid conditions [56]. The active region of kappacins included a phosphorylated
 429 serine residue that was essential for antibacterial activity [56,197] and interestingly, these peptides
 430 showed significantly different pH optima for this activity that resulted from a single residue
 431 difference in the sequence of their active regions [55,197]. Kappacin A showed the highest
 432 antibacterial activity of the two peptides and possessed an aspartic acid residue in its active region,
 433 which was replaced by an alanine residue in the corresponding location of kappacin B [55,197,198].
 434 The functional significance of this difference in sequences is not known although it has been proposed
 435 that the additional negative charge possessed by kappacin A may enhance its ability to bind metal
 436 ions [9]. It had been demonstrated that the presence of Ca^{2+} and Zn^{2+} ions enhanced the antibacterial
 437 activity of kappacins and it was suggested that these ions may form a cationic salt bridge between
 438 kappacins and anionic components of the bacterial membrane, thereby facilitating membrane
 439 binding and antibacterial action [197]. It has been proposed that the membrane interactive
 440 conformation of these peptides may be a proline-kinked amphiphilic α -helix but conformational

441 changes observed in the peptide in the presence of membrane mimic could not be clearly assigned to
442 any particular secondary structures and the structure of kappacins remains unclear [9,56,197,198].
443

444 2.4. Marine invertebrates

445 Myticin C (myt C) exists as a number of isoforms in the bivalve mollusc, *Mytilus galloprovincialis*
446 [199-201] and has been shown to possess activity against fish viruses, including Hemorrhagic
447 Septicaemia virus and Infectious Pancreatic Necrosis virus [202]. Several studies were conducted on
448 the antimicrobial activity of a reduced form of myt C, (myt Cc, [62,63]) based on the proposal that the
449 endogenous reduction of cysteine-stabilized AMPs to produce peptides with higher levels of
450 antimicrobial activity may form part of some innate immune systems [116,117]. It was found that the
451 antiviral activity of myt C, myt Cc and derivatives of both these peptides was enhanced by low pH.
452 These studies also showed that only under acid conditions did these various peptides possess activity
453 against Gram-positive bacteria and Gram-negative bacteria. Structure function studies on *E. coli*
454 suggested that the antimicrobial action of myt C and its variants was membranolytic and involved
455 low pH mediated increases in their levels of α -helicity and β -hairpin elements within their molecular
456 architecture [62,63]. In addition to their pH dependent antimicrobial activity, both myt C and myt Cc
457 possessed chemotactic activity and appeared to be the first chemokine/cytokine-like molecules
458 identified in bivalves [63,202]. These results added to the increasing evidence that AMPs can serve as
459 cytokines [29,203] and interestingly, it is also becoming clear that these latter peptides are able to
460 exhibit antimicrobial activity [204,205] that is enhanced by low pH [206,207]. Molluscs are also a
461 source of histidine containing AMPs with pH dependent activity, such as the peptide, KPS-1, which
462 was isolated from, *Atrina pectinata*, and under acid conditions, inhibited the growth of a range of
463 Gram-negative bacteria, including *P. Aeruginosa*, *S. typhimurium* and *Enterobacter sakazakii* [64]. Ci-
464 PAP-A22 and Ci-MAM-A24 are representative peptides of the Ci-PAP and Ci-MAM families of
465 AMPs from the solitary tunicate (Sea squirt), *Ciona intestinalis* [65,66], which appear to be produced
466 in haemocytes and granulocytes of the organism. [208,209]. Both peptides were found to be
467 predominantly α -helical and to possess antimicrobial activity [68,209,210] that appeared to have a
468 pH dependency with optima that varied according to the target microbes [65-68]. In the case of Ci-
469 PAP-A22, the activity of the peptide against fungi, Gram-negative and Gram-positive bacteria was
470 enhanced by neutral pH except for *B. megaterium* which was more efficiently killed by Ci-PAP-A22
471 at acid pH [65]. In contrast, Ci-MAM-A24 showed enhanced activity against *B. megaterium*, *B. subtilis*,
472 *E. coli* and *P. aeruginosa* at low pH but neutral pH optima for action towards *S. aureus*, *Staphylococcus*
473 *epidermis*, *Serratia marsecens* and *Klebsiella pneumoniae*. The peptide was also found to exhibit pH
474 independent antimicrobial activity, killing comparable levels of *Yersinia enterocolitica* and fungi under
475 low and neutral conditions of pH [66]. The antimicrobial activity of both Ci-PAP-A22 and Ci-MAM-
476 A24 appeared to involve a membranolytic mechanism that had characteristics consistent with a
477 'carpet' or 'toroidal pore, type model (Table 1). To help explain the differences in pH dependent
478 antimicrobial activity shown by these peptides it was suggested that histidine mediated variation in
479 their positive charge may facilitate optimal membrane interaction on a species- specific basis [65,66]
480 and of course varying lipid composition will mean differing bacterial systems exhibit different
481 changes to key parameters such as lipid packing at varying pH. Interestingly, Ci-MAM-A24 was
482 found to be more potent than Ci-PAP-A22 and appears to be the first AMP reported to kill an intra-
483 amoebic pathogen [67]. It was demonstrate that Ci-MAM-A24 was able to kill *Legionella pneumophila*,
484 which is a Gram-negative parasite responsible for Legionnaire's disease [211], whilst the organism
485 was replicating intracellularly in *Acanthamoeba castellanii* [67]. It is well established that *A. castellanii*
486 acts as a vector for this bacterium [212,213], which efficiently replicates in the acidic environment of
487 host amoebal phagosomes [214,215]. Ci-MAM-A24 was also able to kill Mycobacteria, in murine
488 macrophages [210] and these acid fast bacteria are known to replicate in the acidic compartments of
489 these host cells [216]. Given the pH dependent antimicrobial activity of the peptide, it is tempting to

490 speculate that the ability of Ci-MAM-A24 to kill these various bacterial parasites was potentiated by
491 the low pH of their host cell environments. Clavaspirin, clavansins and styelins were isolated from
492 another solitary tunicate, *Styela clava* and these pH dependent AMPs were found to be rich in both
493 histidine and phenylalanine residues [72,73]. In general, it was found that clavaspirin and clavansins
494 possessed pH dependent antibacterial and antifungal activity [69-71,80] with low pH enhancing the
495 ability of these AMPs to adopt α -helical structure and permeabilize the membranes of these
496 organisms [72,73]. It appeared that the protonation of histidine residues under low pH conditions
497 promoted the ability of these AMPs to target microbial membranes whilst the presence of their
498 glycine and phenylalanine residues provided them with the conformational flexibility and structural
499 hydrophobicity to facilitate bilayer partitioning [74-79]. Styelins, which are rich in phenylalanine
500 residues, were found to show activity against both human bacterial pathogens and marine bacteria,
501 such as *Psychrobacter immobilis* and *Planococcus citreus*, [72,217]. The best characterised of these AMPs
502 is styelin D, which possesses α -helical structure and is highly unusual in that it contains twelve post-
503 translationally modified residues [81]. For example, the peptide contained multiple
504 bromotryptophan residues, which are found in the AMPs of other marine organisms [218-222] and
505 play an important role in the life of sea sponges and lower marine invertebrates [223]. Styelin D's
506 post-translationally modified residues enhanced the peptide's membranolytic action at low pH but
507 only against Gram-positive bacteria. It was suggested that a role for these extensive modifications
508 may be in preserving activity against certain organisms under the acid conditions found in
509 haemocytes of *S. clava* where the styelins are active [81].

510
511

512 2.5. Terrestrial invertebrates

513 Hebraein is produced by the tick, *Amblyomma hebraeum* [224] and showed acid pH optima for its
514 activity against *E. coli*, *S. aureus* and the fungus, *C. glabrata* [82], a major cause of vulvovaginal
515 candidiasis in diabetics [225]. The peptide possessed an α -helical structure except for a short C-
516 terminal extension containing multiple histidine residues, which appeared to be required for activity
517 against these organisms [82]. Based on these observations, it was suggested that the acidic pH
518 induced in the physiological environment when a tick blood-feeds would increase the cationicity of
519 hebraein and thereby its membrane interactivity and antimicrobial potency [82]. However, the
520 activity of hebraein against *S. aureus* appeared to be independent of this histidine cluster and *C.*
521 *albicans* was not susceptible to the action of the peptide suggesting that it possessed a variety of
522 antimicrobial mechanisms, which were influenced by the target organism [82]. Interestingly, these
523 latter studies showed hebraein to possess homology and structural similarities to microplusin, which
524 is a Cu^{2+} chelating peptide isolated from another arachnid, the cattle tick, *Rhipicephalus microplus*, with
525 broad range antimicrobial activity [226-228]. Studies on the Gram-positive bacterium, *Micrococcus*
526 *luteus*, and the fungus, *Cryptococcus neoformans*, suggested that Cu^{2+} chelation involving histidine
527 residues promotes the antimicrobial activity of microplusin by depriving vital cellular processes of
528 the ion, such as haeme-copper terminal oxidases that contribute to cell respiration [228-230].
529 Amoebapores are a family of cystein stabilized antimicrobial proteins with α -helical structures that
530 are found in the cytoplasmic granules of the protozoan parasite of primates, *Entamoeba histolytica*
531 [83,85,231], and interestingly there is evidence to suggest that amoeba-like peptides may have been
532 amongst the first eukaryotic AMPs to emerge [232]. Amoebapore A is the best characterized of this
533 family of proteins and has been shown to exhibit pH dependent activity against Gram-positive
534 organisms, such as *M. luteus*, and Gram-negative bacteria, including *E. coli* [83,84,87], which appears
535 to involve pore formation in the membranes of target organisms [84]. Both pore formation and the
536 antibacterial activity of the protein were enhanced by low pH, which appears to derive from an
537 increased ability of amoebapore A to self-associate and form oligomers with some similarities to a
538 barrel-stave pore type mode of action [83-85]. Elucidation of the structure of amoebapore A indicated

539 that a C-terminal histidine residue acted as a molecular switch that triggers the formation of active
540 dimers from inactive monomers, which leads to the construction oligomeric pores in target cell
541 membranes, [86]. In addition to amoebapore A, two isoforms of the protein, amoebapores, B and C,
542 are included in the amoebapore family and all three isoforms differ markedly in their primary
543 structure and spectra of antibacterial activity, synergising their combined efficacy. Amoebapores, B
544 and C are believed to have similar antibacterial mechanisms to amoebapore A and the acidic pH
545 optima of these proteins is consistent with the low pH conditions encountered in the
546 amoebic intracellular vesicles, which form their site of action [83,85,231]. More recently,
547 acanthaporin, which is another protozoan protein with pH dependent antimicrobial activity was
548 described in the *Acanthamoeba culbertsoni*. At neutral pH, acanthaporin appears to exist as an inactive
549 dimer but low pH triggers the histidine mediated production of monomers and the formation of
550 membrane pores, which promoted the activity of the peptide against a variety of bacteria [88].
551 Caenopores, also known as saposin-like proteins (SPP), are cystein stabilized helical proteins that are
552 found in the nematode, *Caenorhabditis elegans* [93,232-235] and are distantly related to amoebapores
553 with which they share structural and functional features [89,92,236,237]. Many of the genes encoding
554 SPP proteins in *C. elegans* are induced in response to microbial challenge [232] and several of their
555 gene products have been reported to exhibit pH dependent antimicrobial activity, including SSP-1
556 [89,90], SPP-3 [91], SPP-5 [92] and SPP-12 [90]. These studies showed that that low pH enhanced the
557 ability of caenopores to kill a wide range of microbes, including Gram-negative bacteria, such as *E.*
558 *coli*; Gram-positive bacteria, including *Bacillus thuringiensis*; yeasts, such as *Saccharomyces cerevisiae*;
559 and amoebae, including *Dictyostelium discoideum*. For each of these proteins, antimicrobial activity
560 appeared to be based on an ability to form pores in membranes of target organisms under acid pH
561 conditions and it was suggested that this ability was mediated by the multiple internal
562 histidine residues possessed by caenopores. Due to these residues, the positive charge of these
563 proteins is enhanced under acid conditions, increasing the potential for interaction with anionic
564 components of microbial membranes and possibly mediating pore formation, as described for
565 amoebapores [90-93,236,237]. It was observed that the pH dependent activity of these proteins would
566 appear to reflect the pH conditions at the site of their functional action, such as SPP-1 and SPP-5,
567 which are active in the acidic environment of the *C. elegans* intestine [92,93,236,237].
568

569 3. Potential applications of pH dependent antimicrobial peptides and proteins

570 In response to the growing demand for new antibiotics with novel mechanisms of action, the
571 number of AMPs and antimicrobial proteins entering clinical trials is accelerating [12] and included
572 within these antimicrobial molecules are a number that have been reviewed here (Table 2). Currently,
573 the only pH dependent anionic AMPs that appear to have been commercially developed are
574 kappacins. Based on their activity against oral pathogens [55,56], preparations including these
575 peptides and zinc have been patented [238] and are available as a dental care products [194,198]. It
576 has also been shown that these peptides exhibit increased antimicrobial activity in foods with high
577 calcium contents [194], which, taken with the history of the safe use of κ -casein, led to the proposal
578 that kappacins may be used as a preservative [239]. In the case PD-3-7, epimers of this peptide appear
579 to be the only amyloid forming amphibian anionic AMPs so far reported and have the potential to
580 progress understanding of the role of residue chirality in the formation of disease-related amyloid
581 and aid the design of amyloid-based nanomaterials [126]. The development of functional amyloids
582 as novel nanostructure materials for multiple purposes, such as drug delivery and tissue repair /
583 engineering, is a growing area of technology [240,241] and recently, techniques have been developed
584 to detect epimeric AMPs in the complex skin secretions of frogs and toads [242].

585 The most researched of the cationic AMPs reviewed here for potential medical development is
586 LL-37, which is a prospective broad range antimicrobial agent that is also able to induce wound
587 healing and angiogenesis as well as modulate apoptosis [243]. This potential is though limited in

588 some cases by the pleiotropic effects of the peptide [244]. For example, the peptide shows a variation
589 in its sensitivity to cancer types, promoting proliferation, migration, and tumorigenesis in breast,
590 lung, and prostate cancers through receptor signaling but suppresses proliferation and induces
591 apoptotic and autophagic cell death in gastric cancer, colon cancer, and T-cell leukemia [107].
592 However, wound treatment is a globally prevalent and economic burden, which makes the
593 pleiotropic ability of LL-37 to exert healing properties and combat multiple microbial pathogens an
594 attractive platform that has been used to develop potential therapeutic strategies for wound
595 treatment [245]. For example, a clinical phase I/II study conducted by Pergamum on LL-37 led to a
596 patent [246] and showed that topical application of the peptide was safe and enhanced wound
597 healing in patients with chronic venous leg ulcers and diabetic patients suffering from infected
598 wounds [245,247]. Wound infection is a major complication in diabetic patients and in particular,
599 infected foot ulcers is one of the most serious and frequent of these complications, which accounts
600 for over 50% of all lower limb amputations performed on these patients [248]. More recently, several
601 studies have developed biodegradable drug delivery system that facilitated the controlled sustained
602 release of LL-37 and other wound healing agents, such as lactate and serpin A1, from nanoparticles.
603 LL-37 and these agents acted synergistically in the treatment of full thickness excisional wounds,
604 significantly promoting wound closure, reducing bacterial contamination and enhancing anti-
605 inflammatory activity. These systems offered several advantages over therapies commonly used to
606 treat chronic wound infections, which are often limited due to factors, such as the lack of controlled
607 delivery and the depth of skin infections [249,250]. A number of LL-37 related peptides have also
608 shown the potential for therapeutic development [243,251], such as OP-145, which was developed by
609 OctoPlus, and when the peptide was included in cream formulations for nasal application, these
610 preparations were found to be efficacious in the eradication of MRSA carriage [252]. The anterior
611 nares are the main reservoir for colonization by *S. aureus* and the nasal carriage of MRSA is an
612 important risk factor for subsequent infection and transmission of this pathogen, which has led to
613 intensive efforts to identify agents able to efficiently reduce MRSA colonization [253]. The completion
614 of phase I/II clinical trials by OP-145 also showed that the peptide was safe and efficacious as a
615 treatment for chronic otitis media, or chronic bacterial middle-ear infection (Table 2) [254]. This
616 disease afflicts millions of people worldwide and is highly recalcitrant to treatment by conventional
617 antibiotics, which is now known to be primarily due to bacterial biofilms [255]. Another derivative
618 of LL-37, 60.4Ac, has also proven to be beneficial in the treatment of patients with otitis media [245]
619 and more recently the peptide showed the potential for development as a novel local therapy to treat
620 patients with burn wounds infected with multidrug-resistant bacteria, including MRSA [256]. Burn
621 wounds are one of the most common and devastating forms of trauma and the infection of these
622 wound by drug resistant bacterial pathogens is rapidly becoming a serious therapeutic challenge in
623 the care of burn patients [257].

