

Central Lancashire Online Knowledge (CLoK)

Title	Orientation and location of the finite helical axis of the equine forelimb joints
Type	Article
URL	https://clock.uclan.ac.uk/28029/
DOI	https://doi.org/10.1002/jmor.20978
Date	2019
Citation	Kaashoek, Mariëlle, Hobbs, Sarah Jane, Clayton, Hilary Mary, Aerts, Peter and Nauwelaerts, Sandra (2019) Orientation and location of the finite helical axis of the equine forelimb joints. <i>Journal of Morphology</i> , 280 (5). pp. 712-721. ISSN 0362-2525
Creators	Kaashoek, Mariëlle, Hobbs, Sarah Jane, Clayton, Hilary Mary, Aerts, Peter and Nauwelaerts, Sandra

It is advisable to refer to the publisher's version if you intend to cite from the work.
<https://doi.org/10.1002/jmor.20978>

For information about Research at UCLan please go to <http://www.uclan.ac.uk/research/>

All outputs in CLoK are protected by Intellectual Property Rights law, including Copyright law. Copyright, IPR and Moral Rights for the works on this site are retained by the individual authors and/or other copyright owners. Terms and conditions for use of this material are defined in the <http://clock.uclan.ac.uk/policies/>

1 Orientation and location of the finite helical axis of the equine
2 forelimb joints

3

4 Helical axis of the equine forelimb joints

5

6 Mariëlle Kaashoek¹, Sarah Jane Hobbs², Hilary Mary Clayton³, Peter Aerts^{1,4} and Sandra
7 Nauwelaerts^{1,5}

8

9 1) Functional Morphology lab, Biology Department, University of Antwerp, Campus Drie
10 Eiken, Building D, Belgium

11 2) Centre for Applied Sport and Exercise Sciences, University of Central Lancashire,
12 Preston, Lancashire, United Kingdom

13 3) Department of Large Animal Clinical Sciences, Michigan State University, East Lansing,
14 MI, United States of America

15 4) Department of Movement and Sports Sciences, University of Ghent, Belgium

16 5) Centre for Research and Conservation Antwerp Zoo, Antwerp, Belgium

17

18 **Corresponding author:**

19 Mariëlle Kaashoek

20 Mariëlle.kaashoek@uantwerpen.be

21 Functional Morphology lab, Biology Department, University of Antwerp, Campus Drie

22 Eiken, Building D, Belgium Universiteitsplein 1, 2610 Wilrijk, Belgium

23

24 Abstract

25 To reduce anatomically unrealistic limb postures in a virtual musculoskeletal model of a
26 horse's forelimb, accurate knowledge on forelimb joint constraints is essential. The aim
27 of this cadaver study is to report all orientation and position changes of the finite helical
28 axes (FHA) as a function of joint angle for different equine forelimb joints. Five horse
29 cadaver forelimbs with standardized cuts at the midlevel of each segment were used.
30 Bone pins with reflective marker triads were drilled into the forelimb bones. Unless joint
31 angles were anatomically coupled, each joint was manually moved independently in all
32 three rotational DOFs (flexion-extension, abduction-adduction, internal-external
33 rotation). The three-dimensional coordinates of the marker triads were recorded using
34 a six infra-red camera system. The FHA and its orientational and positional properties
35 were calculated and expressed against joint angle over the entire range of motion using
36 a finite helical axis method. When coupled, joint angles and FHA were expressed in
37 function of flexion-extension angle. Flexion-extension movement was substantial in all
38 forelimb joints, the shoulder allowed additional considerable motion in all three
39 rotational DOFs. The position of the FHA was constant in the fetlock and a constant
40 orientation of the FHA was found in the shoulder. Orientation and position changes of
41 the FHA over the entire ROM were observed in the elbow, carpus and PIP-DIP joints. We
42 report FHA position and orientation changes as a function of flexion-extension angle to
43 allow for inclusion in a musculoskeletal model of a horse to minimize calculation errors
44 caused by incorrect location of the FHA.

45

46

47 **Keywords:** Musculoskeletal Model, Range of Motion, Helical Axis,

48

49

50 **Research Highlights and Graphical Abstract**

51 When measuring the finite helical axes of the equine forelimb joints over their entire
52 range of motion, changes in orientation were observed in the elbow, carpus, fetlock
53 and PIP-DIP joints. Changes in position were measured in the shoulder, elbow, carpus
54 and PIP-DIP joints.

55

56 **Main Text**

57 **Introduction**

58 A common way to study the locomotion of an animal is conducting *in vivo* gait
59 experiments where external force and segment/joint movement patterns are typically
60 reported. The underlying mechanisms to generate locomotion, such as muscle control,
61 internal loading of anatomical structures, joint forces and muscle energetics are more
62 difficult to obtain using non-invasive techniques (Umberger & Caldwell, 2013). One
63 solution is the use of musculoskeletal models, which are three-dimensional virtual
64 reconstructions of the musculoskeletal system that can estimate these internal variables
65 (Umberger & Caldwell, 2013). Such models use a linked segment approach of rigid
66 bodies (bones) that connect at joints, and may include muscles, ligaments and other
67 structures (Delp & Loan, 1995; Seth, Sherman, Reinbolt, & Delp, 2011).

68

69 The three-dimensional motion of two adjacent segments can be described by defining
70 their finite helical axis (FHA) (SHIAVI et al 1987), which describes the motion of two
71 segments as a rotation about and a translation along an axis (Blankevoort et al 1990). In
72 the models, joint constraints like the FHA are used to eliminate kinematics which are
73 outside the range of natural poses or movements (Kambic, Roberts, & Gatesy, 2017).

