

# Zebrafish-based discovery of antiseizure compounds from the Red Sea: pseurotin A<sub>2</sub> and azaspirofuran A

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## **ABSTRACT**

In search for novel antiseizure drugs (ASDs), the European FP7-funded PharmaSea project used zebrafish embryos and larvae as a drug discovery platform to screen marine natural products to identify promising antiseizure hits *in vivo* for further development. Within the framework of this project, 7 known hetero-spirocyclic  $\gamma$ -lactams, namely pseurotin A, pseurotin A<sub>2</sub>, pseurotin F1, 11-*O*-methylpseurotin A, pseurotin D, azaspirofuran A, and azaspirofuran B, were isolated from the bioactive marine fungus *Aspergillus fumigatus*, and their antiseizure activity was evaluated in the larval zebrafish pentylenetetrazole (PTZ) seizure model. Pseurotin A<sub>2</sub> and azaspirofuran A were identified as antiseizure hits, while their close chemical analogues were inactive. Besides, electrophysiological analysis from the zebrafish midbrain demonstrated that pseurotin A<sub>2</sub> and azaspirofuran A also ameliorate PTZ-induced epileptiform discharges. Next, to determine whether these findings translate to mammals, both compounds were analyzed in the mouse 6-Hz (44 mA) psychomotor seizure model. They lowered the seizure duration dose-dependently, thereby confirming their antiseizure properties and suggesting activity against drug-resistant seizures. Finally, in a thorough ADMET assessment, pseurotin A<sub>2</sub> and azaspirofuran A were found to be drug-like. Based on the prominent antiseizure activity in both species and the drug-likeness, we propose pseurotin A<sub>2</sub> and azaspirofuran A as lead compounds that are worth further investigation for the treatment of epileptic seizures. This study not only provides the first evidence of antiseizure activity of pseurotins and azaspirofurans, but also demonstrates the value of the zebrafish model in (marine) natural product drug discovery in general, and for ASD discovery in particular.

## **KEYWORDS**

epilepsy, antiseizure drug discovery, marine drug discovery, pseurotin A<sub>2</sub>, azaspirofuran A, *Aspergillus fumigatus*

## INTRODUCTION

Epilepsy is one of the most common neurological conditions, affecting more than 70 million people of all ages with no geographical, social, or racial boundaries.<sup>1-3</sup> It is a disease of the brain that is characterized by spontaneous recurrent unprovoked seizures.<sup>4</sup> Despite an exponential growth of marketed antiseizure drugs (ASDs) over the past 25 years, seizures remain uncontrolled in one third of the patients due to drug resistance.<sup>5, 6</sup> As uncontrolled epilepsy is associated with increased physical and physiological comorbidities and increased risk of sudden unexplained death, there is a substantial burden on the patients, their caretakers and society.<sup>6</sup> Hence, more efficacious ASDs that can treat patients with drug-resistant seizures are sorely needed.<sup>7</sup>

The EU PharmaSea project, funded by the Seventh Framework Programme (FP7), is an SME-academia-driven project of 24 partners that was initiated in 2012 to discover novel antibiotic, anti-inflammatory and neuroactive compounds from marine microorganisms isolated from some of the deepest, coldest, and hottest places on the planet.<sup>8</sup> Neuroactive drug discovery focused on the identification of compounds with the potential to treat epilepsy. To the best of our knowledge, we have been the first to perform systematic large-scale screening of marine-derived natural products for antiseizure drug candidates. Marine species offer an attractive source for drug discovery because they produce potent, selective, and structurally novel bioactive secondary metabolites to defend themselves, to locate mates, and to out-compete competitors for limited resources.<sup>8-10</sup> Many of these do not have a terrestrial analogue and are thus unique in terms of chemical structure and bioactivity.<sup>8, 9</sup> Among them are marine-derived natural products that can alter mammalian neurological activity. Classical examples are tetrodotoxins, saxitoxins, kainic and domoic acids, cone snail venom peptides, and sea anemone toxins.<sup>10</sup> Nevertheless, marine natural products are highly underutilized in drug discovery.<sup>11</sup> A main reason is the lack of systematic analysis in high-content bioassays given the often limited quantities of marine natural products available for screening purposes.<sup>11</sup>

The zebrafish model is suitable for large-scale screening and captures the complexity of a whole body organism, including the central nervous system. As a vertebrate, zebrafish are highly similar to humans due to a high genetic, physiological and pharmacological conservation.<sup>12-14</sup> Moreover, given the small size of embryos and larvae, they fit in wells of microtiter plates and hence are suitable for medium to high-throughput testing.<sup>13, 14</sup> Given the low volumes used in 96- and 384-well plates, zebrafish larvae only require small amounts of sample in the low microgram range when added to their swimming water and even less when administered by

injection. This property is of particular interest for marine natural product drug discovery, where material is often scarce.<sup>11</sup> Within the PharmaSea project, marine samples were screened in 96-well plate format using only 10 µg per well per screening round. From the established larval zebrafish seizure and epilepsy models<sup>15</sup>, the larval zebrafish pentylenetetrazole (PTZ) seizure model was chosen for drug discovery because: 1) the model has been extensively characterized in terms of behavioral and non-behavioral seizure markers<sup>16, 17</sup>, 2) it has been pharmacologically characterized with ASDs on the market<sup>16-18</sup>, 3) results translate well to rodent models<sup>17, 19, 20</sup>, and 4) the behavioral assay is suitable for high-throughput screening – seizures can easily and rapidly be induced by a single administration of the convulsant drug to the larva's aqueous environment<sup>16</sup> and can be quantified by video recording<sup>16-18</sup>.

Within the framework of the PharmaSea project, 5 pseurotins (pseurotin A, pseurotin A<sub>2</sub>, pseurotin F1, 11-*O*-methylpseurotin A, and pseurotin D) and 2 azaspirofurans (azaspirofuran A and B) were isolated from extracts of the bioactive marine fungus *Aspergillus fumigatus*, which was collected from a Red Sea sediment in Hurghada, Egypt. **Interestingly, these compounds did not demonstrate antibacterial activity or cytotoxicity in contrast to other constituents from the crude extract<sup>21</sup> and were readily available for further biological evaluation.** Pseurotins are a family of fungal secondary metabolites that have not been well studied so far, except for pseurotin A that was found to exhibit a range of bioactivities at moderate to high test concentrations (up to 50 µg/mL).<sup>22-31</sup> Azaspirofurans are chemically very similar to pseurotins, but feature an ethyl furan ring instead of a vicinal diol (Figure 1). Little is known about the bioactivities of azaspirofurans as they were identified only recently.<sup>32</sup> So far, azaspirofuran A was observed to specifically inhibit the proliferation of the A459 cancer cell line.<sup>32, 33</sup>

All compounds were investigated for antiseizure activity in the larval zebrafish PTZ seizure model. Interestingly, despite close structural similarities, only pseurotin A<sub>2</sub> and azaspirofuran A ameliorated PTZ-induced seizures, suggesting a highly specific interaction. Besides, electrophysiological analysis from the zebrafish midbrain demonstrated that pseurotin A<sub>2</sub> and azaspirofuran A also significantly lowered PTZ-induced epileptiform discharges. Next, the antiseizure activity of pseurotin A<sub>2</sub> and azaspirofuran A was demonstrated in the mouse 6-Hz (44 mA) psychomotor seizure model. These results not only confirmed the translation of findings from zebrafish larvae to mice but also suggested that pseurotin A<sub>2</sub> and azaspirofuran A are effective against drug-resistant focal seizures. Finally, ADMET profiling showed that both compounds are drug-like. Based on the prominent antiseizure activity and the promising

ADMET characteristics, we propose pseurotin A<sub>2</sub> and azaspirofurans A as lead compounds that are worth further investigation for the treatment of seizures.