624 Histatins and their derivatives show the potential for a wide range of therapeutic and
625 biotechnical application [141], particularly in the field of dentistry and bio-dental research [258]. For
626 example, the hst-5 derivative, JH8194, is a promising candidate to act as a surface substrate in dental
627 implants to prevent peri-implantitis and peri-implant mucositis whilst decreasing infections
628 [259,260]. A major focus in the medical development of histatins has been in the preparation of
629 formulations to treat oral diseases and infections [141]. For example, highly effective hydrogel
630 delivery systems for the topical and oral application of hst-5 have been developed for the treatment
631 of oral candidiasis [261], which is the most common opportunistic fungal infection in
632 immunocompromised populations [262]. High potential for the topical treatment of this fungal
633 condition was also demonstrated when derivatives of hst-5 were conjugated to spermidine and tested
634 on immunocompromised murine models [263]. Compared to hst-5, these conjugates exhibited a
635 higher clinical half-life, enhanced uptake into *Candida* cells, and greater candidacidal efficacies, and
636 were proposed to be viable alternatives to azole antifungals [263], which are commonly used to treat
637 oral candidiasis [262]. A compound derived from hst-5 and hs1-3, P-113 (PAC-113), developed by
638 Pacgen, was evaluated in Phase I/ II clinical studies for the treatment of both oral candidiasis and

639 gingivitis, and was found to be safe and effective in the treatment of both conditions (Table 2)[12].
640 Gingivitis is the most common form of periodontal disease, affecting up to 15% of the adult
641 populations worldwide and primarily due to *Porphyromonas gingivalis*. Untreated the condition can
642 lead to periodontitis, the chronic destruction of connective tissues, and ultimately result in loss of
643 teeth [264]. P-113 has been patented [265] and most recently, it has been shown that the candidal
644 efficacy of the peptide was greatly enhanced when it was modified by coupling to other AMPs and
645 their derivatives [144] Another histatin, hist-1, was conjugated to a silver metallopharmaceutical and
646 the conjugate was found to have wound healing properties coupled to potent activity against
647 bacteria, which included MRSA, indicating the potential for development of novel multifunctional
648 therapeutics [266]. Clavanins are attractive candidates for development as drugs against bacteria
649 associated with sepsis, which is rapidly becoming a problematic nosocomial infection [267], and
650 recently developed nanoparticle formulations of these peptides, showed high promise as a drugs
651 against polymicrobial sepsis with morphological characteristics suitable for administration via
652 injection [80]. Derivatives of clavanins have also been developed to combat biofilms formed by, *S.*
653 *mutans*, which is a major contributor to dental plaque and one of the major etiological factors involved
654 in causing caries [268]. Dental caries is one of the most prevalent, preventable infectious diseases
655 affecting humans and is recognized as the primary cause of oral pain and tooth loss [269].

656 The major medical development of the antimicrobial proteins reviewed here appears to be
657 lactoferrin, which as described above is an iron binding protein but, like LL-37, is pleiotropic and also
658 displays broad range antimicrobial activity using a number of mechanisms, which includes the
659 release of derivative AMPs *via* hydrolysis by proteases [162,270,271]. Lactoferrin and its related
660 peptides shows the potential for a number of clinical uses, ranging from wound healing and the
661 detection of bacteria to the treatment of microbial infections both alone and in combination with other
662 clinically relevant agents [272]. A full description of these medical uses is beyond the scope of this
663 review but lactoferrin and its derivatives have featured in multiple clinical trials [160,272,273] and
664 have numerous entries in a recently constructed database of bioactive peptides derived from milk
665 proteins [274]. As major examples, lactoferrin and its derivatives have been extensively investigated
666 as potential drugs for the treatment of common viral infections including the common cold, influenza,
667 viral gastroenteritis and herpes [275] whilst the inhibitory effects of these proteins and peptides
668 against the proliferation of multiple cancers, has suggested a potential role in cancer prevention [276].
669 It is well established that many AMPs and antimicrobial proteins have anticancer activity that
670 generally appears to involve mechanisms of membranolysis that are similar to those used by these
671 molecules in their action against microbes [277,278], which in some cases shows pH dependence [98],
672 as recently described [279,280]. Advanced clinical trials have shown that the administration of
673 lactoferrin has no significant side effects and that the protein has efficacy in treating iron deficiency
674 anemia in pregnant women [281], sepsis in premature neonates, which is a common and severe
675 complication in new-born infants [282] and infections due to *Helicobacter pylori*, which is causally
676 associated with gastritis and peptic ulcer diseases [283]. A major example of the medical potential of
677 lactoferrin is the development of ALX-109 by Alaxia, which is a combination of the protein and
678 hypothiocyanite for the treatment of CF [160]. This drug combination has been granted orphan drug
679 status by American and European licensing agencies and has been shown to enhance the ability of
680 conventional antibiotics to eliminate biofilms of *P. aeruginosa* growing on CF airway epithelial cells
681 [284]. Derivatives of lactoferrin, have also shown the potential for therapeutic development such as
682 hLF(1-11), which was developed by AM Pharma, and in clinical trials the peptide was safely injected
683 into neutropenic stem cell transplantation patients [160]. Neutropenia is defined as a reduction in the
684 absolute number of neutrophils in the blood circulation, predisposing individuals to severe or fatal
685 infections [285], and currently, hLF(1-11), awaits development for the prevention of bacteremia and
686 fungal infections in immunocompromised individuals (Table 2)[273]. Lactoferricin B is cleaved from
687 the N-terminal region of bovine lactoferrin under acid pH conditions and has an extremely wide
688 spectrum of antimicrobial activity against bacterial, fungal and parasite species as well as showing
689 anti-catabolic and anti-inflammatory effects [162,270,271]. Based on these abilities, this peptide has

690 featured in numerous preclinical trials and shows the potential for a variety of therapeutic purposes,
691 including the treatment of ocular infections, osteo-articular gastro-Intestinal and dermatological
692 diseases, along with applications in veterinary practice and the food industry [272]. The commercial
693 importance of lactoferrin and its derivatives is perhaps underlined by the fact that the recombinant
694 human protein has been expressed in transgenic cattle to provide the large-scale production of
695 lactoferrin for pharmaceutical use [286]. The recombinant protein has also been expressed in microbes
696 and higher plants in the search for bioreactors with the capacity for large-scale production, which,
697 led to lactoferrin expression also being used as a tool for the enhancement of plant resistance to
698 pathogens [286].

699

700 4. Discussion

701 AMPs and antimicrobial proteins with pH dependent action against microbes appear to receive
702 relatively little attention in the literature but, as this review has shown, these molecules are produced
703 by a diverse spectrum of eukaryotes, including: vertebrates, such as fish, humans, horses, cattle,
704 rabbits, guinea pigs, mice, frogs and toads, as well as invertebrates, such as ticks, parasites, worms
705 and mollusks (Table 3). Around two thirds of the molecules reviewed here are cationic AMPs and
706 antimicrobial proteins with most of those that remain possessing net negative charges [287]. It is
707 generally recognized that the incidence of anionic antimicrobial molecules is low and that, in general,
708 their occurrence appears to be a strategy to synergize the antimicrobial activity of their cationic
709 counterparts [9,288]. For example, the proteolytic processing of the sweat borne peptide, dermcidin,
710 to yield DCD-1(L), described above, also produces a number of other anionic AMPs, such as SSL-46
711 (net charge -2) and LEK-45 (net charge -2) [289]. These sweat-derived anionic AMPs are continually
712 secreted and are believed to synergize the activity of cationic AMPs in the constitutive innate defense
713 of human skin by modulating surface colonization by microbes rather than responding to injury and
714 inflammation as observed for inducible peptides, such as LL-37 [186].

715 The pH dependence of the antimicrobial molecules reviewed here was found to vary with pH
716 with some, such as E2EM-lin, exhibiting high pH optima (Table 4) whilst others, such as Ci-PAP-A22
717 and Ci-MAM-A24, exhibited optima at either neutral or acid pH depending on the target organisms
718 [65-68]. Again depending on the target microbes, several antimicrobial molecules, including the latter
719 peptides and psoriasin, showed the ability to employ both pH dependent and pH independent
720 activity [38-40,65-68]. However, most of the AMPs and antimicrobial proteins reviewed here
721 exhibited low pH optima, which is consistent with the acidic pH found at their sites of action,
722 particularly the skin [131,290]. Consistent with these observations, the major structure / function
723 relationships that promote the pH dependent activity of the antimicrobial molecules reviewed here
724 are those involving amino acid residues that become protonated under acid conditions, including
725 histidine, aspartic acid and glutamic acid residues. Under these pH conditions, the protonation of
726 these residues will have the overall effect of increasing the cationicity or decreasing the anionicity of
727 the parent molecule thereby, enhancing its ability to target and interact with negatively charged
728 components of microbial membranes. Typical examples include hebraein [224] and clavanins [74-79],
729 and in the case of Ci-PAP-A22 and Ci-MAM-A24, it appears that the histidine mediated variation in
730 the cationicity of these peptides facilitates optimal interaction with target microbial membranes on a
731 species to specific basis [65,66]. However, given the high incidence of histidine residues in the
732 antimicrobial molecules reviewed here, it is worthy of note that the possession of these residues is
733 not necessarily sufficient for a pH dependent mode of antimicrobial action. This point is well
734 illustrated by Pc-pis, from the yellow croaker, *Pseudosciaena crocea*, which includes a number of
735 histidine residues in its primary structure and displays pH independent antimicrobial activity.
736 However, the addition of a histidine residue to its sequence generated a peptide with antimicrobial
737 activity optimal at low pH and a wider spectrum of antimicrobial activity [291].

738 A second major structure / function relationship for histidine, aspartic acid and glutamic acid
739 residues in the antimicrobial action of pH dependent AMPs and proteins reviewed here is to facilitate
740 the binding of metal ions. For example, the binding of Ca²⁺ by MSP at low pH potentiates the activity
741 of the peptide by alleviating inhibitory mechanisms that are mediated by the ion [41] and metal ion
742 binding by histidine residues appears able to promote microbial death through depletion of these
743 ions for a number of antimicrobial molecules, such as histatins [141,143,155]. In contrast, the binding
744 of metal ions appears to potentiates the activity of some antimicrobial molecules reviewed here by
745 promoting their capacity to form peptide-membrane or peptide-peptide salt bridges and thereby
746 disrupt microbial membranes, as proposed for kappacins [197] and DCD-1(L) respectively [54].
747 However, the most common structure / function relationships for histidine, aspartic acid and
748 glutamic acid residues in the antimicrobial action of the molecules reviewed here is to directly
749 promote the disruption of target microbial membranes. For example, in the case of several
750 antimicrobial proteins, the protonation of histidine residues appears to be a molecular switch that
751 initiates oligomerisation and the formation of discrete channels or pores by the protein, as in the case
752 of acanthaporin [88]. In some cases though, histidine, aspartic acid and glutamic acid residues appear
753 to play multiple roles in promoting the activity of their parent antimicrobial molecules. For example,
754 the N-terminal regions of gad-1 and gad-2 include a number of sequential histidine pairs that appear
755 to be important to their ability for lipid targeting and interaction, channel formation and thereby the
756 disruption of microbial membranes at low pH [33,34,94,99,100].

757 A further major structure / function relationship involved in the mechanisms of the antimicrobial
758 molecules reviewed here is pH related conformational change in α -helical architecture, which is by
759 far the most common secondary structural element identified in these AMPs and antimicrobial
760 proteins. Indeed, it well established that histidine, glutamic acid and aspartic acid residues have a
761 strong potential for α -helical formation that is enhanced by low pH [98,292]. The pore forming
762 antimicrobial proteins reviewed here are strongly α -helical (Figure 2) and it is known that changes to
763 the levels of α -helical architecture possessed by these proteins are enhanced by low pH, which
764 promotes their pore forming mechanisms and are key to their ability to kill microbes [39,86,88,93]. A
765 full description of these conformational changes is beyond the scope of this review but as an example,
766 the protonation of C-terminal histidine residues by low pH promotes conformational changes that
767 lead to the construction of hexameric membrane pores via the formation of active dimers from
768 inactive monomers in the case of amoebapores [83,85,231] caenopores [89-93] and psoriasin [38-40].
769 The pore forming mechanism of acanthaporin, shows similarities to those of these latter proteins and
770 also results in the formation of hexameric membrane pores. However, in the case of acanthaporin,
771 the low pH mediated protonation of C-terminal histidine residues promotes conformational changes
772 that induce pore formation via the formation of active dimers from inactive monomers [88]. Strictly,
773 based on its size, lactoferrin is an antimicrobial protein, but it is often classified with AMPs due to its
774 ubiquity in body fluids and its ability to kill bacteria using membrane interactive mechanisms with
775 similarities to those of these latter peptides [29]. However, lactoferrin was first characterized as an
776 iron binding protein and sequestration of the metal was initially believed to form the basis of its
777 antibacterial mechanism although the protein is now known to use multiple iron-independent
778 mechanisms in its activity against microbes [162,293].

779 In relation to the AMPs reviewed here low pH generally increased their levels of α -helical
780 secondary structure and thereby enhanced their capacity for membrane interaction and antimicrobial
781 activity. However, alkaline conditions promoted maximal levels of α -helical structure in E2EM-lin,
782 which appeared to promote monomer association, pore formation and membrane interaction at the
783 peptide's high pH optimum (Table 4, Figure 1). In contrast, to these latter AMPs, gad-1 and gad-2
784 were found to possess minimal levels of α -helical structure under the low pH conditions that were
785 optimal for their membrane interactions and antimicrobial activity [33]. These observations would
786 seem to clearly indicate that pH dependent structural plasticity is an important factor in the
787 antimicrobial mechanisms of many of the AMPs reviewed here. This form of structural plasticity
788 would appear to be key to facilitating the appropriate balance between the amphiphilicity and

789 hydrophobicity of these peptides that is required for their membranolytic action at optimal pH, as
790 proposed for gad-1 and gad-2 [33]. Reinforcing this proposal, other amino acid residues have been
791 reported to contribute to the structural plasticity of the α -helical AMPs reviewed here including
792 glycine, phenylalanine and post-translationally modified residues. These residues appear to enhance
793 the conformational flexibility and structural hydrophobicity of tunicate clavaspilin, clavanins and
794 styelins for bilayer partitioning and antimicrobial action at their low pH optimum [72,73].

795 The antimicrobial mechanisms of several AMPs reviewed here appear to be described by models
796 of membrane interaction, including variants of the carpet, toroidal pore and barrel-stave pore
797 mechanisms. These models of membrane interaction differ fundamentally to the pore forming
798 mechanisms of the antimicrobial proteins described above and were primarily proposed to describe
799 the membrane spanning abilities of AMPs, which are generally up to 50 residues in length [23]. A
800 number of novel antimicrobial mechanisms for AMPs have also been revealed by this review, such
801 as that described for human lactoferrin, which at sub lethal levels appears to kill microbes via the pH
802 dependent dissipation of microbial PMF [51]. The microbial PMF is an emerging potential target for
803 the development of novel AMPs and antimicrobial proteins based on the fact that the temporary
804 membrane perturbations caused by their action can have a large negative impact on bacterial
805 metabolism, affecting a diverse array of cellular processes that depend upon the PMF [294-299]. This
806 review has also described novel examples of pH dependent AMPs produced by the reduction of
807 cysteine stabilized parent AMPs including myt Cc [63,202] and E2EM-lin (Table 4, Figure 1, [114,115])
808 and it has been recently shown that the free cysteines of reduced AMPs play an important role in
809 their antimicrobial activity [116,117]. Moreover, it was speculated above that the antimicrobial
810 activity of E2EM-lin may involve pore formation via self-association and interestingly, recent work
811 has suggested that free cysteine residues may play a role in the antimicrobial activity of AMPs by
812 facilitating the oligomerisation of these peptides [300]. Taken together, these reports show that AMPs
813 with pH dependent antimicrobial activity contribute to the accumulating evidence that the
814 endogenous reduction of cysteine-stabilized AMPs is a strategy used by hosts to generate novel
815 peptides that enhance the efficacy of their antimicrobial capacity [116,117].

816 A number of the AMPs and antimicrobial proteins reviewed here, along with their derivatives,
817 have been developed for multiple medical purposes, which in some cases has led to patents and the
818 successful completion of clinical trials, and include kappacins, LL-37, histatins, lactoferrin and
819 clavanins. Major examples of the application of these AMPs and proteins include the treatment of
820 multiple cancers along with viral infections, such as the common cold; bacterial infections, including
821 those associated with implants, otitis media, neutropenia and CF; and fungal infections, particularly,
822 those detrimental to oral health. These AMPs and proteins also show the potential to induce wound
823 healing, such as for diabetic patients and burn victims, and interestingly, a recent report has indicated
824 that wound healing is accelerated by an acidic environment, which promotes a range of beneficial
825 effects including increases in antimicrobial activity and the enhancement of epithelization and
826 angiogenesis [301]. In general, the therapeutic administration of the AMPs and proteins involve
827 topical application, such as the use of mouth washes, cream formulations and hydrogel delivery
828 systems. These observations raise an interesting point in that most clinical trials to date involve the
829 treatment of skin infections or the prevention of surface colonization by microbes, particularly sessile
830 forms of these organisms, which potentially, can indicate a wide variation in local pH conditions. A
831 comprehensive understanding of the effect of pH on the antimicrobial activity of the molecule under
832 development would therefore seem necessary. Nonetheless, this is not generally the case and data
833 cited in the literature in relation to the antimicrobial activity of AMPs and proteins is usually that
834 determined under neutral pH conditions [133]. These observations clearly suggest that when
835 characterizing the antimicrobial action of AMPs, the optimal pH for their action against individual
836 microbes should be determined. This point is well illustrated by recent studies, which investigated
837 the antimicrobial action of a range of synthetic AMPs and found that high pH inhibited the action of
838 these peptides against fungi and Gram-negative bacteria but the opposite pH trend was observed
839 for Gram-positive bacteria [302].

840 **5. Conclusions**

841 This review has shown that AMPs and proteins with pH dependent antimicrobial activity are
842 increasingly being reported and that progress has been made in understanding the structure /
843 function relationships and mechanisms underpinning this activity. This review has also shown that
844 there has been considerable therapeutic development of pH dependent antimicrobial molecules to
845 treat a variety of infections and other conditions. However, one of the biggest therapeutic and
846 biotechnical developments of these antimicrobial molecules has been to provide guidance to the
847 design of novel compounds with pH dependent activity against: bacteria [303,304], fungi [50,305,306]
848 and cancer cells [307,308] as well as applications involving drug [309,310] and gene delivery [311,312].
849 As a specific example, most AMPs designed to target the low pH of tumor tissue are cationic and
850 cytotoxicity to healthy tissue at physiological pH has often been an issue for these peptides
851 [98,279,280]. To address this issue, a peptide based on magainin 2 from *X. laevis* was designed to
852 possess a negative charge at neutral pH that switched to a strong positive charge at low pH for cancer
853 targeting. Designated, HE, this novel peptide, killed human renal adenocarcinoma at low pH *via*
854 membranolytic mechanisms and was nontoxic towards healthy human cells across low and neutral
855 pH conditions, making it a promising lead compound for cancer therapy [313]. As a further example,
856 chronic infections due to *P. aeruginosa* are responsible for the majority of the morbidity and mortality
857 in patients with CF and the persistence of these infections is largely due to the organism adopting a
858 biofilm mode of growth, thereby acquiring high resistance to most antibiotics [314,315]. In response,
859 the peptide, WLBU2, was designed and found able to prevent biofilm formation by *P. aeruginosa*
860 under the low pH and high salt conditions characteristic of the CF airway without negative effects
861 on human airway epithelial cells. WLBU2 was also found able to synergize the action of commonly
862 used antibiotics, such as tobramycin and meropenem, making the peptide an attractive proposition
863 to help address the critical need for novel therapeutics able to suppress chronic CF lung infections
864 [316]. Using another approach, it has also been proposed that increasing the airway pH in CF
865 individuals by activating CFTR independent HCO₃⁻ transport pathways or by inhibiting proton
866 pumps could help prevent or reduce bacterial and viral infections associated with the disease
867 [165,180,181]. Nonetheless, this review has shown that many pH dependent AMPs and antimicrobial
868 proteins have yet to be fully characterized and it is proposed that these antimicrobial molecules merit
869 far more research attention than they currently receive. Indeed, pH dependent AMPs and
870 antimicrobial proteins appear to represent an untapped source of novel biologically active agents that
871 is awaiting full exploitation and could aid fulfillment of the urgent need for alternatives to
872 conventional antibiotics, helping to avert a return to the pre-antibiotic era.

873

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877

878

879 **References**

- 880 1. Prestinaci, F.; Pezzotti, P.; Pantosti, A. Antimicrobial resistance: A global multifaceted
881 phenomenon. *Pathog. Glob. Health* **2015**, *109*, 309-318.
- 882 2. World Health Organization. *Antimicrobial resistance: Global report on surveillance 2014*.;
883 Geneva, Switzerland., 2014.
- 884 3. O'Neill, J. *Tackling drug-resistant infections globally: Final report and recommendations*.; 2016.
- 885 4. Phoenix, D.A.; Harris, F.; Dennison, S.R. *Novel antimicrobial agents and strategies*. Wiley: 2014.
- 886 5. Nobrega, F.L.; Costa, A.R.; Kluskens, L.D.; Azeredo, J. Revisiting phage therapy: New
887 applications for old resources. *Trends Microbiol* **2015**, *23*, 185-191.
- 888 6. Harris, F.; Pierpoint, L. Photodynamic therapy based on 5-aminolevulinic acid and its use as
889 an antimicrobial agent. *Med Res Rev* **2012**, *32*, 1292-1327.
- 890 7. Harris, F.; Dennison, S.R.; Phoenix, D.A. Using sound for microbial eradication - light at the
891 end of the tunnel? *FEMS Microbiol Lett* **2014**, *356*, 20-22.
- 892 8. Harris, F.; Dennison, S.R.; Phoenix, D.A. Sounding the death knell for microbes? *Trends Mol*
893 *Med.* **2014**, *20*, 363-367.
- 894 9. Harris, F.; Dennison, S.R.; Phoenix, D.A. Anionic antimicrobial peptides from eukaryotic
895 organisms. *Curr Protein Pept Sci* **2009**, *10*, 585-606.
- 896 10. Dutta, P.; Das, S. Mammalian antimicrobial peptides: Promising therapeutic targets against
897 infection and chronic inflammation. *Curr Top Med Chem* **2016**, *16*, 99-129.
- 898 11. Steinbuch, K.B.; Fridman, M. Mechanisms of resistance to membrane-disrupting antibiotics
899 in gram-positive and gram-negative bacteria. *Med ChemComm* **2016**, *7*, 86-102.
- 900 12. Fox, J.L. Antimicrobial peptides stage a comeback. *Nat Biotechnol* **2013**, *31*, 379-382.
- 901 13. Midura-Nowaczek, K.; Markowska, A. Antimicrobial peptides and their analogs: Searching
902 for new potential therapeutics. *Perspect Med Chem* **2014**, *6*, 73-80.
- 903 14. Zasloff, M. *Antimicrobial peptides: Do they have a future as therapeutics?* Birkhauser Verlag Ag,
904 Viadukstrasse 40-44, Po Box 133, Ch-4010 Basel, Switzerland: 2016; p 147-154.
- 905 15. Andersson, D.I.; Hughes, D.; Kubicek-Sutherland, J.Z. Mechanisms and consequences of
906 bacterial resistance to antimicrobial peptides. *Drug Resist Updat* **2016**, *26*, 43-57.
- 907 16. Lee, T.-H.; Hall, K.N.; Aguilar, M.-I. Antimicrobial peptide structure and mechanism of
908 action: A focus on the role of membrane structure. *Curr Top Med Chem* **2016**, *16*, 25-39.
- 909 17. Lee, J.; Lee, D.G. Antimicrobial peptides (amps) with dual mechanisms: Membrane:
910 Disruption and apoptosis. *J Microbiol Biotechnol* **2015**, *25*, 759-764.
- 911 18. Cytrynska, M.; Zdybicka-Barabas, A. Defense peptides: Recent developments. *Biomol*
912 *concepts* **2015**, *6*, 237-251.
- 913 19. Phoenix, D.A.; Dennison, S.R.; Harris, F. Cationic antimicrobial peptides. In *Antimicrobial*
914 *peptides*, Wiley-VCH Verlag GmbH & Co. KGaA: 2013; pp 39-81.
- 915 20. Cruz, J.; Ortiz, C.; Guzman, F.; Fernandez-Lafuente, R.; Torres, R. Antimicrobial peptides:
916 Promising compounds against pathogenic microorganisms. *Curr med chem* **2014**, *21*, 2299-2321.
- 917 21. Thaker, H.D.; Cankaya, A.; Scott, R.W.; Tew, G.N. Role of amphiphilicity in the design of
918 synthetic mimics of antimicrobial peptides with gram-negative activity. *ACS Med Chem Lett* **2013**, *4*,
919 481-485.

- 920 22. Xiong, M.; Lee, M.W.; Mansbach, R.A.; Song, Z.; Bao, Y.; Peek, R.M.; Yao, C.; Chen, L.-F.;
921 Ferguson, A.L.; Wong, G.C.L., *et al.* Helical antimicrobial polypeptides with radial amphiphilicity.
922 *Proc Nat Acad Sci USA* **2015**, *112*, 13155-13160.
- 923 23. Phoenix, D.A.; Dennison, S.R.; Harris, F. Models for the membrane interactions of
924 antimicrobial peptides. In *Antimicrobial peptides*, Wiley-VCH Verlag GmbH & Co. KGaA: 2013; pp
925 145-180.
- 926 24. Hirsch, J.G. Phagocytin: A bactericidal substance from polymorphonuclear leucocytes. *J exp*
927 *med* **1956**, *103*, 589-611.
- 928 25. Hirsch, J.G. Further studies on preparation and properties of phagocytin. *J exp med* **1960**, *111*,
929 323-337.
- 930 26. Kenward, M.A.; Brown, M.R.W.; Fryer, J.J. Influence of calcium or manganese on the
931 resistance to edta, polymyxin-b or cold shock, and the composition of pseudomonas-aeruginosa
932 grown in glucose-depleted or magnesium-depleted batch cultures. *J. Appl. Bacteriol.* **1979**, *47*, 489-503.
- 933 27. Selsted, M.E.; Szklarek, D.; Lehrer, R.I. Purification and antibacterial activity of antimicrobial
934 peptides of rabbit granulocytes. *Infect Immun* **1984**, *45*, 150-154.
- 935 28. Daher, K.A.; Selsted, M.E.; Lehrer, R.I. Direct inactivation of viruses by human granulocyte
936 defensins. *J Virol* **1986**, *60*, 1068-1074.
- 937 29. Phoenix, D.A.; Dennison, S.R.; Harris, F. Antimicrobial peptides: Their history, evolution,
938 and functional promiscuity. In *Antimicrobial peptides*, Wiley-VCH Verlag GmbH & Co. KGaA: 2013;
939 pp 1-37.
- 940 30. Bruhn, O.; Groetzinger, J.; Cascorbi, I.; Jung, S. Antimicrobial peptides and proteins of the
941 horse - insights into a well-armed organism. *Vet Res* **2011**, *42*.
- 942 31. Fan, L.; Sun, J.; Zhou, M.; Zhou, J.; Lao, X.; Zheng, H.; Xu, H. Dramp: A comprehensive data
943 repository of antimicrobial peptides. *Sci Rep* **2016**, *6*.
- 944 32. Wang, G.; Mishra, B.; Lau, K.; Lushnikova, T.; Golla, R.; Wang, X. Antimicrobial peptides in
945 2014. *Pharmaceuticals (Basel, Switzerland)* **2015**, *8*, 123-150.
- 946 33. McDonald, M.; Mannion, M.; Pike, D.; Lewis, K.; Flynn, A.; Brannan, A.M.; Browne, M.J.;
947 Jackman, D.; Madera, L.; Coombs, M.R.P., *et al.* Structure-function relationships in histidine-rich
948 antimicrobial peptides from atlantic cod. *Biochimic Biophys Acta* **2015**, *1848*, 1451-1461.
- 949 34. Khatami, M.H.; Bromberek, M.; Saika-Voivod, I.; Booth, V. Molecular dynamics simulations
950 of histidine-containing cod antimicrobial peptide paralogs in self-assembled bilayers. *Biochimic*
951 *Biophys Acta* **2014**, *1838*, 2778-2787.
- 952 35. Shang, D.; Sun, Y.; Wang, C.; Ma, L.; Li, J.; Wang, X. Rational design of anti-microbial
953 peptides with enhanced activity and low cytotoxicity based on the structure of the arginine/histidine-
954 rich peptide, chensinin-1. *J Appl Microbiol* **2012**, *113*, 677-685.
- 955 36. Shang, D.J.; Sun, Y.; Wang, C.; Wei, S.; Ma, L.J.; Sun, L. Membrane interaction and
956 antibacterial properties of chensinin-1, an antimicrobial peptide with atypical structural features from
957 the skin of rana chensinensis. *Appl Microbiol Biotechnol* **2012**, *96*, 1551-1560.
- 958 37. Goessler-Schoefberger, R.; Hesser, G.; Muik, M.; Wechselberger, C.; Jilek, A. An orphan
959 dermaseptin from frog skin reversibly assembles to amyloid-like aggregates in a ph-dependent
960 fashion. *FEBS J* **2009**, *276*, 5849-5859.