## RESULTS AND DISCUSSION

### Isolation and structural elucidation of compounds from the bioactive marine fungus *Aspergillus fumigatus*

The marine fungal isolate MR2012 used in this study was isolated from a Red Sea sediment in Hurghada, Egypt and taxonomically identified on a molecular basis as *Aspergillus fumigatus*<sup>21</sup>. Compounds pseurotin A<sup>34</sup>, pseurotin A<sub>2</sub><sup>27</sup>, pseurotin F1<sup>35</sup>, 11-*O*-methylpseurotin A<sup>34</sup>, pseurotin D<sup>36</sup>, and azaspirofurans A and B<sup>32</sup>, all known hetero-spirocyclic  $\gamma$ -lactams, were isolated from the CH<sub>2</sub>Cl<sub>2</sub> fraction of the fungal fermentation<sup>21, 37</sup> (Figure 1). Their structures were confirmed by HRESIMS analysis and by comparing the 1D and 2D NMR spectra and optical rotation with literature data as indicated (see supporting information for their 1D NMR data).

### Azaspirofurans A and pseurotin A<sub>2</sub> ameliorate seizures in the zebrafish PTZ seizure model

To investigate whether the isolated compounds display antiseizure activity, they were tested in the larval zebrafish PTZ seizure model both after a short (2 hours (h)) and long (18 h) incubation time at their maximum tolerated concentration (MTC) (Table 1). The MTC was defined as the highest concentration at which no larvae died nor showed signs of toxicity or locomotor impairment in comparison to vehicle (VHC)-treated control larvae. In case no MTC was reached, 100  $\mu$ g/mL was used as the test concentration. In line with studies previously reported<sup>16, 17</sup>, addition of the GABA<sub>A</sub>-receptor antagonist PTZ to the swimming water of 7-days post-fertilization (dpf) zebrafish larvae strongly elevated larval locomotion ( $p \leq 0.001$ ) (Figure 2 and 3) as a result of induced seizures (or seizure-like behavior) that were recognized as typical high-speed swimming, whirlpool-like circling, clonus-like seizures, and loss of posture, as previously described<sup>16</sup>. Azaspirofurans A ( $p \leq 0.01$ ) and pseurotin A<sub>2</sub> ( $p \leq 0.001$ ) significantly lowered PTZ-induced seizures of larvae after 2 and 18 h of compound exposure, respectively (Figure 2). Remarkably, none of the chemical analogues showed notable activities despite their close structural similarities. These results suggest that azaspirofurans A and pseurotin A<sub>2</sub> specifically interact with their antiseizure target(s). In contrast to azaspirofurans A, azaspirofurans B lacks the methoxyl group (Figure 1), which seems necessary for the activity

against PTZ-induced seizures. It is unlikely that the activity of azaspirofurans A is due to an improved compound uptake as the LogP of azaspirofurans A and B is predicted to be 3.98 and 3.81 (see methods), respectively, and good compound bioavailability in zebrafish larvae is expected from a LogP of 1 onwards<sup>38</sup>. The close analogues pseurotin A and A<sub>2</sub> are diastereomers with different configurations at C-8 and C-9 (Figure 1).<sup>27</sup> Their structural differences do not affect uptake, which is also expected to be adequate as the compounds are predicted to have a LogP of 3.23 (see methods). So, likely the structural differences result in distinct pharmacological activities rather than in altered bioavailabilities. Nevertheless the pharmacokinetics of the individual compounds in zebrafish larvae are currently unknown, so it cannot be ruled out that the actual brain concentrations of azaspirofurans A and pseurotin A<sub>2</sub> are higher than those of their inactive analogues. Of note, like azaspirofurans A, pseurotin A, pseurotin D, and 11-*O*-methylpseurotin A also possess the methoxyl group and are not active, which can be due to the absence of the ethyl furan ring. These observations suggest that the molecular target(s) of azaspirofurans A and pseurotin A<sub>2</sub> are not necessarily the same. Interestingly, antiseizure activity has not yet been reported for azaspirofurans or pseurotins.

To investigate the concentration-response relationship, azaspirofurans A and pseurotin A<sub>2</sub> were retested at their MTC, MTC/2, and MTC/4 (two-fold serial dilution) in the zebrafish PTZ seizure model at their optimal incubation time (2 and 18 h, respectively) in three independent experiments (Figure 3). Azaspirofurans A lowered PTZ-induced seizure behavior to the same extent as before at the MTC over the 30-minute (min) recording period (Figure 3A). A more detailed analysis of the 5-min time intervals from the 30-min recording period revealed a significant reduction in the 10-30 min time window ( $p \leq 0.001$ ) (Figure 3B). No antiseizure activity was seen at lower concentrations. Pseurotin A<sub>2</sub> showed concentration-dependent activity against PTZ-induced seizure behavior, both within the 30-min recording period ( $p \leq 0.01$ , Figure 3C) as during the 5-min time intervals ( $p \leq 0.001$ , Figure 3D).

### **Azaspirofurans A and pseurotin A<sub>2</sub> ameliorate epileptiform brain activity in the zebrafish PTZ seizure model**

To determine whether azaspirofurans A and pseurotin A<sub>2</sub> can ameliorate the PTZ-induced hyperexcitable state of the brain that is characterized by epileptiform discharges<sup>39</sup>, local field potential (LFP) recordings<sup>40</sup> were non-invasively measured from the midbrain (optic tectum) of zebrafish larvae. To that end larvae were treated with either VHC or test compound (MTC

and optimal incubation time was used) followed by a 15 min during exposure to PTZ or VHC prior to the electrophysiology measurements (Figure 4 and 5). A larva was considered to have epileptiform brain activity when at least 3 electrical discharges were seen in the 10-min recording period that fulfilled the pre-defined requirements of an epileptiform event (see methods). Pre-incubation with azaspirofurane A significantly reduced ( $p \leq 0.001$ ) the percentage of larvae with PTZ-induced epileptiform activity with almost 60 % in comparison to PTZ-treated controls (Figure 4A). Larvae also showed significantly less epileptiform events ( $p \leq 0.001$ ) when pre-exposed to azaspirofurane A, resulting in a shorter cumulative duration of events ( $p \leq 0.001$ ) over the 10-min recording period (Figure 4B and C). Pseurotin A<sub>2</sub> only non-significantly lowered the percentage of larvae with PTZ-induced epileptiform activity with 33 %, in comparison to controls (Figure 4D). However, larvae did show significantly less epileptiform events ( $p \leq 0.01$ ), resulting in a shorter cumulative duration of events ( $p \leq 0.05$ ) over the 10-min recording period (Figure 4E and F). Thus, both compounds not only ameliorate PTZ-induced seizures but are likely to do so by lowering the PTZ-induced hyperexcitable state of the brain. Of note, azaspirofurane A or pseurotin A<sub>2</sub> did not induce abnormal electrical discharges in comparison to VHC-treated controls.

### **Azaspirofurane A and pseurotin A<sub>2</sub> ameliorate focal seizures in the mouse 6-Hz (44 mA) psychomotor seizure model**

Although the zebrafish model has a high degree of genetic, physiological, and pharmacological conservation<sup>13</sup>, it is more distinct from humans than rodents. Therefore, we were interested to see if the observed antiseizure properties of azaspirofurane A and pseurotin A<sub>2</sub> would translate to a rodent model. The mouse 6-Hz (44 mA) psychomotor seizure model was chosen because it can detect compounds with novel antiseizure mechanisms and with potential against drug-resistant seizures.<sup>41, 42</sup> In this model drug-resistant focal impaired awareness seizures<sup>43</sup>, previously referred to as complex partial or psychomotor seizures<sup>44</sup>, are induced by low frequency, long duration corneal electrical stimulation. Mice injected i.p. with VHC (30 min before electrical stimulation) had a mean ( $\pm$  SD) seizure duration of 50 seconds (s) ( $\pm$  19 s) (Figure 6). In line with previous studies<sup>41, 45</sup>, i.p. administration of the positive control valproate (300 mg/kg dose, 30 min before electrical stimulation) protected all mice against the induced seizures, significantly reducing the seizure duration to a mean of 4 s ( $\pm$  6 s) ( $p \leq 0.001$ ). In contrast, i.p. administration of the negative control phenytoin (10 mg/kg dose, 120 min before electrical stimulation, as reported by Barton and colleagues<sup>41</sup>) did not significantly affect the