- 961 38. Glaser, R.; Harder, J.; Lange, H.; Bartels, J.; Christophers, E.; Schroder, J.M. Antimicrobial
962 psoriasin (s100a7) protects human skin from escherichia coli infection. *Nat Immunol* **2005**, *6*, 57-64.
- 963 39. Michalek, M.; Gelhaus, C.; Hecht, O.; Podschun, R.; Schroeder, J.M.; Leippe, M.; Groetzingler,
964 J. The human antimicrobial protein psoriasin acts by permeabilization of bacterial membranes. *Dev*
965 *Comp Immunol* **2009**, *33*, 740-746.
- 966 40. Mildner, M.; Stichenwirth, M.; Abtin, A.; Eckhart, L.; Sam, C.; Glaser, R.; Schroder, J.M.;
967 Gmeiner, R.; Mlitz, V.; Pammer, J., *et al.* Psoriasin (s100a7) is a major escherichia coli-cidal factor of
968 the female genital tract. *Mucosal Immunol.* **2010**, *3*, 602-609.
- 969 41. Edstrom Hagerwall, A.M.; Rydengard, V.; Fernlund, P.; Morgelin, M.; Baumgarten, M.; Cole,
970 A.M.; Malmsten, M.; Kragelund, B.B.; Sorensen, O.E. Beta-microseminoprotein endows post coital
971 seminal plasma with potent candidacidal activity by a calcium- and ph-dependent mechanism. *PLoS*
972 *pathogens* **2012**, *8*, e1002625.
- 973 42. Lopez-Garcia, B.; Lee, P.H.A.; Yamasaki, K.; Gallo, R.L. Anti-fungal activity of cathelicidins
974 and their potential role in candida albicans skin infection. *J Invest Dermatol* **2005**, *125*, 108-115.
- 975 43. Lombardi, L.; Maisetta, G.; Batoni, G.; Tavanti, A. Insights into the antimicrobial properties
976 of hepcidins: Advantages and drawbacks as potential therapeutic agents. *Molecules* **2015**, *20*, 6319-
977 6341.
- 978 44. Tavanti, A.; Maisetta, G.; Del Gaudio, G.; Petruzzelli, R.; Sanguinetti, M.; Batoni, G.; Senesi,
979 S. Fungicidal activity of the human peptide hepcidin 20 alone or in combination with other
980 antifungals against candida glabrata isolates. *Peptides* **2011**, *32*, 2484-2487.
- 981 45. Maisetta, G.; Petruzzelli, R.; Brancatisano, F.L.; Esin, S.; Vitali, A.; Campa, M.; Batoni, G.
982 Antimicrobial activity of human hepcidin 20 and 25 against clinically relevant bacterial strains: Effect
983 of copper and acidic pH. *Peptides* **2010**, *31*, 1995-2002.
- 984 46. Mak, P.; Siwek, M.; Pohl, J.; Dubin, A. Menstrual hemocidin hbb115–146 is an acidophilic
985 antibacterial peptide potentiating the activity of human defensins, cathelicidin and lysozyme. *Am J*
986 *Reprod Immunol* **2007**, *57*, 81-91.
- 987 47. Maisetta, G.; Vitali, A.; Scorciapino, M.A.; Rinaldi, A.C.; Petruzzelli, R.; Brancatisano, F.L.;
988 Esin, S.; Stringaro, A.; Colone, M.; Luzzi, C., *et al.* Ph-dependent disruption of escherichiacoli atcc 25922
989 and model membranes by the human antimicrobial peptides hepcidin 20 and 25. *FEBS J* **2013**, *280*,
990 2842-2854.
- 991 48. Del Gaudio, G.; Lombardi, L.; Maisetta, G.; Esin, S.; Batoni, G.; Sanguinetti, M.; Senesi, S.;
992 Tavanti, A. Antifungal activity of the non cytotoxic human peptide hepcidin 20 against fluconazole
993 resistant candida glabrata in human vaginal fluid. *Antimicrob Agents Chemother* **2013**.
- 994 49. Mochon, A.B.; Liu, H. The antimicrobial peptide histatin-5 causes a spatially restricted
995 disruption on the candida albicans surface, allowing rapid entry of the peptide into the cytoplasm.
996 *Plos Pathogens* **2008**, *4*.
- 997 50. Kacprzyk, L.; Rydengard, V.; Morgelin, M.; Davoudi, M.; Pasupuleti, M.; Malmsten, M.;
998 Schmidtchen, A. Antimicrobial activity of histidine-rich peptides is dependent on acidic conditions.
999 *Biochim Biophys Acta* **2007**, *1768*, 2667-2680.
- 1000 51. Viejo-Diaz, M.; Andres, M.T.; Fierro, J.F. Modulation of in vitro fungicidal activity of human
1001 lactoferrin against candida albicans by extracellular cation concentration and target cell metabolic
1002 activity. *Antimicrob. Agents Chemother.* **2004**, *48*, 1242-1248.

- 1003 52. Song, C.; Weichbrodt, C.; Salnikov, E.S.; Dynowski, M.; Forsberg, B.O.; Bechinger, B.;
1004 Steinem, C.; de Groot, B.L.; Zachariae, U.; Zeth, K. Crystal structure and functional mechanism of a
1005 human antimicrobial membrane channel. *Proc Natl Acad Sci USA* **2013**, *110*, 4586-4591.
- 1006 53. Paulmann, M.; Arnold, T.; Linke, D.; Özdirekcan, S.; Kopp, A.; Gutschmann, T.; Kalbacher, H.;
1007 Wanke, I.; Schuenemann, V.J.; Habeck, M., *et al.* Structure-activity analysis of the dermcidin-derived
1008 peptide dcd-11, an anionic antimicrobial peptide present in human sweat. *J Biol Chem* **2012**, *287*, 8434-
1009 8443.
- 1010 54. Becucci, L.; Valensin, D.; Innocenti, M.; Guidelli, R. Dermcidin, an anionic antimicrobial
1011 peptide: Influence of lipid charge, ph and zn²⁺ on its interaction with a biomimetic membrane. *Soft*
1012 *Matter* **2014**, *10*, 616-626.
- 1013 55. Dashper, S.G.; Liu, S.W.; Reynolds, E.C. Antimicrobial peptides and their potential as oral
1014 therapeutic agents. *Int J Pept Res Ther* **2007**, *13*, 505-516.
- 1015 56. Malkoski, M.; Dashper, S.G.; O'Brien-Simpson, N.M.; Talbo, G.H.; Macris, M.; Cross, K.J.;
1016 Reynolds, E.C. Kappacin, a novel antibacterial peptide from bovine milk. *Antimicrob Agents Chemother*
1017 **2001**, *45*, 2309-2315.
- 1018 57. Yeaman, M.R.; Tang, Y.Q.; Shen, A.J.; Bayer, A.S.; Selsted, M.E. Purification and in vitro
1019 activities of rabbit platelet microbicidal proteins. *Infect Immun* **1997**, *65*, 1023-1031.
- 1020 58. Sawyer, J.G.; Martin, N.L.; Hancock, R.E. Interaction of macrophage cationic proteins with
1021 the outer membrane of pseudomonas aeruginosa. *Infect Immun* **1988**, *56*, 693-698.
- 1022 59. Lehrer, R.I.; Lichtenstein, A.K.; Ganz, T. Defensins: Antimicrobial and cytotoxic peptides of
1023 mammalian cells. *Annu Rev Immunol* **1993**, *11*, 105-128.
- 1024 60. Bellamy, W.; Wakabayashi, H.; Takase, M.; Kawase, K.; Shimamura, S.; Tomita, M. Killing of
1025 candida-albicans by lactoferricin-b, a potent antimicrobial peptide derived from the n-terminal region
1026 of bovine lactoferrin. *Med. Microbiol. Immunol.* **1993**, *182*, 97-105.
- 1027 61. Bellamy, W.R.; Wakabayashi, H.; Takase, M.; Kawase, K.; Shimamura, S.; Tomita, M. Role of
1028 cell-binding in the antibacterial mechanism of lactoferricin b. *J Appl Bacteriol* **1993**, *75*, 478-484.
- 1029 62. Domeneghetti, S.; Franzoi, M.; Damiano, N.; Norante, R.; El Haifawy, N.M.; Mammi, S.;
1030 Marin, O.; Bellanda, M.; Venier, P. Structural and antimicrobial features of peptides related to myticin
1031 c, a special defense molecule from the mediterranean mussel mytilus galloprovincialis. *J Agric Food*
1032 *Chem* **2015**, *63*, 9251-9259.
- 1033 63. Martinez-Lopez, A.; Antonio Encinar, J.; Maria Medina-Gali, R.; Balseiro, P.; Garcia-Valtanen,
1034 P.; Figueras, A.; Novoa, B.; Estepa, A. Ph-dependent solution structure and activity of a reduced form
1035 of the host-defense peptide myticin c (myt c) from the mussel mytilus galloprovincialis. *Marine Drugs*
1036 **2013**, *11*, 2328-2346.
- 1037 64. Yoo, S.; Kim, J.-Y.; Park, S.-C.; Choi, D.Y.; Seo, C.H.; Hahm, K.-S.; Park, Y. Effect of acidic ph
1038 on antibacterial action of peptide isolated from korean pen shell (*atrina pectinata*). *J Pept Scie* **2011**,
1039 *17*, 353-357.
- 1040 65. Fedders, H.; Leippe, M. A reverse search for antimicrobial peptides in *ciona intestinalis*:
1041 Identification of a gene family expressed in hemocytes and evaluation of activity. *Dev Comp Immunol*
1042 **2008**, *32*, 286-298.

- 1043 66. Fedders, H.; Michalek, M.; Groetzinger, J.; Leippe, M. An exceptional salt-tolerant
1044 antimicrobial peptide derived from a novel gene family of haemocytes of the marine invertebrate
1045 ciona intestinalis. *Biochem. J.* **2008**, *416*, 65-75.
- 1046 67. Schlusshuber, M.; Humblot, V.; Casale, S.; Methivier, C.; Verdon, J.; Leippe, M.; Berjeaud,
1047 J.-M. Potent antimicrobial peptides against legionella pneumophila and its environmental host,
1048 acanthamoeba castellanii. *Appl Microbiol Biotechnol* **2015**, *99*, 4879-4891.
- 1049 68. Fedders, H.; Podschun, R.; Leippe, M. The antimicrobial peptide ci-mam-a24 is highly active
1050 against multidrug-resistant and anaerobic bacteria pathogenic for humans. *Int J Antimicrob Agents*
1051 **2010**, *36*, 264-266.
- 1052 69. Mulder, K.C.; de Lima, L.A.; Aguiar, P.S.; Carneiro, F.C.; Franco, O.L.; Dias, S.C.; Parachin,
1053 N.S. Production of a modified peptide clavanin in pichia pastoris: Cloning, expression, purification
1054 and in vitro activities. *AMB Express* **2015**, *5*, 1-8.
- 1055 70. Silva, O.N.; Fensterseifer, I.C.M.; Rodrigues, E.A.; Holanda, H.H.S.; Novaes, N.R.F.; Cunha,
1056 J.P.A. Clavanin a improves outcome of complications from different bacterial infections. *Antimicrob*
1057 *Agents Chemother* **2015**, *59*.
- 1058 71. In, I.H.; Zhao, C.; Nguyen, T.; Menzel, L.; Waring, A.J.; Lehrer, R.I.; Sherman, M.A.
1059 Clavaspilin, an antibacterial and haemolytic peptide from styela clava. *J Peptid Res* **2001**, *58*, 445-456.
- 1060 72. Lehrer, R.I.; Andrew Tincu, J.; Taylor, S.W.; Menzel, L.P.; Waring, A.J. Natural peptide
1061 antibiotics from tunicates: Structures, functions and potential uses. *Integr Comp Biol* **2003**, *43*, 313-322.
- 1062 73. Lehrer, R.I.; Lee, I.H.; Menzel, L.; Waring, A.; Zhao, C. Clavanins and styelins, alpha-helical
1063 antimicrobial peptides from the hemocytes of styela clava. *Adv exp med biol* **2001**, *484*, 71-76.
- 1064 74. Lee, I.H.; Cho, Y.; Lehrer, R.I. Effects of pH and salinity on the antimicrobial properties of
1065 clavanins. *Infect Immun* **1997**, *65*, 2898-2903.
- 1066 75. Lee, I.H.; Zhao, C.Q.; Cho, Y.; Harwig, S.S.L.; Cooper, E.L.; Lehrer, R.I. Clavanins, alpha-
1067 helical antimicrobial peptides from tunicate hemocytes. *FEBS Lett* **1997**, *400*, 158-162.
- 1068 76. van Kan, E.J.M.; Demel, R.A.; Breukink, E.; van der Bent, A.; de Kruijff, B. Clavanin
1069 permeabilizes target membranes via two distinctly different pH-dependent mechanisms. *Biochemistry*
1070 **2002**, *41*, 7529-7539.
- 1071 77. van Kan, E.J.M.; Demel, R.A.; van der Bent, A.; de Kruijff, B. The role of the abundant
1072 phenylalanines in the mode of action of the antimicrobial peptide clavanin. *Biochim Biophys Acta* **2003**,
1073 *1615*, 84-92.
- 1074 78. van Kan, E.J.M.; Ganchev, D.N.; Snel, M.M.E.; Chupin, V.; van der Bent, A.; de Kruijff, B. The
1075 peptide antibiotic clavanin a interacts strongly and specifically with lipid bilayers. *Biochemistry* **2003**,
1076 *42*, 11366-11372.
- 1077 79. van Kan, E.J.M.; van der Bent, A.; Demel, R.A.; de Kruijff, B. Membrane activity of the peptide
1078 antibiotic clavanin and the importance of its glycine residues. *Biochemistry* **2001**, *40*, 6398-6405.
- 1079 80. Saude, A.C.M.; Ombredane, A.S.; Silva, O.N.; Barbosa, J.A.R.G.; Moreno, S.E.; Guerra Araujo,
1080 A.C.; Falcao, R.; Silva, L.P.; Dias, S.C.; Franco, O.L. Clavanin bacterial sepsis control using a novel
1081 methacrylate nanocarrier. *Int J Nanomedicine* **2014**, *9*, 5055-5069.
- 1082 81. Taylor, S.W.; Craig, A.G.; Fischer, W.H.; Park, M.; Lehrer, R.I. Styelin d, an extensively
1083 modified antimicrobial peptide from ascidian hemocytes. *J Biol Chem* **2000**, *275*, 38417-38426.

- 1084 82. Lai, R.; Takeuchi, H.; Lomas, L.O.; Jonczy, J.; Rigden, D.J.; Rees, H.H.; Turner, P.C. A new
1085 type of antimicrobial protein with multiple histidines from the hard tick, *amblyomma hebraeum*.
1086 *FASEB J* **2004**.
- 1087 83. Andra, J.; Herbst, R.; Leippe, M. Amoebapores, archaic effector peptides of protozoan origin,
1088 are discharged into phagosomes and kill bacteria by permeabilizing their membranes. *Dev Comp*
1089 *Immunol* **2003**, *27*, 291-304.
- 1090 84. Bruhn, H.; Riekens, B.; Berninghausen, O.; Leippe, M. Amoebapores and nk-lysin, members
1091 of a class of structurally distinct antimicrobial and cytolytic peptides from protozoa and mammals:
1092 A comparative functional analysis. *Biochem. J.* **2003**, *375*, 737-744.
- 1093 85. Leippe, M. Pore-forming toxins from pathogenic amoebae. *Appl Microbiol Biotechnol* **2014**, *98*,
1094 4347-4353.
- 1095 86. Leippe, M.; Bruhn, H.; Hecht, O.; Grotzinger, J. Ancient weapons: The three-dimensional
1096 structure of amoebapore a. *Trends Parasitol* **2005**, *21*, 5-7.
- 1097 87. Mann, B.J.; Loftus, B.J. The molecular biology and pathogenicity of *entamoeba histolytica*. In
1098 *Pathogen genomics: Impact on human health*, Shaw, K.J., Ed. Humana Press: Totowa, NJ, 2002; pp 281-
1099 302.
- 1100 88. Michalek, M.; Sonnichsen, F.D.; Wechselberger, R.; Dingley, A.J.; Hung, C.W.; Kopp, A.;
1101 Wienk, H.; Simanski, M.; Herbst, R.; Lorenzen, I., *et al.* Structure and function of a unique pore-
1102 forming protein from a pathogenic acanthamoeba. *Nat Chem Biol* **2013**, *9*, 37-42
- 1103 89. Banyai, L.; Patthy, L. Amoebapore homologs of *caenorhabditis elegans*. *BBA Prot Struc M*
1104 **1998**, *1429*, 259-264.
- 1105 90. Hoekendorf, A.; Stanisak, M.; Leippe, M. The saposin-like protein spp-12 is an antimicrobial
1106 polypeptide in the pharyngeal neurons of *caenorhabditis elegans* and participates in defence against
1107 a natural bacterial pathogen. *Biochem. J.* **2012**, *445*, 205-212.
- 1108 91. Hoekendorf, A.; Leippe, M. Spp-3, a saposin-like protein of *caenorhabditis elegans*, displays
1109 antimicrobial and pore-forming activity and is located in the intestine and in one head neuron. *Dev*
1110 *Comp Immunol* **2012**, *38*, 181-186.
- 1111 92. Roeder, T.; Stanisak, M.; Gelhaus, C.; Bruchhaus, I.; Groetzinger, J.; Leippe, M. Caenopores
1112 are antimicrobial peptides in the nematode *caenorhabditis elegans* instrumental in nutrition and
1113 immunity. *Dev Comp Immunol* **2010**, *34*, 203-209.
- 1114 93. Dierking, K.; Yang, W.; Schulenburg, H. Antimicrobial effectors in the nematode
1115 *Caenorhabditis elegans*: an outgroup to the Arthropoda. *Philos Trans R Soc Lond B Biol Sci.* **2016**, *371*.
1116 DOI: 10.1098/rstb.2015.0299
- 1117 94. Browne, M.J.; Feng, C.Y.; Booth, V.; Rise, M.L. Characterization and expression studies of
1118 gaduscidin-1 and gaduscidin-2; paralogous antimicrobial peptide-like transcripts from atlantic cod
1119 (*gadus morhua*). *Dev Comp Immunol* **2011**, *35*, 399-408.
- 1120 95. Rise, M.L.; Hall, J.R.; Alcock, B.P.; Hori, T.S. Dynamic expression profiles of virus-responsive
1121 and putative antimicrobial "peptide-encoding transcripts during atlantic cod (*gadus morhua*)
1122 embryonic and. Early larval development. *Gene* **2012**, *509*, 232-246.
- 1123 96. Huang, Y.; He, L.; Li, G.; Zhai, N.; Jiang, H.; Chen, Y. Role of helicity of α -helical antimicrobial
1124 peptides to improve specificity. *Prot Cell* **2014**, *5*, 631-642.
- 1125 97. Burton, M.F.; Steel, P.G. The chemistry and biology of Il-37. *Natl Prod Rep* **2009**, *26*, 1572-1584.