seizure duration (mean duration of 30 s ( $\pm$  13 s)) as previously published<sup>41, 46, 47</sup>. Administration of 40 mg/kg azaspirofurane A (i.p. injected, 30 min before electrical stimulation) significantly lowered the seizure duration to a mean of 24 s ( $\pm$  18 s) ( $p \leq 0.05$ ) in comparison to VHC-controls. Administration of 40 mg/kg pseurotin A<sub>2</sub> (i.p. injected, 30 min before electrical stimulation) significantly lowered the seizure duration to a mean of 26 s ( $\pm$  18 s) ( $p \leq 0.05$ ) in comparison to controls. Treatment with azaspirofurane A as well as with pseurotin A<sub>2</sub> showed a trend for dose-dependent reduction in seizure duration. 20 and 10 mg/kg azaspirofurane A non-significantly lowered the seizure duration to a mean of 41 and 46 s ( $\pm$  15 and 20 s), respectively. 10 and 2.5 mg/kg pseurotin A<sub>2</sub> non-significantly lowered the seizure duration to a mean of 27 and 42 s ( $\pm$  26 and 11 s), respectively. These data confirm the antiseizure properties of azaspirofurane A and pseurotin A<sub>2</sub> in a standard mouse model of drug-resistant focal seizures. The identification and validation of these novel antiseizure hits thereby demonstrate the effectiveness of using the larval zebrafish model for ASD discovery. Besides, this study provides another example of the translation of results from zebrafish to rodent seizure models. Little is known about the molecular mechanisms underlying the bioactivities of azaspirofurane A and pseurotin A<sub>2</sub> in general. Therefore, it is difficult to speculate on the antiseizure drug targets that could be involved. Of note, pseurotin A<sub>2</sub> was reported to exert anti-inflammatory activity, a promising mode of action that has been proposed for the development of innovative ASDs and AEDs as neuroinflammation is involved in the origin of seizures and epilepsy.<sup>29, 48, 49</sup> In addition, pseurotin A is known to induce neuronal cell differentiation.<sup>22</sup> This long-term effect is unlikely to be involved in the antiseizure actions of azaspirofurane A and pseurotin A<sub>2</sub> in this study because of the short-term exposure (2 and 18 h of exposure, respectively), but it is of particular interest for AED development. Further research is needed to unravel the molecular mechanisms of azaspirofurane A and pseurotin A<sub>2</sub> that are responsible for their antiseizure activity against PTZ-induced seizures in zebrafish and focal seizures in mice.

### **ADMET profiling of azaspirofurane A and pseurotin A<sub>2</sub>**

Finally, to define the drug-likeness of the antiseizure lead compounds, the ADMET profiles of azaspirofurane A and pseurotin A<sub>2</sub> were elucidated using standard *in vitro* assays. The ADMET results are summarized in Table 2. No notable cytotoxicity or cardiotoxicity was observed for either one of the compounds. Both compounds showed an acceptable solubility and azaspirofurane A also demonstrated a high permeability, a desired combination that is not common. Furthermore, azaspirofurane A only weakly inhibited the CYP2D6 and CYP2C9



enzymes and did not affect the CYP3A4 enzyme, while pseurotin A<sub>2</sub> did not inhibit any of the three types of CYP450 enzymes. Thus, azaspirofuran A and pseurotin A<sub>2</sub> are unlikely to present drug-drug interactions. In addition, they are metabolically stable with a half-life ( $t_{1/2}$ ) of 33 and > 60 min for azaspirofuran A and pseurotin A<sub>2</sub>, respectively. Finally, azaspirofuran A showed a high level of plasma protein binding (95 %), which is not optimal but can be addressed, and pseurotin A<sub>2</sub> has a much lower plasma protein binding of only 37 % with good recovery. Thus, azaspirofuran A and pseurotin A<sub>2</sub> show promising ADMET characteristics and are therefore drug-like. Hence, we propose azaspirofuran A and pseurotin A<sub>2</sub> as lead compounds worth further investigation for the treatment of epileptic seizures in general, and drug-resistant focal seizures in particular.

## CONCLUSIONS

In this study, 7 known hetero-spirocyclic  $\gamma$ -lactams were isolated from the marine sediment-derived fungus *Aspergillus fumigatus* MR2012 and investigated for antiseizure activity in the larval zebrafish PTZ seizure model. Pseurotin A<sub>2</sub> and azaspirofuran A were found to have promising antiseizure activity, not present in the other structural analogues tested and thus suggested to have a structure-specific interaction with, possibly new, antiseizure drug targets. The antiseizure activity of pseurotin A<sub>2</sub> and azaspirofuran A translated to a mouse model of drug-resistant focal seizures and in addition, both compounds were observed to be drug-like. Based on the prominent antiseizure activity in zebrafish and mice and their drug-likeness, we propose pseurotin A<sub>2</sub> and azaspirofuran A as lead compounds that are worth further investigation for the treatment of epileptic seizures. Thereby, this study provides the first evidence of antiseizure activity of pseurotins and azaspirofurans, and gives another example of the translation of results from zebrafish larvae to mice. Moreover, this study demonstrates the value of the zebrafish model in (marine) natural product drug discovery in general, and for ASD discovery in particular. A detailed structure-activity relationship investigation is needed to understand the structural necessities of pseurotins and azaspirofurans to exert their antiseizure action. Moreover, further research is needed to unravel the molecular mechanisms of pseurotin A<sub>2</sub> and azaspirofuran A that are responsible for their antiseizure activity against PTZ-induced seizures in zebrafish and focal seizures in mice.

## **METHODS**

### **Chemical experimental procedures**

NMR data were acquired on a Varian Inova 600 MHz NMR spectrometer. High resolution mass spectrometric data were obtained using a Thermo LTQ Orbitrap coupled to an HPLC system (PDA detector, PDA autosampler, and pump). The following conditions were used: capillary voltage of 45 V, capillary temperature of 260 °C, auxiliary gas flow rate of 10-20 arbitrary units, sheath gas flow rate of 40-50 arbitrary units, spray voltage of 4.5 kV, and mass range of 100-2000 amu (maximal resolution of 30,000). For LC/MS, a C<sub>18</sub> analytical HPLC column (5 µm, 4.6 mm × 150 mm) was used with a mobile phase of 0 to 100 % MeOH over 30 min at a flow rate of 1 mL min<sup>-1</sup>. Biotage Flash system SP1-XOB1, Charlottesville WA, USA was used for initial purification. Compound purification was conducted using Agilent 1200 HPLC system with a Waters Sunfire C<sub>18</sub> column (5 µm, 100 Å, 10 mm × 250 mm), connected to a binary pump, and monitored using a photodiode array detector.

### **Microbial strain**

The marine fungal isolate MR2012 used in this study was isolated from a Red Sea sediment in Hurghada, Egypt in September 2011, and taxonomically identified on a molecular basis as *Aspergillus fumigatus*<sup>21</sup>.

### **Microbial fermentation, extraction, and isolation**

The fungal isolate MR2012 initially cultured on a solid medium composed of (g/L) glucose 10, yeast extract 10, malt extract 4. A 6 liter fermentation was conducted on a medium composed of (g/L) sucrose 100, glucose 10, casamino acids 0.1, yeast extract 5, MOPS 21, K<sub>2</sub>SO<sub>4</sub> 0.25 × 10<sup>-6</sup>, MgCl<sub>2</sub>·6H<sub>2</sub>O 1.0 × 10<sup>-6</sup> for 12 days at 30 °C with shaking at 180 rpm. At the end of the incubation period, Diaion HP-20 resin was added to the culture media and shaken for 6 h at 180 rpm, then cultures were centrifuged (3000 rpm for 20 min) where the residue composed of the fungal mycelia and resin were washed with distilled water twice and extracted with MeOH, and subjected to LC-HRESIMS analysis. This extract was fractionated successively with *n*-hexane (3 × 250 mL), CH<sub>2</sub>Cl<sub>2</sub> (3 × 300 mL), and then EtOAc (3 × 250 mL). Each solvent fraction was evaporated *in vacuo* and subjected to LC-HRESIMS and <sup>1</sup>H NMR analysis, which revealed that the CH<sub>2</sub>Cl<sub>2</sub> fraction was the one of interest to follow. This CH<sub>2</sub>Cl<sub>2</sub> fraction was loaded on Flash Biotage using a FLASH 65i cartridge, solvent MeOH/water 0-100 %, flow rate 60 mL/min over

20 min and UV collection wavelengths 225 and 254 nm to produce 6 fractions. All of these fractions were monitored by LC-HRESIMS. Injection of fraction 4 into Agilent HPLC system using semi-preparative Sunfire C<sub>18</sub> column (250 × 10 mm, 5 μm) with CH<sub>3</sub>CN:H<sub>2</sub>O 30-90 % over 30 min with a 2 mL/min flow led to the isolation of 38 mg of pseurotin A, 30 mg of pseurotin A<sub>2</sub>, 2 mg of pseurotin F1, 6 mg of 11-*O*-methylpseurotin A, and 5 mg of pseurotin D. Injection of fraction 5 into Agilent HPLC system using semi-preparative Sunfire C<sub>18</sub> column (250 × 10 mm, 5 μm) with CH<sub>3</sub>CN:H<sub>2</sub>O 40-80 % over 30 min with a 2 mL/min flow led to the isolation of 26 mg of azaspirofuran A and 23 mg of azaspirofuran B.

### **Compound preparation**

For experiments with zebrafish larvae, dry samples were dissolved in 100 % dimethyl sulfoxide (DMSO, spectroscopy grade) as 100-fold concentrated stocks and diluted in embryo medium to a final concentration of 1 % DMSO content. Control groups were treated with 1 % DMSO (VHC) in accordance with the final solvent concentration of tested compounds. For mice experiments, a mixture of poly-ethylene glycol M.W. 200 (PEG200), 100 % DMSO (spectroscopy grade), and demineralized water (PEG200:DMSO:water; 0.25:0.25:0.5) was used as solvent and VHC.

### **Compound LogP prediction**

LogP (ACD/LogP) values were obtained from ChemSpider and predicted by means of the ACD/Labs Percepta Platform (PhysChem Module) based on the compound structure<sup>50</sup>.

### **Experimental animals**

All animal experiments carried out were approved by the Ethics Committee of the University of Leuven (approval numbers 101/2010, 061/2013, 150/2015, and 023/2017) and by the Belgian Federal Department of Public Health, Food Safety & Environment (approval number LA1210199).

#### *Zebrafish*

Adult zebrafish (*Danio rerio*) stocks of AB strain (Zebrafish International Resource Center, Oregon, USA) were maintained at 28 °C, on a 14/10-h light/dark cycle under standard

aquaculture conditions. Fertilized eggs were collected via natural spawning and raised in embryo medium (1.5 mM HEPES, pH 7.2, 17.4 mM NaCl, 0.21 mM KCl, 0.12 mM MgSO<sub>4</sub>, and 0.18 mM Ca(NO<sub>3</sub>)<sub>2</sub>, and 0.6 μM methylene blue) at 28 °C, under constant light.

### *Mice*

Male NMRI mice (weight 18-20 g) were acquired from Charles River Laboratories and housed in poly-acrylic cages under a 14/10-h light/dark cycle at 21 °C. The animals were fed a pellet diet and water *ad libitum*, and were allowed to acclimate for one week before experimental procedures were conducted. Prior to the experiment, mice were isolated in a poly-acrylic cage with a pellet diet and water *ad libitum* for habituation overnight in the experimental room, to minimize stress.

## **Zebrafish pentylenetetrazole seizure model**

### *Toxicity evaluation*

Maximum tolerated concentration (MTC) was determined prior to further experiments and used as the highest test concentration. Experimental procedure was described before<sup>51</sup>. In brief, the MTC was investigated by exposing 12 larvae of 6 dpf to a range of concentrations in a 100 μL volume during 18 h. The following parameters were investigated after 2 and 18 h of exposure: touch response, morphology, posture, edema, signs of necrosis, swim bladder, and heartbeat. MTC was defined as the highest concentration at which no larvae died nor showed signs of toxicity or locomotor impairment in comparison to VHC-treated control larvae. In case no MTC was reached, 100 μg/mL was used as the highest test concentration.

### *Behavioral analysis*

Experimental procedure was described before<sup>17, 20</sup>. In brief, a single 7-dpf larva (in case of 2 h incubation) or 6-dpf larva (in case of 18 h incubation) was placed in each well of a 96-well plate and treated with either VHC (1 % DMSO) or compound in a 100 μL volume. Larvae were incubated in dark for 2 or 18 h at 28 °C, whereafter 100 μL of either VHC (embryo medium) or 40 mM PTZ was added to each well. Next, within 5 min the 96-well plate was placed in an automated tracking device (ZebraBox Viewpoint, France) and larval behavior was video recorded for 30 min. The complete procedure was performed in dark conditions using infrared light. Total locomotor activity was recorded by ZebraLab software (Viewpoint, France) and

expressed in actinteg units, which is the sum of pixel changes detected during the defined time interval (5 min). Larval behavior was depicted as mean actinteg values in the 30-min recording period and over 5 min time intervals. Data are expressed as mean  $\pm$  SD or as mean  $\pm$  SEM when the means of independent experiments were pooled.

### *Electrophysiology*

Experimental procedure was described before<sup>51-53</sup>. In brief, non-invasive LFP recordings were measured from the midbrain of 7-dpf zebrafish larvae pre-incubated with VHC only, PTZ only, compound and VHC, or compound and PTZ. Larvae were incubated for approximately 2 or 18 h with VHC (1 % DMSO) or compound in a 100  $\mu$ L volume (28 °C). After incubation, 100  $\mu$ L VHC (embryo medium) or 40 mM PTZ (20 mM working concentration) was added to the well for 15 min (28 °C) prior to the recording. For electrophysiological recordings (room temperature), larvae were immobilized in 2 % low melting point agarose (Invitrogen) and the signal electrode [an electrode inside a soda-glass pipet (1412227, Hilgenberg) pulled with a DMZ Universal Puller (Zeitz, Germany), diameter  $\pm$  20 microns, containing artificial cerebrospinal fluid (ACSF: 124 mM NaCl, 10 mM glucose, 2 mM KCl, 2 mM MgSO<sub>4</sub>, 2 mM CaCl<sub>2</sub>, 1.25 mM KH<sub>2</sub>PO<sub>4</sub>, and 26 mM NaHCO<sub>3</sub>, 300-310 mOsmols)] was positioned on the skin covering the optic tectum. Each recording lasted 600 s and was analyzed manually by quantifying the number, cumulative duration, and mean duration of epileptiform-like events with Clampfit 10.2 software (Molecular Devices Corporation, USA). An electrical discharge was considered epileptiform if it was a polyspiking event comprising at least 3 spikes with a minimum amplitude of three times the baseline amplitude and a duration of at least 100 ms. Data are expressed as mean  $\pm$  SD.

### **Mouse 6-Hz (44 mA) psychomotor seizure model**

Experimental procedure was described before<sup>19, 45, 51</sup>. In brief, NMRI mice (average weight 28 g, range 23-32 g) were randomly divided into control and treatment groups (n = 5-10). 50  $\mu$ L (injection volume was adjusted to the individual weight) of VHC (PEG200:DMSO:water; 0.25:0.25:0.5) or treatment (an ASD or test compound dissolved in VHC) was i.p. injected in mice and after 30 or 120 min (in case of phenytoin, as reported by Barton and colleagues<sup>41</sup>) psychomotor seizures were induced by low frequency, long duration corneal electrical stimulation (6 Hz, 0.2 ms, rectangular pulse width, 3 s duration, 44 mA) using an ECT Unit 5780 (Ugo Basile, Comerio, Italy). Mice were manually restrained and a drop of ocular

anesthetic (0.5 % lidocaine) was applied to the corneas before stimulation. Following electrical current stimulation, the mouse was released in a transparent cage for behavioral observation, which was video-recorded. VHC-treated mice typically displayed stun, twitching of the vibrissae, forelimb clonus, and Straub tail. In addition, facial and mouth jerking as well as head nodding were observed occasionally. Seizure durations were measured during the experiment by experienced researchers, familiar with the different seizure behaviors. In addition, seizure durations were determined by blinded video analysis to confirm or correct the initial observations. Data are expressed as mean  $\pm$  SD.