- 1126 98. Harris, F.; Dennison, S.R.; Singh, J.; Phoenix, D.A. On the selectivity and efficacy of defense
1127 peptides with respect to cancer cells. *Med Res Rev* **2013**, *33*, 190-234.
- 1128 99. Sengupta, D.; Leontiadou, H.; Mark, A.E.; Marrink, S.-J. Toroidal pores formed by
1129 antimicrobial peptides show significant disorder. *BBA - Biomembranes* **2008**, *1778*, 2308-2317.
- 1130 100. Burkhard, B.; Christopher, A. The polymorphic nature of membrane-active peptides from
1131 biophysical and structural investigations. *Curr Prot Pept Sci* **2012**, *13*, 602-610.
- 1132 101. Shagaghi, N.; Palombo, E.A.; Clayton, A.H.A.; Bhave, M. Archetypal tryptophan-rich
1133 antimicrobial peptides: Properties and applications. *World J Microbiol Biotechnol* **2016**, *32*, 1-10.
- 1134 102. Chan, D.I.; Prenner, E.J.; Vogel, H.J. Tryptophan- and arginine-rich antimicrobial peptides:
1135 Structures and mechanisms of action. *Biochimica et Biophysica Acta (BBA) - Biomembranes* **2006**, *1758*,
1136 1184-1202.
- 1137 103. Dong, W.B.; Sun, Y.; Shang, D.J. Interactions between chensinin-1, a natural antimicrobial
1138 peptide derived from rana chensinensis, and lipopolysaccharide. *Biopolymers* **2015**, *103*, 719-726.
- 1139 104. Phoenix, D.A.; Harris, F.; Mura, M.; Dennison, S.R. The increasing role of
1140 phosphatidylethanolamine as a lipid receptor in the action of host defence peptides. *Prog lipid res*
1141 **2015**, *59*, 26-37.
- 1142 105. Dennison, S.R.; Harris, F.; Mura, M.; Morton, L.H.G.; Zvelindovsky, A.; Phoenix, D.A. A
1143 novel form of bacterial resistance to the action of eukaryotic host defense peptides, the use of a lipid
1144 receptor. *Biochemistry* **2013**, *52*, 6021-6029.
- 1145 106. Mansour, S.C.; Pena, O.M.; Hancock, R.E.W. Host defense peptides: Front-line
1146 immunomodulators. *Trends Immunol* **2016**, *35*, 443-450.
- 1147 107. Kuroda, K.; Okumura, K.; Isogai, H.; Isogai, E. The human cathelicidin antimicrobial peptide
1148 ll-37 and mimics are potential anticancer drugs. *Front Oncol* **2015**, *5*.
- 1149 108. Hancock, R.E.W.; Haney, E.F.; Gill, E.E. The immunology of host defence peptides: Beyond
1150 antimicrobial activity. *Nat. Rev. Immunol.* **2016**, *16*, 321-334.
- 1151 109. Conlon, J.M. Reflections on a systematic nomenclature for antimicrobial peptides from the
1152 skins of frogs of the family ranidae. *Peptides* **2008**, *29*, 1815-1819.
- 1153 110. Kim, H.; Lee, B.J.; Lee, M.H.; Hong, S.G.; Ryu, P.D. Mechanisms of selective antimicrobial
1154 activity of gaegurin 4. *Korean J Physiol Pharmacol* **2009**, *13*, 39-47.
- 1155 111. Kumar, V.T.V.; Holthausen, D.; Jacob, J.; George, S. Host defense peptides from asian frogs
1156 as potential clinical therapies. *Antibiotics-Basel* **2015**, *4*, 136-159.
- 1157 112. Haney, E.F.; Hunter, H.N.; Matsuzaki, K.; Vogel, H.J. Solution nmr studies of amphibian
1158 antimicrobial peptides: Linking structure to function? *BBA-Biomembranes* **2009**, *1788*, 1639-1655.
- 1159 113. Kozić, M.; Vukičević, D.; Simunić, J.; Rončević, T.; Antcheva, N.; Tossi, A.; Juretić, D.
1160 Predicting the minimal inhibitory concentration for antimicrobial peptides with rana-box domain. *J*
1161 *Chem Inf Model* **2015**, *55*, 2275-2287.
- 1162 114. Kim, H.J.; Kim, S.S.; Lee, M.H.; Lee, B.J.; Ryu, P.D. Role of c-terminal heptapeptide in pore-
1163 forming activity of antimicrobial agent, gaegurin 4. *J Pept Res* **2004**, *64*, 151-158.
- 1164 115. Won, H.-S.; Kang, S.-J.; Lee, B.-J. Action mechanism and structural requirements of the
1165 antimicrobial peptides, gaegurins. *BBA - Biomembranes* **2009**, *1788*, 1620-1629.
- 1166 116. Schroeder, B.O.; Stange, E.F.; Wehkamp, J. Waking the wimp: Redox-modulation activates
1167 human beta-defensin 1. *Gut microbes* **2011**, *2*, 262-266.

- 1168 117. Schroeder, B.O.; Wu, Z.; Nuding, S.; Groscurth, S.; Marcinowski, M.; Beisner, J.; Buchner, J.;
1169 Schaller, M.; Stange, E.F.; Wehkamp, J. Reduction of disulphide bonds unmasks potent antimicrobial
1170 activity of human beta-defensin 1. *Nature* **2011**, *469*, 419-+.
- 1171 118. Lillywhite, H.B. Water relations of tetrapod integument. *J Exp Biol* **2006**, *209*, 202-226.
- 1172 119. Dennison, S.R.; Harris, F.; Phoenix, D.A. Chapter three - langmuir-blodgett approach to
1173 investigate antimicrobial peptide-membrane interactions. In *Advances in planar lipid bilayers and*
1174 *liposomes*, Aleš, I.; Chandrashekhar, V.K., Eds. Academic Press: 2014; Vol. Volume 20, pp 83-110.
- 1175 120. Eun, S.Y.; Jang, H.K.; Han, S.K.; Ryu, P.D.; Lee, B.J.; Han, K.H.; Kim, S.J. A helix-induced
1176 oligomeric transition of gaegurin 4, an antimicrobial peptide isolated from a korean frog. *Mol Cells*
1177 **2006**, *21*, 229-236.
- 1178 121. Phoenix, D.A.; Dennison, S.R.; Harris, F. Graphical techniques to visualize the amphiphilic
1179 structures of antimicrobial peptides. In *Antimicrobial peptides*, Wiley-VCH Verlag GmbH & Co. KGaA:
1180 2013; pp 115-144.
- 1181 122. Wechselberger, C. Cloning of cdnas encoding new peptides of the dermaseptin-family.
1182 *Biochim Biophys Acta* **1998**, *1388*, 279-283.
- 1183 123. Maji, S.K.; Perrin, M.H.; Sawaya, M.R.; Jessberger, S.; Vadodaria, K.; Rissman, R.A.; Singru,
1184 P.S.; Nilsson, K.P.R.; Simon, R.; Schubert, D., *et al.* Functional amyloids as natural storage of peptide
1185 hormones in pituitary secretory granules. *Science (New York, N.Y.)* **2009**, *325*, 328-332.
- 1186 124. Franco, O.L. Peptide promiscuity: An evolutionary concept for plant defense. *FEBS Lett* **2011**,
1187 *585*, 995-1000.
- 1188 125. Harris, F.; Dennison, S.R.; Phoenix, D.A. Aberrant action of amyloidogenic host defense
1189 peptides: A new paradigm to investigate neurodegenerative disorders? *Faseb J* **2012**, *26*, 1776-1781.
- 1190 126. Gossler-Schofberger, R.; Hesser, G.; Reif, M.M.; Friedmann, J.; Duscher, B.; Toca-Herrera, J.L.;
1191 Oostenbrink, C.; Jilek, A. A stereochemical switch in the adrs model system, a candidate for a
1192 functional amyloid. *Arch. Biochem. Biophys.* **2012**, *522*, 100-106.
- 1193 127. Fritz, G.; Heizmann, C.W. 3d structures of the calcium and zinc binding s100 proteins. In
1194 *Handbook of metalloproteins*, John Wiley & Sons, Ltd: 2006.
- 1195 128. Rezvanpour, A.; Shaw, G.S. Unique s100 target protein interactions. *Gen Physiol Biophys* **2009**,
1196 *28*, F39-F46.
- 1197 129. Santamaria-Kisiel, L.; Rintala-Dempsey, A.C.; Shaw, G.S. Calcium-dependent and -
1198 independent interactions of the s100 protein family. *Biochem. J.* **2006**, *396*, 201-214.
- 1199 130. Zimmer, D.B.; Sadosky, P.W.; Weber, D.J. Molecular mechanisms of s100-target protein
1200 interactions. *Microsc. Res. Tech.* **2003**, *60*, 552-559.
- 1201 131. Brogden, N.K.; Mehalick, L.; Fischer, C.L.; Wertz, P.W.; Brogden, K.A. The emerging role of
1202 peptides and lipids as antimicrobial epidermal barriers and modulators of local inflammation. *Skin*
1203 *Pharmacol Physiol* **2012**, *25*, 167-181.
- 1204 132. Harder, J.; Schroder, J.M.; Glaser, R. The skin surface as antimicrobial barrier: Present
1205 concepts and future outlooks. *Exp. Dermatol.* **2013**, *22*, 1-5.
- 1206 133. Wiesner, J.; Vilcinskas, A. Antimicrobial peptides: The ancient arm of the human immune
1207 system. *Virulence* **2010**, *1*, 440-464.

- 1208 134. Shukeir, N.; Garde, S.; Wu, J.Z.J.; Panchal, C.; Rabbani, S.A. Prostate secretory protein of 94
1209 amino acids (psp-94) and its peptide (pck3145) as potential therapeutic modalities for prostate cancer.
1210 *Anti-Cancer Drugs* **2005**, *16*, 1045-1051.
- 1211 135. Sutcliffe, S.; De Marzo, A.M.; Sfanos, K.S.; Laurence, M. Msmb variation and prostate cancer
1212 risk: Clues towards a possible fungal etiology. *The Prostate* **2014**, *74*, 569-578.
- 1213 136. Kosciuczuk, E.M.; Lisowski, P.; Jarczak, J.; Strzalkowska, N.; Jozwik, A.; Horbanczuk, J.;
1214 Krzyzewski, J.; Zwierzchowski, L.; Bagnicka, E. Cathelicidins: Family of antimicrobial peptides. A
1215 review. *Mol Biol Rep* **2012**, *39*, 10957-10970.
- 1216 137. Marcinkiewicz, M.; Majewski, S. The role of antimicrobial peptides in chronic inflammatory
1217 skin diseases. *Postępy Dermatol Alergol* **2016**, *33*, 6-12.
- 1218 138. Reinholz, M.; Ruzicka, T.; Schaubert, J. Cathelicidin II-37: An antimicrobial peptide with a role
1219 in inflammatory skin disease. *Annf Dermatol* **2012**, *24*, 126-135.
- 1220 139. Linde, A.; Lushington, G.H.; Abello, J.; Melgarejo, T. Clinical relevance of cathelicidin in
1221 infectious disease. *J Clin Cell Immunol* **2013**, *S13:003*.
- 1222 140. Ganz, T.; Nemeth, E. Hpcidin and iron homeostasis. *Biochim biophys acta* **2012**, *1823*, 1434-
1223 1443.
- 1224 141. Melino, S.; Santone, C.; Di Nardo, P.; Sarkar, B. Histatins: Salivary peptides with copper(ii)-
1225 and zinc(ii)-binding motifs perspectives for biomedical applications. *Febs J* **2014**, *281*, 657-672.
- 1226 142. de Sousa-Pereira, P.; Amado, F.; Abrantes, J.; Ferreira, R.; Esteues, P.J.; Vitorino, R. An
1227 evolutionary perspective of mammal salivary peptide families: Cystatins, histatins, statherin and
1228 prps. *Arch Oral Biol* **2013**, *58*, 451-458.
- 1229 143. Calderón-Santiago, M.; Luque de Castro, M.D. The dual trend in histatins research. *Trends*
1230 *Anal Chem* **2009**, *28*, 1011-1018.
- 1231 144. Han, J.; Jyoti, M.A.; Song, H.-Y.; Jang, W.S. Antifungal activity and action mechanism of
1232 histatin 5-halocidin hybrid peptides against candida spp. *PLoS One* **2016**, *11*.
- 1233 145. White, M.R.; Helmerhorst, E.J.; Ligtenberg, A.; Karpel, M.; Tecle, T.; Siqueira, W.L.;
1234 Oppenheim, F.G.; Hartshorn, K.L. Multiple components contribute to ability of saliva to inhibit
1235 influenza viruses. *Oral Microbiol Immunol* **2009**, *24*, 18-24.
- 1236 146. Vukosavljevic, D.; Custodio, W.; Del Bel Cury, A.A.; Siqueira, W.L. The effect of histatin 5,
1237 adsorbed on pmma and hydroxyapatite, on candida albicans colonization. *Yeast* **2012**, *29*, 459-466.
- 1238 147. Jang, W.S.; Edgerton, M. Salivary histatins: Structure, function, and mechanisms of
1239 antifungal activity. In *Candida and candidiasis, second edition*, American Society of Microbiology: 2012.
- 1240 148. Fabian, T.K.; Hermann, P.; Beck, A.; Fejerdy, P.; Fabian, G. Salivary defense proteins: Their
1241 network and role in innate and acquired oral immunity. *Int. J. Mol. Sci.* **2012**, *13*, 4295-4320.
- 1242 149. Galgut, P.N. The relevance of ph to gingivitis and periodontitis. *J Int Acad Periodontol* **2001**, *3*,
1243 61-67.
- 1244 150. Hold, K.M.; de Boer, B.S.; Zuidema, J.; Maes, R.A.A. Saliva as an analytical tool in toxicology
1245 *International Journal of Drug Testing* **1999** *1*, 1-36.
- 1246 151. Forssten, S.D.; Björklund, M.; Ouwehand, A.C. Streptococcus mutans, caries and simulation
1247 models. *Nutrients* **2010**, *2*, 290-298.
- 1248 152. Davis, D.A. How human pathogenic fungi sense and adapt to ph: The link to virulence. *Curr*
1249 *Opin Microbiol* **2009**, *12*, 365-370.

- 1250 153. Metwalli, K.H.; Khan, S.A.; Krom, B.P.; Jabra-Rizk, M.A. Streptococcus mutans, candida
1251 albicans, and the human mouth: A sticky situation. *PLoS Pathogens* **2013**, *9*, e1003616.
- 1252 154. Xu, T.; Levitz, S.M.; Diamond, R.D.; Oppenheim, F.G. Anticandidal activity of major human
1253 salivary histatins. *Infect Immun* **1991**, *59*, 2549-2554.
- 1254 155. Puri, S.; Li, R.; Ruszaj, D.; Tati, S.; Edgerton, M. Iron binding modulates candidacidal
1255 properties of salivary histatin 5. *J Dent Res* **2015**, *94*, 201-208.
- 1256 156. Kanwar, J.R.; Roy, K.; Patel, Y.; Zhou, S.-F.; Singh, M.R.; Singh, D.; Nasir, M.; Sehgal, R.;
1257 Sehgal, A.; Singh, R.S., *et al.* Multifunctional iron bound lactoferrin and nanomedicinal approaches to
1258 enhance its bioactive functions. *Molecules* **2015**, *20*, 9703-9731.
- 1259 157. Singh, P.K.; Parsek, M.R.; Greenberg, E.P.; Welsh, M.J. A component of innate immunity
1260 prevents bacterial biofilm development. *Nature* **2002**, *417*, 552-555.
- 1261 158. Ammons, M.C.; Ward, L.S.; Fisher, S.T.; Wolcott, R.D.; James, G.A. In vitro susceptibility of
1262 established biofilms composed of a clinical wound isolate of pseudomonas aeruginosa treated with
1263 lactoferrin and xylitol. *Int J Antimicrob Agents* **2009**, *33*, 230-236.
- 1264 159. Ammons, M.C.; Ward, L.S.; James, G.A. Anti-biofilm efficacy of a lactoferrin/xylitol wound
1265 hydrogel used in combination with silver wound dressings. *Int Wound J* **2011**, *8*, 268-273.
- 1266 160. Ammons, M.C.; Copié, V. Lactoferrin: A bioinspired, anti-biofilm therapeutic. *Biofouling*
1267 **2013**, *29*, 443-455.
- 1268 161. Hurdle, J.G.; O'Neill, A.J.; Chopra, I.; Lee, R.E. Targeting bacterial membrane function: An
1269 underexploited mechanism for treating persistent infections. *Nat Rev Microbiol* **2011**, *9*, 62-75.
- 1270 162. Sinha, M.; Kaushik, S.; Kaur, P.; Sharma, S.; Singh, T.P. Antimicrobial lactoferrin peptides:
1271 The hidden players in the protective function of a multifunctional protein. *Internatl j pept* **2013**, *2013*,
1272 390230-390230.
- 1273 163. Yeaman, M.R. Platelets: At the nexus of antimicrobial defence. *Nat rev. Microbiol* **2014**, *12*, 426-
1274 437.
- 1275 164. Yeaman, M.R. The role of platelets in antimicrobial host defense. *Clin Infect Dis* **1997**, *25*, 951-
1276 968.
- 1277 165. Berkebile, A.R.; McCray, P.B. Effects of airway surface liquid ph on host defense in cystic
1278 fibrosis. *International Journal of Biochemistry & Cell Biology* **2014**, *52*, 124-129.
- 1279 166. Lecaille, F.; Lalmanach, G.; Andrault, P.M. Antimicrobial proteins and peptides in human
1280 lung diseases: A friend and foe partnership with host proteases. *Biochimie* **2016**, *122*, 151-168.
- 1281 167. Cutting, G.R. Cystic fibrosis genetics: From molecular understanding to clinical application.
1282 *Nature Reviews Genetics* **2015**, *16*, 45-56.
- 1283 168. Laubel, D.M.; Yiml, S.; Ryan, L.K.; Kisich, K.O.; Diamond, G. Antimicrobial peptides in the
1284 airway. *Curr.Top.Microbiol.Immunol.* **2006**, *306*, 153-182.
- 1285 169. Waterer, G.W. Airway defense mechanisms. *Clin. Chest Med.* **2012**, *33*, 199-+.
- 1286 170. Gray, R.D.; McCullagh, B.N.; McCray, P.B. Nets and cf lung disease: Current status and
1287 future prospects. *Antibiotics-Basel* **2015**, *4*, 62-75.
- 1288 171. Rahman, S.; Gadjeva, M. Does netosis contribute to the bacterial pathoadaptation in cystic
1289 fibrosis? *Frontiers in Immunology* **2014**, *5*, 378.
- 1290 172. Nel, J.G.; Theron, A.J.; Pool, R.; Durandt, C.; Tintinger, G.R.; Anderson, R. Neutrophil
1291 extracellular traps and their role in health and disease. *South African J Scie* **2016**, *112*, 36-44.

- 1292 173. Walton, W.G.; Ahmad, S.; Little, M.R.; Kim, C.S.K.; Tyrrell, J.; Lin, Q.; Di, Y.P.; Tarran, R.;
1293 Redinbo, M.R. Structural features essential to the antimicrobial functions of human splunc1.
1294 *Biochemistry* **2016**, *55*, 2979-2991.
- 1295 174. Liu, Y.; Bartlett, J.A.; Di, M.E.; Bomberger, J.M.; Chan, Y.R.; Gakhar, L.; Mallampalli, R.K.;
1296 McCray, P.B.; Di, Y.P. Splunc1/bpifa1 contributes to pulmonary host defense against klebsiella
1297 pneumoniae respiratory infection. *American J Pathol* **2013**, *182*, 1519-1531.
- 1298 175. Liu, H.; Zhang, X.; Wu, J.; French, S.W.; He, Z. New insights on the palate, lung, and nasal
1299 epithelium clone (plunc) proteins: Based on molecular and functional analysis of its homolog of
1300 yh1/splunc1. *Exp Mol Pathol* **2016**, *100*, 363-369.
- 1301 176. Ahmad, S.; Tyrrell, J.; Walton, W.G.; Tripathy, A.; Redinbo, M.R.; Tarran, R. Splunc1 has
1302 antimicrobial and antibiofilm activity against burkholderia cepacia complex. *Antimicrob Agents*
1303 *Chemother* **2016**.
- 1304 177. Chen, J.H.; Stoltz, D.A.; Karp, P.H.; Ernst, S.E.; Pezzulo, A.A.; Moninger, T.O.; Rector, M.V.;
1305 Reznikov, L.R.; Launspach, J.L.; Chaloner, K., *et al.* Loss of anion transport without increased sodium
1306 absorption characterizes newborn porcine cystic fibrosis airway epithelia. *Cell* **2010**, *143*, 911-923.
- 1307 178. Abou Alaiwa, M.H.; Beer, A.M.; Pezzulo, A.A.; Launspach, J.L.; Horan, R.A.; Stoltz, D.A.;
1308 Starner, T.D.; Welsh, M.J.; Zabner, J. Neonates with cystic fibrosis have a reduced nasal liquid ph; a
1309 small pilot study. *J Cys Fibros* **2014**, *13*, 373-377.
- 1310 179. Garland, A.L.; Walton, W.G.; Coakley, R.D.; Tan, C.D.; Gilmore, R.C.; Hobbs, C.A.; Tripathy,
1311 A.; Clunes, L.A.; Bencharit, S.; Stutts, M.J., *et al.* Molecular basis for ph-dependent mucosal
1312 dehydration in cystic fibrosis airways. *Proc Nat Acad Sci USA* **2013**, *110*, 15973-15978.
- 1313 180. Pezzulo, A.A.; Tang, X.X.; Hoegger, M.J.; Abou Alaiwa, M.H.; Ramachandran, S.; Moninger,
1314 T.O.; Karp, P.H.; Wohlford-Lenane, C.L.; Haagsman, H.P.; van Eijk, M., *et al.* Reduced airway surface
1315 ph impairs bacterial killing in the porcine cystic fibrosis lung. *Nature* **2012**, *487*, 109-+.
- 1316 181. Abou Alaiwa, M.H.; Reznikov, L.R.; Gansemer, N.D.; Sheets, K.A.; Horswill, A.R.; Stoltz,
1317 D.A.; Zabner, J.; Welsh, M.J. Ph modulates the activity and synergism of the airway surface liquid
1318 antimicrobials beta-defensin-3 and ll-37. *Proc Nat Acad Sci USA* **2014**, *111*, 18703-18708.
- 1319 182. Parkins, M.D.; Floto, R.A. Emerging bacterial pathogens and changing concepts of bacterial
1320 pathogenesis in cystic fibrosis. *J Cys Fibros* **2015**, *14*, 293-304.
- 1321 183. Ciofu, O.; Hansen, C.R.; Hoiby, N. Respiratory bacterial infections in cystic fibrosis. *Curr Opin*
1322 *Pulm Med* **2013**, *19*, 251-258.
- 1323 184. Garnett, J.P. Splunc1: Link between acidity and dehydration of the airway surface liquid in
1324 cf. *Thorax* **2014**, *69*, 1004.
- 1325 185. Tang, X.X.; Ostedgaard, L.S.; Hoegger, M.J.; Moninger, T.O.; Karp, P.H.; McMenimen, J.D.;
1326 Choudhury, B.; Varki, A.; Stoltz, D.A.; Welsh, M.J. Acidic ph increases airway surface liquid viscosity
1327 in cystic fibrosis. *J Clin Inv* **2016**, *126*, 879-891.
- 1328 186. Zeth, K. Dermcidin: What is its antibiotic potential? *Future Microbiol* **2013**, *8*, 817-819.
- 1329 187. Schitteck, B. The multiple facets of dermcidin in cell survival and host defense. *J Innat Immun*
1330 **2012**, *4*, 349-360.
- 1331 188. Burian, M.; Schitteck, B. The secrets of dermcidin action. *Int. J. Med. Microbiol.* **2015**, *305*, 283-
1332 286.

- 1333 189. Čipáková, I.; Gašperík, J.; Hostinová, E. Expression and purification of human antimicrobial
1334 peptide, dermcidin, in escherichia coli. *Prote Expres Purific* **2006**, *45*, 269-274.
- 1335 190. Lai, Y.P.; Peng, Y.F.; Zuo, Y.; Li, J.; Huang, J.; Wang, L.F.; Wu, Z.R. Functional and structural
1336 characterization of recombinant dermcidin-11, a human antimicrobial peptide. *Biochem Biophys Res*
1337 *Commun* **2005**, *328*, 243-250.
- 1338 191. Schitteck, B.; Hipfel, R.; Sauer, B.; Bauer, J.; Kalbacher, H.; Stevanovic, S.; Schirle, M.;
1339 Schroeder, K.; Blin, N.; Meier, F., *et al.* Dermcidin: A novel human antibiotic peptide secreted by sweat
1340 glands. *Nat immunol* **2001**, *2*, 1133-1137.
- 1341 192. Steffen, H.; Rieg, S.; Wiedemann, I.; Kalbacher, H.; Deeg, M.; Sahl, H.G.; Peschel, A.; Götz, F.;
1342 Garbe, C.; Schitteck, B. Naturally processed dermcidin-derived peptides do not permeabilize bacterial
1343 membranes and kill microorganisms irrespective of their charge. *Antimicrob. Agents Chemother.* **2006**,
1344 *50*, 2608-2620.
- 1345 193. Vuong, C.; Voyich, J.M.; Fischer, E.R.; Braughton, K.R.; Whitney, A.R.; DeLeo, F.R.; Otto, M.
1346 Polysaccharide intercellular adhesin (pia) protects staphylococcus epidermidis against major
1347 components of the human innate immune system. *Cell microbiol* **2004**, *6*, 269-275.
- 1348 194. Benkerroum, N. Antimicrobial peptides generated from milk proteins: A survey and
1349 prospects for application in the food industry. A review. *Internat J Dairy Technol* **2010**, *63*, 320-338.
- 1350 195. Huang, R.; Li, M.; Gregory, R.L. Bacterial interactions in dental biofilm. *Virulence* **2011**, *2*, 435-
1351 444.
- 1352 196. Xu, X.; He, J.; Xue, J.; Wang, Y.; Li, K.; Zhang, K.; Guo, Q.; Liu, X.; Zhou, Y.; Cheng, L., *et al.*
1353 Oral cavity contains distinct niches with dynamic microbial communities. *Environ Microbiol* **2015**, *17*,
1354 699-710.
- 1355 197. Dashper, S.G.; O'Brien-Simpson, N.M.; Cross, K.J.; Paolini, R.A.; Hoffmann, B.; Catmull, D.V.;
1356 Malkoski, M.; Reynolds, E.C. Divalent metal cations increase the activity of the antimicrobial peptide
1357 kappacin. *Antimicrob Agents Chemother* **2005**, *49*, 2322-2328.
- 1358 198. McCarthy, R.; Mills, S.; Ross, R.P.; Fitzgerald, G.F.; Stanton, C. *Bioactive peptides from casein*
1359 *and whey proteins*. Blackwell Publishing 2014; p 23-54.
- 1360 199. Costa, M.M.; Dios, S.; Alonso-Gutierrez, J.; Romero, A.; Novoa, B.; Figueras, A. Evidence of
1361 high individual diversity on myticin c in mussel (*mytilus galloprovincialis*). *Dev Comp Immunol* **2009**,
1362 *33*, 162-170.
- 1363 200. Vera, M.; Martinez, P.; Poisa-Beiro, L.; Figueras, A.; Novoa, B. Genomic organization,
1364 molecular diversification, and evolution of antimicrobial peptide myticin-c genes in the mussel
1365 (*mytilus galloprovincialis*). *PLoS One* **2011**, *6*.
- 1366 201. Pallavicini, A.; del Mar Costa, M.; Gestal, C.; Dreos, R.; Figueras, A.; Venier, P.; Novoa, B.
1367 High sequence variability of myticin transcripts in hemocytes of immune-stimulated mussels
1368 suggests ancient host-pathogen interactions. *Dev Comp Immunol* **2008**, *32*, 213-226.
- 1369 202. Balseiro, P.; Falco, A.; Romero, A.; Dios, S.; Martinez-Lopez, A.; Figueras, A.; Estepa, A.;
1370 Novoa, B. *Mytilus galloprovincialis* myticin c: A chemotactic molecule with antiviral activity and
1371 immunoregulatory properties. *PLoS One* **2011**, *6*.
- 1372 203. Brogden, K.A.; Bates, A.M.; Fischer, C.L. *Antimicrobial peptides in host defense: Functions beyond*
1373 *antimicrobial activity*. in *Antimicrobial Peptides*. Springer International publishing. Harder, J.
1374 Schroder JM 2016; p 129-146.