## **ADMET profiling**

### *Cell viability and MTT assays*

Experimental procedure was described before<sup>54</sup>. Three cell lines were used: a) Hep G2 (HB-8065), a well-differentiated human hepatocellular carcinoma cell line, b) THLE-2, human liver epithelial cells transformed with SV40 large T-antigen and c) SHSY5Y, a thrice-cloned sub-line of a human metastatic bone tumor. Cells were seeded at a concentration of  $1 \times 10^4$  cells/well in 200  $\mu$ L culture medium and incubated at 37 °C in 5 % CO<sub>2</sub> using 96-well plates for 24 h. Next, the medium was replaced with medium complemented with test compounds at different concentrations. After another 24 h incubation, the medium was replaced by 100  $\mu$ L of a MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) solution (5 mg/mL in PBS and diluted at 0.5 mg/mL in MEM without phenol red). The plates were gently shaken and incubated for 3 h at 37 °C in a 5 % CO<sub>2</sub> incubator. The supernatant was removed and 100  $\mu$ L of 100 % DMSO was added. The plates were gently shaken to solubilize the formed formazan. The absorbance was measured using a multireader (Victor2 (Wallac)) at a wavelength of 570 nm.

### *Cardiotoxicity*

Fluorescence-based assays were performed using HEK293 cell lines that stably express the Nav1.5-channel, Cav1.2-channel, or hERG K<sup>+</sup>-channel, using FMP Red Dye (Molecular Devices), calcium-sensitive fluorescent dye Fluo-4 (Invitrogen) and FluxOR™ reagent (Invitrogen), respectively, and a FLIPR Tetra High-Throughput Cellular Screening System (Molecular Devices), according to manufacturer's protocols (Molecular Devices). Tetrodotoxin, israpidine and nicarbidine, and astemizole and haloperidol were used as standard

sodium channel, calcium channel, and hERG channel blockers. Data were analyzed using Genedata Screener.

#### *CYP450 enzyme inhibition assay*

Experimental procedure was described before<sup>55</sup>. Assessment of CYP450 inhibition was conducted in 96-well plate format at 37 °C. Test compounds were dissolved in DMSO/acetonitrile (ACN) (2 µL) and diluted in 98 µL NADPH solution (2 mM). Reactions were started by addition of 100 µL potassium phosphate buffer (200 mM, pH 7.4) containing Human Liver Microsomes (HLM) (0.5 mg/mL). Probe reactions for CYP3A4, CYP2D6 and CYP2C9 were conducted with 50 µM testosterone, 22 µM dextromethorphan and 10 µM diclofenac for 15 min. Reactions were terminated with the addition of a quench solution (90 µL) of ACN containing internal standards for LC-MS/MS determination (60 ppb cortisone, 100 ppb 4'-hydroxydiclofenac-<sup>13</sup>C<sub>6</sub>, 60 ppb levallorphan).

#### *Metabolic stability assay*

The assay was performed with a mixture of test compounds (1 µM), NADPH (4 mM) and HLM (1 mg/mL) incubated at 37 °C. Reactions were quenched at 0, 15, 30, 45, 60, and 90 min, using an equal volume of ACN and then diluted 1:1 with water prior to analysis by LC-MS/MS.

The analysis was performed using an Agilent Series 1290 LC system (Agilent Technologies, Santa Clara, CA, USA) using a Supelco Discovery HS C<sub>18</sub> (2.1×50 mm) 3 µm column that was held at 30 °C. Solvent A contained water with 0.1 % formic acid and solvent B contained ACN with 0.1 % formic acid, and the flow rate was set at 400 µL/min. The gradient elution was performed as follows: 0-0.5 min 0 % eluent B; 0.5-7 min 100 % eluent B; 7-9 min 100 % eluent B; 9-9.2 min 0 % eluent B; and 9.2-10.5 min 0 % eluent B. An API 4000 mass spectrometer in positive ESI mode (AB SCIEX, Concord, ON, Canada) was used with a generic method for data acquisition on all compounds. Data processing was performed using MultiQuant Software (AB SCIEX, Concord, ON, Canada) to process the data. Peak areas were used to plot the Ln % remaining relative to time (t) = 0. The slope of the natural log of the percent remaining versus time was calculated to determine the first-order rate constant (k) and the half-life (t<sub>1/2</sub>) of the test compounds according to the following equation: t<sub>1/2</sub> = 0.693/k (min)

### *Kinetic solubility assay*

Experimental procedure was described before<sup>56</sup>. The kinetic solubility assay was conducted in 96-well, flat-bottom, transparent polystyrene plates (Costar 9018, Corning, Tewksbury MA). Six two-fold serial dilutions of an initial 10 mM test compound solution were prepared in DMSO. After a 2-h incubation period (to avoid missing slow precipitation) absorbance was measured at 620 nm by an EnVision multilabel plate reader. The kinetic solubility was estimated from the concentration of test compound that produced an increase in absorbance above the background levels (i.e., 1 % DMSO in buffer).

### *Plasma protein binding assay*

Experimental procedure was described before<sup>57</sup>. Rapid equilibrium dialysis was performed with RED device inserts (Thermo Scientific, Meridian Rd., Rockford, IL) containing dialysis membrane with a molecular weight cut-off of 8000 Daltons. Serum (200  $\mu$ L) containing test compound (5  $\mu$ M) was added to the serum chamber of the insert and 350  $\mu$ L of buffer was added to the buffer chamber of the insert. Dialysis was done at 37 °C with shaking at 100 rpm for 5 h. Following dialysis, an aliquot of 50  $\mu$ L was removed from each well (plasma and buffer side) and diluted with an equal volume of opposite matrix to nullify the matrix effect. Then a fraction (50  $\mu$ L) of each dialyzed sample was crashed with 150  $\mu$ L of ACN containing internal standard and vortexed for 1 min. The samples were centrifuged at 13,300 rpm at 4 °C for 12 min and 100  $\mu$ L of supernatant was used for LC-MS/MS analysis.

### *Parallel artificial membrane permeability assay (PAMPA)*

The Gentest Pre-coated PAMPA Plate System (Corning) was used to perform the permeability assays. The 96-well filter plate, pre-coated with lipids, was used as the permeation acceptor and a matching 96-well receiver plate was used as the permeation donor. Compound solutions were prepared by diluting 10 mM DMSO stock solutions in PBS with a final concentration of 10  $\mu$ M. The compound solutions were added to the wells (300  $\mu$ L/well) of the receiver plate and PBS was added to the wells (200  $\mu$ L/well) of the pre-coated filter plate. The filter plate was then coupled with the receiver plate and the plate assembly was incubated at room temperature without agitation for 5 h. Next, the plates were separated and 50  $\mu$ L solution from each well of both the filter plate and the receiver plate was transferred to a vial with 150  $\mu$ L ACN and centrifuged at 13,300 rpm for 10 min at 4 °C. The supernatant was diluted in a solution



water/ACN (50/50). The final concentration of compounds in both donor wells and acceptor wells was analyzed by LC-MS/MS. Permeability of the compounds was calculated using the following equation:

$$\text{Permeability (cm/s): } Pe = \{-\ln[1-CA(t)/Ceq]\}/[A*(1/VD+1/VA)*t]$$

A = filter area (0.3 cm<sup>2</sup>), VD = donor well volume (0.3 mL), VA = acceptor well volume (0.2 mL), t = incubation time (s), CA(t) = compound concentration in acceptor well at time t, CD(t) = compound concentration in donor well at time t, and Ceq = (CD(t)\*VD+CA(t)\*VA)/(VD+VA)

## **ABBREVIATIONS**

ACN, acetonitrile; ACSF, artificial cerebrospinal fluid; ASD, antiseizure drug; dpf, days post-fertilization; DMSO, dimethyl sulfoxide; FP7, Seventh Framework Programme; h, hours; HLM, Human Liver Microsomes; LFP, local field potential; min, minute; MTC, maximum tolerated concentration; PEG200, poly-ethylene glycol M.W. 200; PTZ, pentylenetetrazole; s, seconds; t<sub>1/2</sub>, half-life; VHC, vehicle

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### **Author contributions**

PAMW, MJ, DC, and MR were responsible for study design. PAMW, MJ, DC, MR, FR, and RE were responsible for experimental design. Experiments were performed by DC, MR, JNT, MPB, ND, RV, CD, JP del P, and AJS. DC and MR were responsible for data acquisition and

analysis. DC, MR, and PAMW wrote the manuscript. DC prepared the figures and tables. All authors edited and approved the final version of the manuscript.