- 1375 204. Valdivia-Silva, J.; Medina-Tamayo, J.; Garcia-Zepeda, E.A. Chemokine-derived peptides:
1376 Novel antimicrobial and antineoplastic agents. *Int. J. Mol. Sci.* **2015**, *16*, 12958-12985.
- 1377 205. Wolf, M.; Moser, B. Antimicrobial activities of chemokines: Not just a side-effect? *Front*
1378 *immunol* **2012**, *3*, 213.
- 1379 206. Yount, N.Y.; Waring, A.J.; Gank, K.D.; Welch, W.H.; Kupferwasser, D.; Yeaman, M.R.
1380 Structural correlates of antimicrobial efficacy in il-8 and related human kinocidins. *BBA-Biomembranes*
1381 **2007**, *1768*, 598-608.
- 1382 207. Bjorstad, A.; Fu, H.M.; Karlsson, A.; Dahlgren, C.; Bylund, J. Interleukin-8-derived peptide
1383 has antibacterial activity. *Antimicrob. Agents Chemother.* **2005**, *49*, 3889-3895.
- 1384 208. Di Bella, M.A.; Fedders, H.; De Leo, G.; Leippe, M. Localization of antimicrobial peptides in
1385 the tunic of *Ciona intestinalis* (ascidiacea, tunicata) and their involvement in local inflammatory-like
1386 reactions. *Results immunol* **2011**, *1*, 70-75.
- 1387 209. Di Bella, M.A.; Fedders, H.; Leippe, M.; De Leo, G. Antimicrobial peptides in the tunic of
1388 *Ciona intestinalis* In *Worldwide research efforts in the fighting against microbial pathogens from basic research*
1389 *to technological developments*, Mendez-Vilas, A., Ed. Universal-Publishers 2013; pp 63-67.
- 1390 210. Jena, P.; Mishra, B.; Leippe, M.; Hasilik, A.; Griffiths, G.; Sonawane, A. Membrane-active
1391 antimicrobial peptides and human placental lysosomal extracts are highly active against
1392 mycobacteria. *Peptides* **2011**, *32*, 881-887.
- 1393 211. Carratala, J.; Garcia-Vidal, C. An update on legionella. *Curr Opin Infect Dis* **2010**, *23*, 152-157.
- 1394 212. Fields, B.S.; Benson, R.F.; Besser, R.E. Legionella and legionnaires' disease: 25 years of
1395 investigation. *Clin Microbiol Rev* **2002**, *15*, 506-+.
- 1396 213. Declerck, P. Biofilms: The environmental playground of legionella pneumophila. *Env*
1397 *Microbiol* **2010**, *12*, 557-566.
- 1398 214. Sturgill-Koszycki, S.; Swanson, M.S. Legionella pneumophila replication vacuoles mature
1399 into acidic, endocytic organelles. *J Exp Med* **2000**, *192*, 1261-1272.
- 1400 215. Isaac, D.T.; Isberg, R. Master manipulators: An update on legionella pneumophila icm/dot
1401 translocated substrates and their host targets. *Future Microbiol* **2014**, *9*, 343-359.
- 1402 216. Vandal, O.H.; Nathan, C.F.; Ehrt, S. Acid resistance in mycobacterium tuberculosis. *J Bacteriol*
1403 **2009**, *191*, 4714-4721.
- 1404 217. Lee, I.H.; Cho, Y.; Lehrer, R.I. Styelins, broad-spectrum antimicrobial peptides from the
1405 solitary tunicate, *styela clava*. *Comp Biochem Physiol B* **1997**, *118*, 515-521.
- 1406 218. Tasiemski, A.; Schikorski, D.; Le Marrec-Croq, F.; Pontoire-Van Camp, C.; Boidin-Wichlacz,
1407 C.; Sautiere, P.E. Hedistin: A novel antimicrobial peptide containing bromotryptophan constitutively
1408 expressed in the nk cells-like of the marine annelid, *Nereis diversicolor*. *Dev Comp Immunol* **2007**, *31*,
1409 749-762.
- 1410 219. Shinnar, A.E.; Butler, K.L.; Park, H.J. Cathelicidin family of antimicrobial peptides:
1411 Proteolytic processing and protease resistance. *Bioorg Chem* **2003**, *31*, 425-436.
- 1412 220. Nguyen, B.; Le Caer, J.-P.; Mourier, G.; Thai, R.; Lamthanh, H.; Servent, D.; Benoit, E.; Molgó,
1413 J. Characterization of a novel *Conus bandanus* conopeptide belonging to the m-superfamily
1414 containing bromotryptophan. *Marine Drugs* **2014**, *12*, 3449-3465.
- 1415 221. Buczek, O.; Bulaj, G.; Olivera, B.M. Conotoxins and the posttranslational modification of
1416 secreted gene products. *Cell Mol Life Sci CMLS* **2005**, *62*, 3067-3079.

- 1417 222. Gerwig, G.J.; Hocking, H.G.; Stöcklin, R.; Kamerling, J.P.; Boelens, R. Glycosylation of
1418 conotoxins. *Marine Drugs* **2013**, *11*, 623-642.
- 1419 223. Bittner, S.; Scherzer, R.; Harlev, E. The five bromotryptophans. *Amino Acids* **2007**, *33*, 19-42.
- 1420 224. Hajdušek, O.; Šíma, R.; Ayllón, N.; Jalovecká, M.; Perner, J.; de la Fuente, J.; Kopáček, P.
1421 Interaction of the tick immune system with transmitted pathogens. *Front Cell Infect Microbiol* **2013**, *3*,
1422 26.
- 1423 225. Nyirjesy, P.; Sobel, J.D. Genital mycotic infections in patients with diabetes. *Postgrad Med*
1424 **2013**, *125*, 33-46.
- 1425 226. Fogaça, A.C.; Lorenzini, D.M.; Kaku, L.M.; Esteves, E.; Bulet, P.; Daffre, S. Cysteine-rich
1426 antimicrobial peptides of the cattle tick boophilus microplus: Isolation, structural characterization
1427 and tissue expression profile. *Dev Comp Immunol* **2004**, *28*, 191-200.
- 1428 227. Esteves, E.; Fogaca, A.C.; Maldonado, R.; Silva, F.D.; Manso, P.P.; Pelajo-Machado, M.; Valle,
1429 D.; Daffre, S. Antimicrobial activity in the tick rhipicephalus (boophilus) microplus eggs: Cellular
1430 localization and temporal expression of microplusin during oogenesis and embryogenesis. *Dev Comp*
1431 *Immunol* **2009**, *33*, 913-919.
- 1432 228. Joazeiro, A.C.; Coutinho, M.L.; Martins, J.R.; Masuda, A.; Seixas, A.; Vaz, I.D. Antimicrobial
1433 peptides in rhipicephalus (boophilus) microplus. *Acta Sci. Vet.* **2012**, *40*, 14.
- 1434 229. Silva, F.D.; Rezende, C.A.; Rossi, D.C.; Esteves, E.; Dyszy, F.H.; Schreier, S.; Gueiros-Filho, F.;
1435 Campos, C.B.; Pires, J.R.; Daffre, S. Structure and mode of action of microplusin, a copper ii-chelating
1436 antimicrobial peptide from the cattle tick rhipicephalus (boophilus) microplus. *J biol chem* **2009**, *284*,
1437 34735-34746.
- 1438 230. Silva, F.D.; Rossi, D.C.P.; Martinez, L.R.; Frases, S.; Fonseca, F.L.; Campos, C.B.L.; Rodrigues,
1439 M.L.; Nosanchuk, J.D.; Daffre, S. Effects of microplusin, a copper-chelating antimicrobial peptide,
1440 against cryptococcus neoformans. *Fems Microbiol Lett* **2011**, *324*, 64-72.
- 1441 231. Leippe, M.; Herbst, R. Ancient weapons for attack and defense: The pore-forming
1442 polypeptides of pathogenic enteric and free-living amoeboid protozoa. *J. Eukaryot. Microbiol.* **2004**, *51*,
1443 516-521.
- 1444 232. Bogaerts, A.; Beets, I.; Schoofs, L.; Verleyen, P. Antimicrobial peptides in caenorhabditis
1445 elegans. *Isj-Invertebrate Survival Journal* **2010**, *7*, 45-52.
- 1446 233. Ewbank, J.J.; Zugasti, O. C. Elegans: Model host and tool for antimicrobial drug discovery.
1447 *Dis Mod Mech* **2011**, *4*, 300-304.
- 1448 234. Squiban, B.; Kurz, C.L. C. Elegans: An all in one model for antimicrobial drug discovery. *Curr*
1449 *drug targets* **2011**, *12*, 967-977.
- 1450 235. Zhang, R.; Hou, A. Host-microbe interactions in caenorhabditis elegans. *ISRN microbiology*
1451 **2013**, *2013*, 356451-356451.
- 1452 236. Tarr, D.E.K. Distribution and characteristics of abfs, cecropins, nemapores, and lysozymes in
1453 nematodes. *Dev Comp Immunol* **2012**, *36*, 502-520.
- 1454 237. Tarr, D.E.K. Nematode antimicrobial peptides. *Isj-Invertebrate Survival Journal* **2012**, *9*, 122-
1455 133.
- 1456 238. Reynolds, E.C.; Dashper, S.G.; O'Brien-Simpson, N.M.; Talbo, G.H.; Malkosi, M. Derived
1457 from milk protein casein; for use in dentistry. US patent 7588752 B2, 2009.

- 1458 239. Kent, R.M.; Fitzgerald, G.F.; Hill, C.; Stanton, C.; Ross, R.P. Novel approaches to improve the
1459 intrinsic microbiological safety of powdered infant milk formula. *Nutrients* **2015**, *7*, 1217-1244.
- 1460 240. Mankar, S.; Anoop, A.; Sen, S.; Maji, S.K. Nanomaterials: Amyloids reflect their brighter side.
1461 *Nano Reviews* **2011**, *2*, 10.3402/nano.v3402i3400.6032.
- 1462 241. Kim, S.; Kim, J.H.; Lee, J.S.; Park, C.B. Beta-sheet-forming, self-assembled peptide
1463 nanomaterials towards optical, energy, and healthcare applications. *Small* **2015**, *11*, 3623-3640.
- 1464 242. Pinkse, M.; Evaristo, G.; Pieterse, M.; Yu, Y.; Verhaert, P. Ms approaches to select peptides
1465 with post-translational modifications from amphibian defense secretions prior to full sequence
1466 elucidation. *EuPA Open Proteomics* **2014**, *5*, 32-40.
- 1467 243. Xhindoli, D.; Pacor, S.; Benincasa, M.; Scocchi, M.; Gennaro, R.; Tossi, A. The human
1468 cathelicidin ll-37--a pore-forming antibacterial peptide and host-cell modulator. *Biochim Biophys Acta*
1469 **2016**, *1858*, 546-566.
- 1470 244. Fabisiak, A.; Murawska, N.; Fichna, J. Ll-37: Cathelicidin-related antimicrobial peptide with
1471 pleiotropic activity. *Pharmacol Rep* **2016**, *68*, 802-808.
- 1472 245. Duplantier, A.J.; van Hoek, M.L. The human cathelicidin antimicrobial peptide ll-37 as a
1473 potential treatment for polymicrobial infected wounds. *Front immunol* **2013**, *4*, 143.
- 1474 246. Gronberg, A.; Dieterich, C.; Mahlapuu, M. New treatment of chronic ulcers. Google Patents:
1475 2015.
- 1476 247. Gronberg, A.; Mahlapuu, M.; Stahle, M.; Whately-Smith, C.; Rollman, O. Treatment with ll-
1477 37 is safe and effective in enhancing healing of hard-to-heal venous leg ulcers: A randomized,
1478 placebo-controlled clinical trial. *Wound Repair and Regeneration* **2014**, *22*, 613-621.
- 1479 248. Yazdanpanah, L.; Nasiri, M.; Adarvishi, S. Literature review on the management of diabetic
1480 foot ulcer. *World J Diab* **2015**, *6*, 37-53.
- 1481 249. Fumakia, M.; Ho, E.A. Nanoparticles encapsulated with ll37 and serpin a1 promotes wound
1482 healing and synergistically enhances antibacterial activity. *Mol pharm* **2016**, *13*, 2318-2331.
- 1483 250. Chereddy, K.K.; Her, C.H.; Comune, M.; Moia, C.; Lopes, A.; Porporato, P.E.; Vanacker, J.;
1484 Lam, M.C.; Steinstraesser, L.; Sonveaux, P., *et al.* Plga nanoparticles loaded with host defense peptide
1485 ll37 promote wound healing. *J Control Release* **2014**, *194*, 138-147.
- 1486 251. Wang, G.; Mishra, B.; Eband, R.F.; Eband, R.M. High-quality 3d structures shine light on
1487 antibacterial, anti-biofilm and antiviral activities of human cathelicidin ll-37 and its fragments. *BBA -*
1488 *Biomembranes* **2014**, *1838*, 2160-2172.
- 1489 252. Goblyos, A.; Schimmel, K.J.; Valentijn, A.R.; Fathers, L.M.; Cordfunke, R.A.; Chan, H.L.;
1490 Oostendorp, J.; Nibbering, P.H.; Drijfhout, J.W.; Hiemstra, P.S., *et al.* Development of a nose cream
1491 containing the synthetic antimicrobial peptide p60.4ac for eradication of methicillin-resistant
1492 staphylococcus aureus carriage. *J pharm sci* **2013**, *102*, 3539-3544.
- 1493 253. Abad, C.L.; Pulia, M.S.; Safdar, N. Does the nose know? An update on mrsa decolonization
1494 strategies. *Curr infec dis rep* **2013**, *15*, 455-464.
- 1495 254. Peek, F.A.W.; Nell, M.J.; Brand, R.; Jansen-Werkhoven, T.M.; van Hoogdalem, E.J.; Frijns,
1496 J.H.M. In *Double-blind placebo-controlled study of the novel peptide drug p60.4ac in chronic middle ear*
1497 *infection*, 49th Intersci. Conf. Antimicrob. Agents Chemother., San Francisco, USA, 2009; San
1498 Francisco, USA, pp L1-337.

- 1499 255. Hall-Stoodley, L.; Hu, F.Z.; Gieseke, A.; Nistico, L.; Nguyen, D.; Hayes, J.; Forbes, M.;
1500 Greenberg, D.P.; Dice, B.; Burrows, A., *et al.* Direct detection of bacterial biofilms on the middle-ear
1501 mucosa of children with chronic otitis media. *JAMA* **2006**, *296*, 202-211.
- 1502 256. Haisma, E.M.; de Breij, A.; Chan, H.; van Dissel, J.T.; Drijfhout, J.W.; Hiemstra, P.S.; El
1503 Ghalbzouri, A.; Nibbering, P.H. LI-37-derived peptides eradicate multidrug-resistant staphylococcus
1504 aureus from thermally wounded human skin equivalents. *Antimicrob Agents Chemothe* **2014**, *58*, 4411-
1505 4419.
- 1506 257. Church, D.; Elsayed, S.; Reid, O.; Winston, B.; Lindsay, R. Burn wound infections. *Clin.*
1507 *Microbiol. Rev.* **2006**, *19*, 403-434.
- 1508 258. Khurshid, Z.; Najeeb, S.; Mali, M.; Moin, S.F.; Raza, S.Q.; Zohaib, S.; Sefat, F.; Zafar, M.S.
1509 Histatin peptides: Pharmacological functions and their applications in dentistry. *Saudi Pharmaceutical*
1510 *J.* <http://dx.doi.org/10.1016/j.jsps.2016.04.027>
- 1511 259. Liu, Z.; Ma, S.; Duan, S.; Xuliang, D.; Sun, Y.; Zhang, X.; Xu, X.; Guan, B.; Wang, C.; Hu, M.,
1512 *et al.* Modification of titanium substrates with chimeric peptides comprising antimicrobial and
1513 titanium-binding motifs connected by linkers to inhibit biofilm formation. *ACS Appl Mater Interfaces*
1514 **2016**, *8*, 5124-5136.
- 1515 260. Makihira, S.; Nikawa, H.; Shuto, T.; Nishimura, M.; Mine, Y.; Tsuji, K.; Okamoto, K.; Sakai,
1516 Y.; Sakai, M.; Imari, N., *et al.* Evaluation of trabecular bone formation in a canine model surrounding
1517 a dental implant fixture immobilized with an antimicrobial peptide derived from histatin. *J. Mater.*
1518 *Sci.-Mater. Med.* **2011**, *22*, 2765-2772.
- 1519 261. Kong, E.F.; Tsui, C.; Boyce, H.; Ibrahim, A.; Hoag, S.W.; Karlsson, A.J.; Meiller, T.F.; Jabra-
1520 Rizk, M.A. Development and in vivo evaluation of a novel histatin-5 bioadhesive hydrogel
1521 formulation against oral candidiasis. *Antimicrob. Agents Chemother.* **2016**, *60*, 881-889.
- 1522 262. Garcia-Cuesta, C.; Sarrion-Pérez, M.-G.; Bagán, J.V. Current treatment of oral candidiasis: A
1523 literature review. *J Clin Exp Dent* **2014**, *6*, e576-e582.
- 1524 263. Tati, S.; Li, R.; Puri, S.; Kumar, R.; Davidow, P.; Edgerton, M. Histatin 5-spermidine
1525 conjugates have enhanced fungicidal activity and efficacy as a topical therapeutic for oral candidiasis.
1526 *Antimicrob. Agents Chemother.* **2014**, *58*, 756-766.
- 1527 264. How, K.Y.; Song, K.P.; Chan, K.G. Porphyromonas gingivalis: An overview of
1528 periodontopathic pathogen below the gum line. *Front Microbiol* **2016**, *7*, 14.
- 1529 265. Cheng, D.J.; Oppenheim, F.G.; Helmerhorst, E.J. Antifungal formulation and method of
1530 preparation. Google Patents: 2009.
- 1531 266. Pal, S.; Tak, Y.K.; Han, E.; Rangasamy, S.; Song, J.M. A multifunctional composite of an
1532 antibacterial higher-valent silver metallopharmaceutical and a potent wound healing polypeptide: A
1533 combined killing and healing approach to wound care. *New J Chem* **2014**, *38*, 3889-3898.
- 1534 267. Silva, O.N.; Fensterseifer, I.C.M.; Rodrigues, E.A.; Holanda, H.H.S.; Novaes, N.R.F.; Cunha,
1535 J.P.A.; Rezende, T.M.B.; Magalhães, K.G.; Moreno, S.E.; Jerônimo, M.S., *et al.* Clavanin a improves
1536 outcome of complications from different bacterial infections. *Antimicrob Agents Chemother* **2015**, *59*,
1537 1620-1626.
- 1538 268. Li, L.; He, J.; Eckert, R.; Yarbrough, D.; Lux, R.; Anderson, M.; Shi, W. Design and
1539 characterization of an acid-activated antimicrobial peptide. *Chem Biol Drug Des* **2010**, *75*, 127-132.
- 1540 269. Yadav, K.; Prakash, S. Dental caries: A review. *Asian J Biomed Pharm Sci* **2016**, *6*, 1-7.