### **Funding sources**

This work was supported by the PharmaSea project ([www.pharma-sea.eu](http://www.pharma-sea.eu)), funded by the EU Seventh Framework Programme (contract number 312184).

### **Conflict of interest**

There is no potential conflict of interest.

### **ACKNOWLEDGEMENT**

This work was supported by the PharmaSea project ([www.pharma-sea.eu](http://www.pharma-sea.eu)), funded by the EU Seventh Framework Programme (contract number 312184). We would like to acknowledge Dr. Francisca Vicente, Dr. Carmen Ramos, and Dr. Bastien Cautain for their contribution with regards to the ADMET analysis. Concerning the graphical table of contents, we acknowledge somersault18:24 BVBA ([www.somersault1824.com](http://www.somersault1824.com)) for providing scientific illustrations.

### **SUPPORTING INFORMATION**

1D NMR data of pseurotin A, pseurotin A<sub>2</sub>, pseurotin F1, 11-*O*-methylpseurotin A, pseurotin D, and azaspirofurans A and B.

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## FIGURE LEGENDS

**Figure 1. Chemical structures isolated from extracts of the marine fungus *Aspergillus fumigatus*.**

**Figure 2. Behavioral antiseizure analysis in the zebrafish PTZ seizure model.** Antiseizure activity analysis of compounds at their maximum tolerated concentrations (MTC, Table 1) in the zebrafish pentylenetetrazole (PTZ) seizure model after 2 h (A) and 18 h (B) of incubation. PTZ-induced seizure-like behavior is expressed as mean actinteg units/5 min ( $\pm$  SD) during the 30-min recording period. Number of replicate wells per condition:  $n = 48$  (A) and  $n = 70-71$  (B) for VHC + PTZ and VHC + VHC controls, and  $n = 8-12$  (A) and  $n = 9-14$  (B) for compound + PTZ conditions. Data are pooled from multiple experimental plates. Statistical analysis: one-way ANOVA with Dunnett's multiple comparison test (GraphPad Prism 5). Significance levels: \*  $p \leq 0.05$ ; \*\*  $p \leq 0.01$ ; \*\*\*  $p \leq 0.001$ .

**Figure 3. Behavioral antiseizure analysis of azaspirofurans A and pseurotin A<sub>2</sub> in the zebrafish PTZ seizure model.** Antiseizure activity of azaspirofurans A (A-B) and pseurotin A<sub>2</sub> (C-D) in the zebrafish pentylenetetrazole (PTZ) seizure model after 2 h (A-B) and 18 h (C-D) of incubation, respectively. PTZ-induced seizure-like behavior is expressed as mean actinteg units/5 min ( $\pm$  SEM) during the 30-min recording period (A, C) and over 5-min time intervals (B, D). Means are pooled from three independent experiments with each 9-12 replicate wells per condition. Statistical analysis: (A, C) one-way ANOVA with Dunnett's multiple comparison test, (B, D) two-way ANOVA with Bonferroni posttests (GraphPad Prism 5). Significance levels: \*  $p \leq 0.05$ ; \*\*  $p \leq 0.01$ ; \*\*\*  $p \leq 0.001$ . Abbreviation: VHC, vehicle.

**Figure 4. Electrophysiological antiseizure analysis of azaspirofurans A and pseurotin A<sub>2</sub> in the zebrafish PTZ seizure model.** Noninvasive local field potential recordings from the optic tectum of larvae pre-exposed to vehicle (VHC) and pentylenetetrazole (PTZ), VHC only, compound and PTZ, or compound and VHC. (A-C) Larvae were incubated with 12.5  $\mu\text{g/mL}$  azaspirofurans A for 2 h, (D-F) or with 12.5  $\mu\text{g/mL}$  pseurotin A<sub>2</sub> for 18 h, conform with the optimal condition used in the behavioral assay. (A, D) Larvae are considered to possess epileptiform brain activity when three or more events occurred during a 10-min recording. Epileptiform discharges are quantified by the number (mean  $\pm$  SD) (B, E) and cumulative



duration (mean  $\pm$  SD) (C, F) of events per 10-min recording. Number of replicate wells per condition: n = 33 (A-C) and n = 22 (D-F) for VHC + PTZ controls, n = 31 (A-C) and n = 21 (D-F) for VHC + VHC controls, n = 15 (A-C) and n = 19 (D-F) for compound + PTZ conditions, and n = 15 (A-C) and n = 19 (D-F) for compound + VHC conditions. Statistical analysis: (A, D) Fisher's exact test with Bonferroni posttest, (B-C, E-F) Kruskal-Wallis test with Dunn's multiple comparison test (GraphPad Prism 5). Significance levels: \*  $p \leq 0.05$ ; \*\*  $p \leq 0.01$ ; \*\*\*  $p \leq 0.001$ .

**Figure 5. Representative local field potential recordings.** 10-min noninvasive local field potential recordings from the optic tectum of larvae pre-exposed to vehicle (VHC) and pentylenetetrazole (PTZ), VHC only, compound and PTZ, or compound and VHC. Larvae were incubated with 12.5  $\mu\text{g/mL}$  azaspirofurane A for 2 h or 12.5  $\mu\text{g/mL}$  pseurotin A<sub>2</sub> for 18 h, conform with the optimal condition used in the behavioral assay.

**Figure 6. Antiseizure activity analysis of azaspirofurane A and pseurotin A<sub>2</sub> in the mouse 6-Hz (44 mA) psychomotor seizure model.** Psychomotor seizures were electrically induced 30 min after i.p. injection of vehicle (VHC, n = 10), positive control valproate (n = 6), or test compound (n = 6-7), and 120 min after i.p. injection of negative control phenytoin (n = 5). Seizures are quantified by the duration (mean  $\pm$  SD). Statistical analysis: one-way ANOVA with Dunnett's multiple comparison test (GraphPad Prism 5). Significance levels: \*  $p \leq 0.05$ ; \*\*  $p \leq 0.01$ ; \*\*\*  $p \leq 0.001$ .

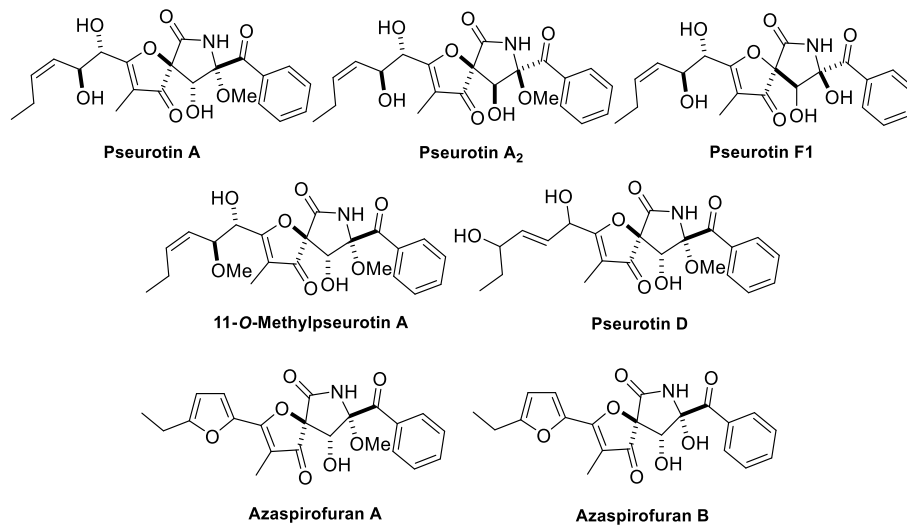
## TABLE LEGENDS

**Table 1. Maximum tolerated concentrations (MTCs) of test compounds in 7-dpf zebrafish larvae.** In case no MTC was reached, 100  $\mu\text{g/mL}$  was used as the test concentration.

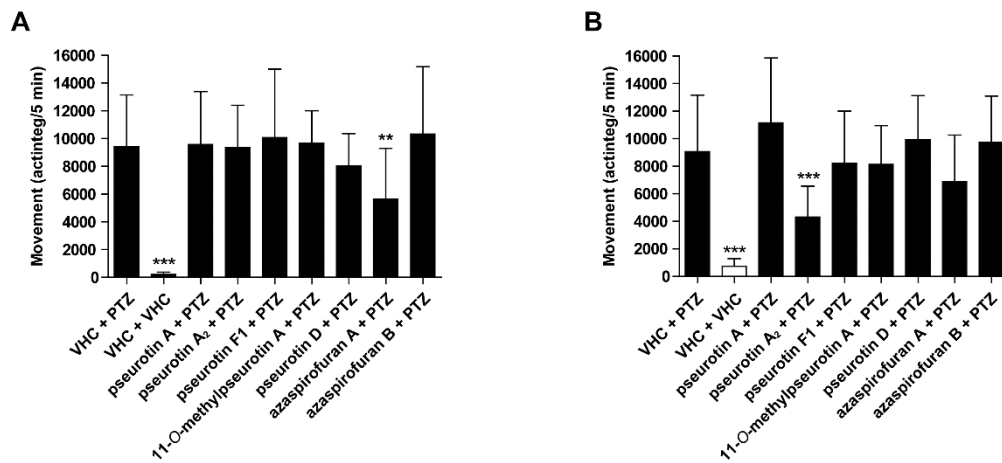
**Table 2. ADMET analysis of azaspirofurane A and pseurotin A<sub>2</sub>.** Abbreviations: concentration at which an assay is inhibited by 50 %, IC<sub>50</sub>.