- 1541 270. Legrand, D.; Pierce, A.; Mazurier, J. Secreted lactoferrin and lactoferrin-related peptides:
1542 Insight into structure and biological functions. In *Bioactive proteins and peptides as functional foods and*
1543 *nutraceuticals*, Wiley-Blackwell: 2010; pp 179-202.
- 1544 271. Mayeur, S.; Spahis, S.; Pouliot, Y.; Levy, E. Lactoferrin, a pleiotropic protein in health and
1545 disease. *Antioxid. Redox Signal.* **2016**, *24*, 813-835.
- 1546 272. Bruni, N.; Capucchio, M.T.; Biasibetti, E.; Pessione, E.; Cirrincione, S.; Giraudo, L.; Corona,
1547 A.; Dosio, F. Antimicrobial activity of lactoferrin-related peptides and applications in human and
1548 veterinary medicine. *Molecules* **2016**, *21*.
- 1549 273. Brouwer, C.P.; Rahman, M.; Welling, M.M. Discovery and development of a synthetic
1550 peptide derived from lactoferrin for clinical use. *Peptides* **2011**, *32*, 1953-1963.
- 1551 274. Theolier, J.; Fliss, I.; Jean, J.; Hammami, R. Milkamp: A comprehensive database of
1552 antimicrobial peptides of dairy origin. *Dairy Sci Technol* **2014**, *94*, 181-193.
- 1553 275. Wakabayashi, H.; Oda, H.; Yamauchi, K.; Abe, F. Lactoferrin for prevention of common viral
1554 infections. *J Infect Chemotherapy* **2014**, *20*, 666-671.
- 1555 276. Zhang, Y.; Lima, C.F.; Rodrigues, L.R. Anticancer effects of lactoferrin: Underlying
1556 mechanisms and future trends in cancer therapy. *Nutr Rev* **2014**, *72*, 763-773.
- 1557 277. Freire, J.M.; Gaspar, D.; Veiga, A.S.; Castanho, M. Shifting gear in antimicrobial and
1558 anticancer peptides biophysical studies: From vesicles to cells. *J Peptid Sc* **2015**, *21*, 178-185.
- 1559 278. Liu, X.; Li, Y.; Li, Z.; Lan, X.; Leung, P.H.M.; Li, J.; Yang, M.; Ko, F.; Qin, L. Mechanism of
1560 anticancer effects of antimicrobial peptides. *J Fiber Bioeng Inform* **2015**, *8*, 25-36.
- 1561 279. Burns, K.E.; McCleerey, T.P.; Thévenin, D. Ph-selective cytotoxicity of phlip-antimicrobial
1562 peptide conjugates. *Scientific Reports* **2016**, *6*, 28465.
- 1563 280. Boohaker, R.J.; Lee, M.W.; Vishnubhotla, P.; Perez, J.M.; Khaled, A.R. The use of therapeutic
1564 peptides to target and to kill cancer cells. *Curr med chem* **2012**, *19*, 3794-3804.
- 1565 281. Nappi, C.; Tommaselli, G.A.; Morra, I.; Massaro, M.; Formisano, C.; Di Carlo, C. Efficacy and
1566 tolerability of oral bovine lactoferrin compared to ferrous sulfate in pregnant women with iron
1567 deficiency anemia: A prospective controlled randomized study. *Acta Obstet Gynecol Scand* **2009**, *88*,
1568 1031-1035.
- 1569 282. Manzoni, P.; Rinaldi, M.; Cattani, S.; Pagni, L.; Romeo, M.G.; Messner, H.; Stolfi, I.;
1570 Decembrino, L.; Laforgia, N.; Vagnarelli, F., *et al.* Bovine lactoferrin supplementation for prevention
1571 of late-onset sepsis in very low-birth-weight neonates: A randomized trial. *JAMA* **2009**, *302*, 1421-
1572 1428.
- 1573 283. de Bortoli, N.; Leonardi, G.; Ciancia, E.; Merlo, A.; Bellini, M.; Costa, F.; Mumolo, M.G.;
1574 Ricchiuti, A.; Cristiani, F.; Santi, S., *et al.* Helicobacter pylori eradication: A randomized prospective
1575 study of triple therapy versus triple therapy plus lactoferrin and probiotics. *Am J Gastroenterol* **2007**,
1576 *102*, 951-956.
- 1577 284. Moreau-Marquis, S.; Coutermarsh, B.; Stanton, B.A. Combination of hypothiocyanite and
1578 lactoferrin (alx-109) enhances the ability of tobramycin and aztreonam to eliminate pseudomonas
1579 aeruginosa biofilms growing on cystic fibrosis airway epithelial cells. *J Antimicrob Chemo* **2015**, *70*,
1580 160-166.
- 1581 285. Boxer, L.A. How to approach neutropenia. *ASH Education Program Book* **2012**, *2012*, 174-182.

- 1582 286. Cooper, C.A.; Maga, E.A.; Murray, J.D. Production of human lactoferrin and lysozyme in the
1583 milk of transgenic dairy animals: Past, present, and future. *Transgenic Res.* **2015**, *24*, 605-614.
- 1584 287. Wang, G.S.; Li, X.; Wang, Z. Apd3: The antimicrobial peptide database as a tool for research
1585 and education. *Nucleic Acids Res* **2016**, *44*, D1087-D1093.
- 1586 288. Harris, F.; Dennison, S.R.; Phoenix, D.A. Anionic antimicrobial peptides from eukaryotic
1587 organisms and their mechanisms of action. *Curr Chem Biol* **2011**, *5*, 142-153.
- 1588 289. Rieg, S.; Seeber, S.; Steffen, H.; Humeny, A.; Kalbacher, H.; Stevanovic, S.; Kimura, A.; Garbe,
1589 C.; Schitteck, B. Generation of multiple stable dermcidin-derived antimicrobial peptides in sweat of
1590 different body sites. *Journal of Investigative Dermatology* **2006**, *126*, 354-365.
- 1591 290. Schmid-Wendtner, M.H.; Korting, H.C. The ph of the skin surface and its impact on the
1592 barrier function. *Skin Pharmacol Physiol* **2006**, *19*, 296-302.
- 1593 291. Mao, Y.; Niu, S.; Xu, X.; Wang, J.; Su, Y.; Wu, Y.; Zhong, S. The effect of an adding histidine
1594 on biological activity and stability of pc-pis from *pseudosciaena crocea*. *Plos One* **2013**, *8*.
- 1595 292. Pace, C.N.; Scholtz, J.M. A helix propensity scale based on experimental studies of peptides
1596 and proteins. *Biophys J* **1998**, *75*, 422-427.
- 1597 293. Embleton, N.D.; Berrington, J.E.; McGuire, W.; Stewart, C.J.; Cummings, S.P. Lactoferrin:
1598 Antimicrobial activity and therapeutic potential. *Seminars in Fetal and Neonatal Medicine* **2013**, *18*, 143-
1599 149.
- 1600 294. Krulwich, T. *Bacterial energetics: A treatise on structure and function*. Elsevier Science: 2012.
- 1601 295. Krulwich, T.A.; Sachs, G.; Padan, E. Molecular aspects of bacterial ph sensing and
1602 homeostasis. *Nature Reviews Microbiology* **2011**, *9*, 330-343.
- 1603 296. Kaneti, G.; Meir, O.; Mor, A. Controlling bacterial infections by inhibiting proton-dependent
1604 processes. *Biochimica et biophysica acta* **2016**, *1858*, 995-1003.
- 1605 297. Peters, B.M.; Shirliff, M.E.; Jabra-Rizk, M.A. Antimicrobial peptides: Primeval molecules or
1606 future drugs? *PLoS pathogens* **2010**, *6*.
- 1607 298. Wilmes, M.; Cammue, B.P.A.; Sahl, H.G.; Thevissen, K. Antibiotic activities of host defense
1608 peptides: More to it than lipid bilayer perturbation. *Nat Prod Rep* **2011**, *28*, 1350-1358.
- 1609 299. Farha, M.A.; Verschoor, C.P.; Bowdish, D.; Brown, E.D. Collapsing the proton motive force
1610 to identify synergistic combinations against *staphylococcus aureus*. *Chem Biol* **2013**, *20*, 1168-1178.
- 1611 300. Lee, J.; Lee, D.; Choi, H.; Kim, H.H.; Kim, H.; Hwang, J.S.; Lee, D.G.; Kim, J.I. Synthesis and
1612 antimicrobial activity of cysteine-free coprisin nonapeptides. *Biochem Biophys Res Commun* **2014**, *443*,
1613 483-488.
- 1614 301. Nagoba, B.S.; Suryawanshi, N.M.; Wadher, B.; Selkar, S. Acidic environment and wound
1615 healing: A review. *Wounds-a Compendium of Clinical Research and Practice* **2015**, *27*, 5-11.
- 1616 302. Walkenhorst, W.F.; Klein, J.W.; Vo, P.; Wimley, W.C. Ph dependence of microbe sterilization
1617 by cationic antimicrobial peptides. *Antimicrob. Agents Chemother.* **2013**, *57*, 3312-3320.
- 1618 303. Lienkamp, K.; Madkour, A.E.; Tew, G.N. Antibacterial peptidomimetics: Polymeric synthetic
1619 mimics of antimicrobial peptides. In *Polymer composites - polyolefin fractionation - polymeric*
1620 *peptidomimetics - collagens*, Abe, A.; Kausch, H.H.; Moller, M.; Pasch, H., Eds. 2013; Vol. 251, pp 141-
1621 172.
- 1622 304. Kharidia, R.; Tu, Z.; Chen, L.; Liang, J.F. Activity and selectivity of histidine-containing lytic
1623 peptides to antibiotic-resistant bacteria. *Arch Microbiol* **2012**, *194*, 769-778.

- 1624 305. Arnusch, C.J.; Albada, H.B.; Liskamp, R.M.J.; Shai, Y. Nanostructure determines antifungal
1625 activity of de novo designed ph dependent histidine containing ultra-short lipopeptides. *Biophys J*
1626 **2010**, *98*, 278A-279A.
- 1627 306. Arnusch, C.J.; Albada, H.B.; van Vaardegem, M.; Liskamp, R.M.J.; Sahl, H.-G.; Shadkchan,
1628 Y.; Osheroov, N.; Shai, Y. Trivalent ultrashort lipopeptides are potent ph dependent antifungal agents.
1629 *J Med Chem* **2012**, *55*, 1296-1302.
- 1630 307. Lu, S.; Bennett, W.F.D.; Ding, Y.; Zhang, L.; Fan, H.Y.; Zhao, D.; Zheng, T.; Ouyang, P.-K.; Li,
1631 J.; Wu, Y., *et al.* Design and characterization of a multifunctional ph-triggered peptide c8 for selective
1632 anticancer activity. *Adv Healthc Mat* **2015**, *4*, 2709-2718.
- 1633 308. Callahan, D.J.; Liu, W.; Li, X.; Dreher, M.R.; Hassouneh, W.; Kim, M.; Marszalek, P.; Chilkoti,
1634 A. Triple stimulus-responsive polypeptide nanoparticles that enhance intratumoral spatial
1635 distribution. *Nano Lett* **2012**, *12*, 2165-2170.
- 1636 309. Han, S.-S.; Li, Z.-Y.; Zhu, J.-Y.; Han, K.; Zeng, Z.-Y.; Hong, W.; Li, W.-X.; Jia, H.-Z.; Liu, Y.;
1637 Zhuo, R.-X., *et al.* Dual-ph sensitive charge-reversal polypeptide micelles for tumor-triggered
1638 targeting uptake and nuclear drug delivery. *Small* **2015**, *11*, 2543-2554.
- 1639 310. Hwang, J.-H.; Choi, C.W.; Kim, H.-W.; Kim, D.H.; Kwak, T.W.; Lee, H.M.; Kim, C.H.; Chung,
1640 C.W.; Jeong, Y.-I.; Kang, D.H. Dextran-b-poly(l-histidine) copolymer nanoparticles for ph-responsive
1641 drug delivery to tumor cells. *Internat J Nanomed* **2013**, *8*, 3197-3207.
- 1642 311. Ferrer-Miralles, N.; Luis Corchero, J.; Kumar, P.; Cedano, J.A.; Gupta, K.C.; Villaverde, A.;
1643 Vazquez, E. Biological activities of histidine-rich peptides; merging biotechnology and
1644 nanomedicine. *Microbial Cell Factories* **2011**, *10*.
- 1645 312. Majdoul, S.; Seye, A.K.; Kichler, A.; Holic, N.; Galy, A.; Bechinger, B.; Fenard, D. Molecular
1646 determinants of vectofusin-1 and its derivatives for the enhancement of lentivirally mediated gene
1647 transfer into hematopoietic stem/progenitor cells. *J Bioll Chem* **2016**, *291*, 2161-2169.
- 1648 313. Wakabayashi, N.; Yano, Y.; Kawano, K.; Matsuzaki, K. A ph-dependent charge reversal
1649 peptide for cancer targeting. *Eur biophys j : EBJ* **2016**.
- 1650 314. Sousa, A.M.; Pereira, M.O. *Pseudomonas aeruginosa* diversification during infection
1651 development in cystic fibrosis lungs—a review. *Pathogens* **2014**, *3*, 680-703.
- 1652 315. Folkesson, A.; Jelsbak, L.; Yang, L.; Johansen, H.K.; Ciofu, O.; Hoiby, N.; Molin, S. Adaptation
1653 of *pseudomonas aeruginosa* to the cystic fibrosis airway: An evolutionary perspective. *Nat Rev Micro*
1654 **2012**, *10*, 841-851.
- 1655 316. Lashua, L.P.; Melvin, J.A.; Deslouches, B.; Pilewski, J.M.; Montelaro, R.C.; Bomberger, J.M.
1656 Engineered cationic antimicrobial peptide (ecap) prevents *pseudomonas aeruginosa* biofilm growth
1657 on airway epithelial cells. *J Antimicrob Chemother.* **2016**, *71*, 2200-2207.
- 1658
1659
1660
1661
1662
1663