## FIGURES

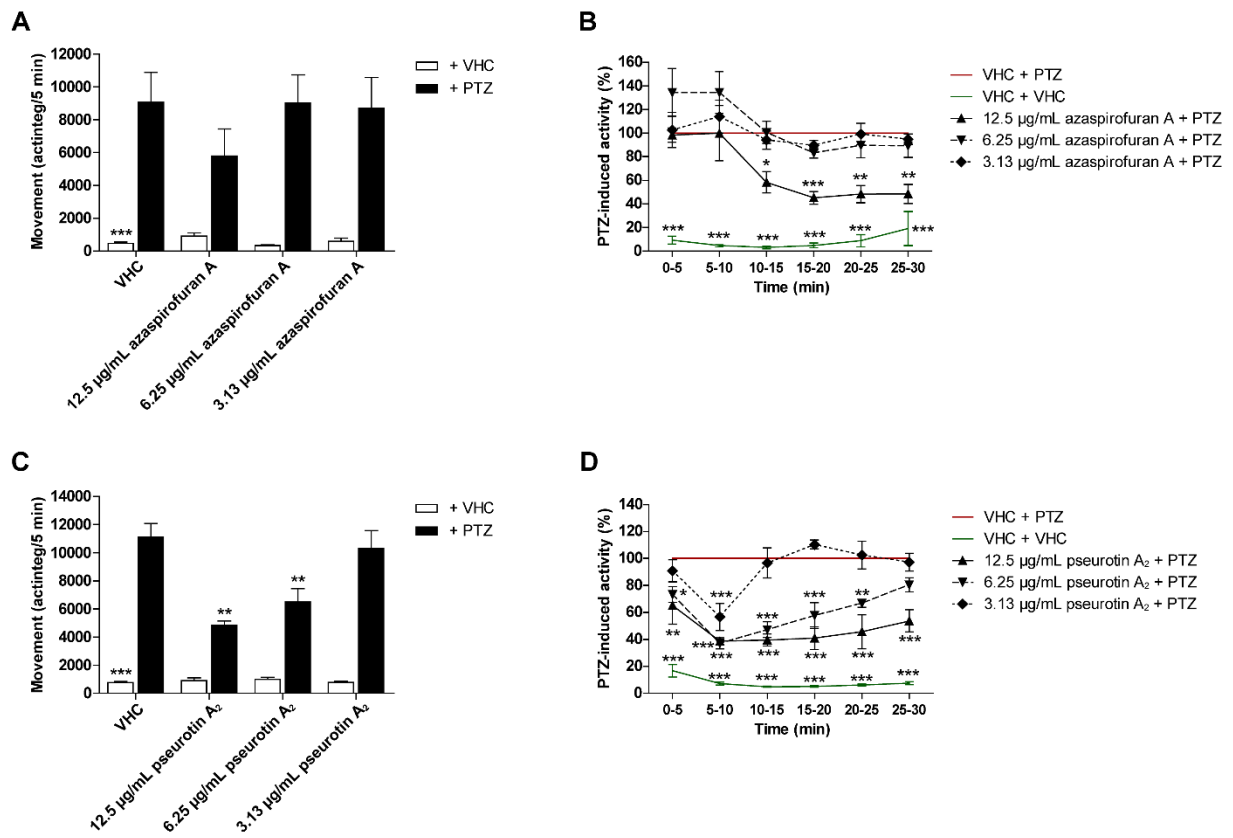
**Figure 1.**



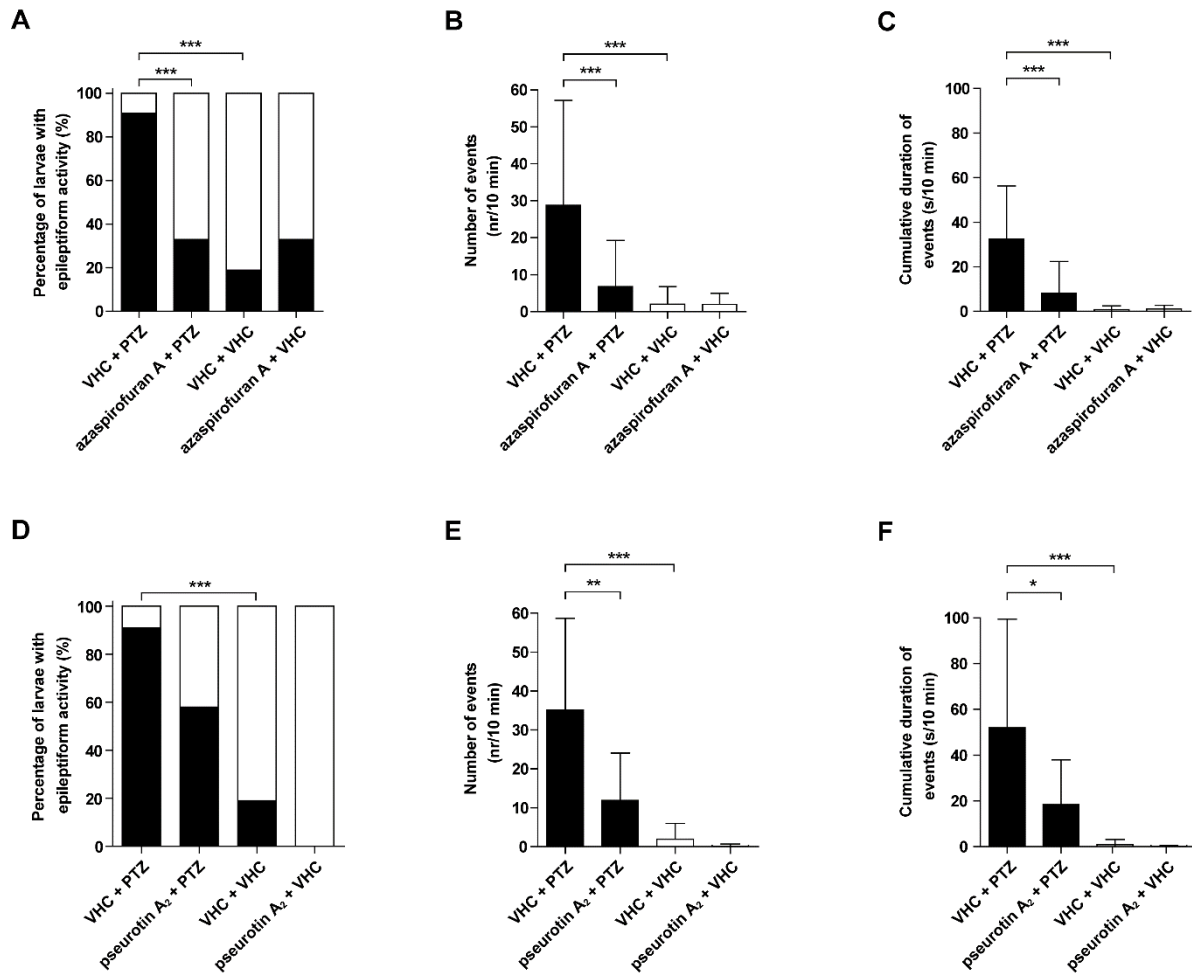
**Figure 2.**



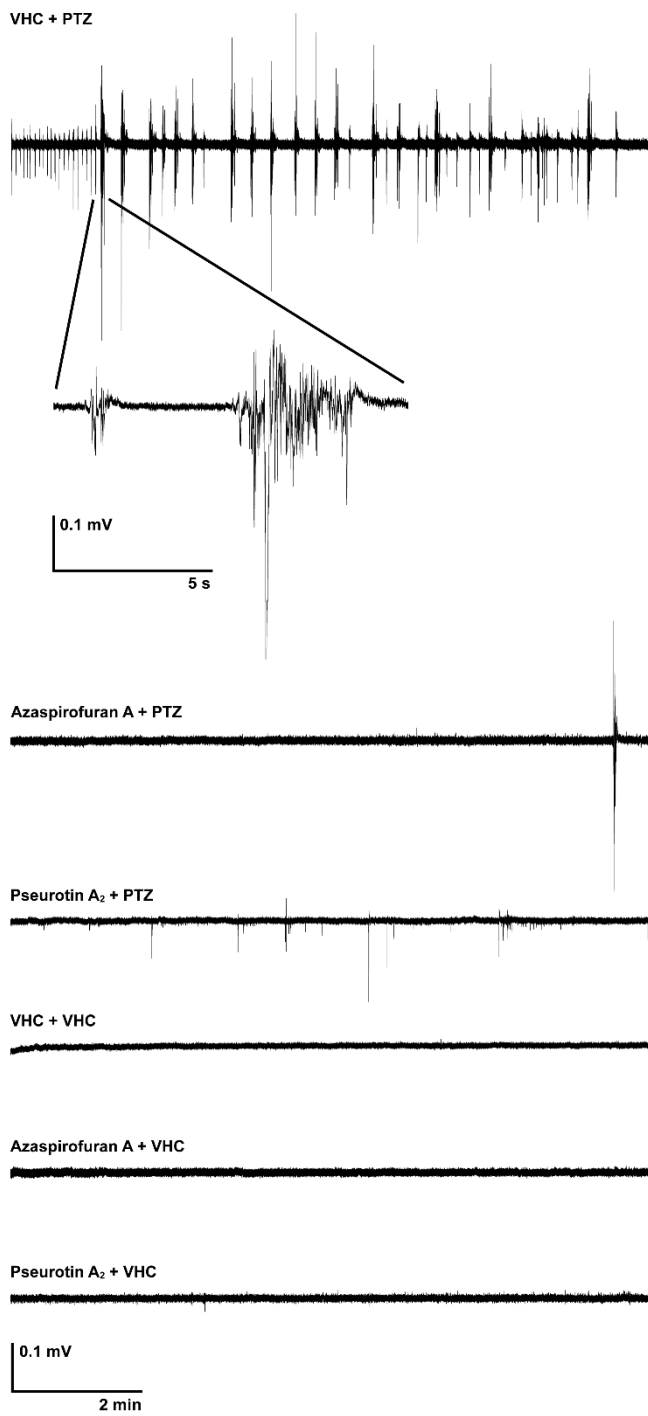
**Figure 3.**



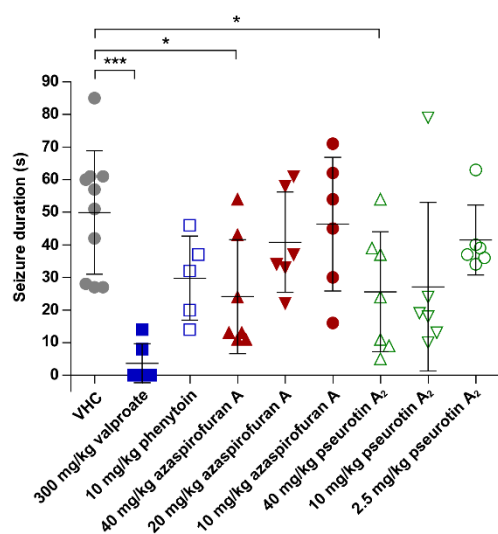
**Figure 4.**



**Figure 5.**



**Figure 6.**



**TABLES**

**Table 1.**

Compound	MTC (µg/mL)
Pseurotin A	100
Pseurotin A <sub>2</sub>	12.5
Pseurotin F1	50
11- <i>O</i> -methylpseurotin A	100
Pseurotin D	100
Azaspirofurans A	12.5
Aaspirofurans B	12.5

**Table 2.**

<b>ADMET test</b>	<b>Azapirofuran A</b>		<b>Pseurotin A<sub>2</sub></b>	
<b>Cytotoxicity</b>				
<b>HEPG2 cells</b>	IC <sub>50</sub> > 50 μM	No effect	IC <sub>50</sub> > 50 μM	No effect
<b>THLE2 cells</b>	IC <sub>50</sub> > 50 μM	Weak decrease	IC <sub>50</sub> > 50 μM	Weak decrease
<b>SHSY5Y cells</b>	IC <sub>50</sub> > 50 μM	Weak decrease	IC <sub>50</sub> = 17.9 μM	Decrease
<b>Cardiotoxicity</b>				
<b>Na<sub>v</sub>1.5 channel</b>	IC <sub>50</sub> = 39.06 μM	Low inhibitory effect	IC <sub>50</sub> > 50 μM	No effect
<b>Ca<sub>v</sub>1.2 channel</b>	IC <sub>50</sub> > 50 μM	No effect	IC <sub>50</sub> > 50 μM	No effect
<b>hERG channel</b>	IC <sub>50</sub> > 50 μM	No effect	IC <sub>50</sub> > 50 μM	No effect
<b>CYP450 enzymes</b>				
<b>CYP3A4</b>	IC <sub>50</sub> > 88 μM	No effect	IC <sub>50</sub> > 88 μM	No effect
<b>CYP2D6</b>	IC <sub>50</sub> = 23.4 μM	Weak inhibition	IC <sub>50</sub> > 88 μM	No effect
<b>CYP2C9</b>	IC <sub>50</sub> = 47.3 μM	Weak inhibition	IC <sub>50</sub> > 88 μM	No effect
<b>Other factors</b>				
<b>Hepatic clearance</b>	20.17 μL/min/mg protein (t <sub>1/2</sub> = 33.12 min)	Medium	< 8.6 μL/min/mg protein (t <sub>1/2</sub> > 60 min)	Low
<b>Kinetic solubility</b>	>100 μM	Acceptable	>100 μM	Acceptable
<b>Protein binding</b>	95.42 %	Low percentage of free drug; 66.7 % recovery	36.53 %	High percentage of free drug; 100 % recovery
<b>Permeability</b>	18.64x10 <sup>-6</sup> cm s <sup>-1</sup>	High	0.05x10 <sup>-6</sup> cm s <sup>-1</sup>	Low

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### Manuscript title

Zebrafish-based discovery of antiseizure compounds from the Red Sea: pseurotin A<sub>2</sub> and azaspirofuran A

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