74

75 Different methods can be used to determine the axis of rotation of a joint, for example
76 the symmetrical axis of rotation approach, finite helical axis and instantaneous helical
77 axis methods, the latter two are amongst the most commonly used methods (Ehrig &
78 Heller, 2019). In current study, the finite helical axis (FHA) was used to calculate the
79 joint axes, this method uses three-dimensional motion data of two adjacent segments
80 to determine the position and orientation of the FHA (Ehrig & Heller, 2019; Shiavi et al.,
81 1987). A clear progression of the FHA in a joint can be obtained by dividing the entire
82 ROM into multiple windows and calculating the FHA over each window (Blankevoort,
83 Huiskes, & de Lange, 1990; Spoor & Veldpaus, 1980; van den Bogert, Reinschmidt, &
84 Lundberg, 2008). Throughout movement, the FHA can shift and/or change in
85 orientation, for instance in the human knee, where the position of the FHA changes
86 during flexion (Blankevoort et al., 1990; van den Bogert et al., 2008). Orientation
87 changes of the FHA occur when a joint rotates about two or three of the orthogonal axes
88 simultaneously. Translations of the FHA can be observed when one bone slides along
89 one or more of the three orthogonal axes.

90

91 An accurate location of the three-dimensional FHA is important when building
92 musculoskeletal models (Bru & Pasqui, 2010). The kinematic and kinetic output of the
93 model are directly affects when the position and/or orientation of the modelled joint
94 axis deviates from the actual joint axis (Holden & Stanhope, 1998; Stagni, Leardini,
95 Cappozzo, Benedetti, & Cappello, 2000). Furthermore, errors will be caused in the
96 calculation of moment arms and joint moments (Camomilla et al., 2017). So far, forelimb
97 joints of the equine athlete have been modelled using one degree of freedom joint
98 descriptions. Previous models of Brown et al. (2003) and Swanstrom et al. (2005)
99 describe the translations of the joint axes in craniocaudal/dorsopalmar and
100 proximodistal as a function of flexion-extension angle. In the current study, we expand
101 on these data by measuring multiple horses and adding the orientation properties of the
102 FHA over the entire ROM for all equine forelimb joints to the description of the
103 craniocaudal/dorsopalmar and proximodistal position of the FHA using a helical axis
104 approach. The three-dimensional data of the FHA are reported as a function of joint
105 angle and can be used when constructing a musculoskeletal model of the horse's
106 forelimb.

107

108 **Material and Methods**

109 **Subjects**

110 Five horse cadavers (age: 16.75 ± 1.35 years, weight 532 ± 6.58 kg, varying breeds) were
111 obtained from the pathology department at Ghent University, where the experiments
112 were performed between April 2016 and November 2018. A formal ethical approval was
113 waived by the chairperson of the ethical committee, based on Belgian and European

114 legislation (EU directive 2010/63/EU), as all tissues were derived post mortem from the
115 necropsy room or from a commercial abattoir. The horses included in this study did not
116 show any signs of musculoskeletal injuries and either died of natural causes or were
117 euthanized for non-locomotor issues. The cadavers were stored in a cooler at 4 degrees
118 Celsius and experiments were performed approximately three days post mortem.

119

120 **Preparation of Specimens**

121 Each left forelimb was removed from the trunk by cutting the soft tissues between the
122 scapula and the rib cage. Six forelimb segments were defined, from proximal to distal,
123 the shoulder, brachium, antebrachium, metacarpus, pastern and hoof (Fig 1). To ensure
124 that all anatomically possible positions and orientations of the distal segment with
125 respect to the proximal segment in each joint were obtained and to maximize the ROM,
126 standardized cuts through the soft tissue were made midway along the length of each
127 forelimb segment to eliminate muscle, tendon, fascia and skin stiffness (Fig 1). Joint
128 capsules, tendon attachment sites and ligaments surrounding the joint were kept intact.

129

130 A bone-pin was drilled into the shaft of the main bone of each segment: scapula
131 (shoulder), humerus (brachium), fused radius and ulna (radio-ulna) (antebrachium),
132 third metacarpal bone (metacarpus), proximal phalanx (pastern) and distal phalanx via
133 the hoof wall (PIP-DIP joints) (Fig 1). Reflective marker triads with a marker diameter of
134 15 mm, were attached to the bone pins. The secure attachment of the marker triads
135 ensured that they represented the exact movements of the bones to which they were
136 attached. The joints were, from proximal to distal, the shoulder, elbow, carpus, fetlock

137 and PIP-DIP joints, which included the proximal and distal interphalangeal joints (Fig 1).

138 The FHA of the distal sesamoid bones were not tracked in this study.

139

140 **Dynamic Trials**

141 The limbs were either placed horizontal on a table with the lateral side facing upwards

142 or partly lifted from the table depending on the rotational degree of freedom (rDOF)

143 (flexion-extension, abduction-adduction, internal-external rotation) that was measured.

144 Dynamic trials were conducted where each joint was manually moved through its full

145 ROM in each of the three rDOFs separately. Each dynamic trial consisted of at least three

146 movement cycles in one of the three rDOFs for a specific joint. The abduction-adduction

147 and internal-external rotation trials were performed at one position of the flexion-

148 extension angle, except for the carpus which was measured in two positions, at

149 extension ($\sim 0^\circ$) and at flexion ($\sim 90^\circ$). During the manual movement of the joints the

150 segments were moved until the experimenter was not able to move the limb any further

151 in that direction due to bone-bone or muscle-skin contact. Out of plane motion was kept

152 to a minimum while still allowing motion in the other rDOF in case two or more rDOF

153 were coupled (Kambic et al., 2017). The three-dimensional positions of all markers were

154 recorded within a pre-calibrated field of view ($\sim 1.5\text{m} \times 1.5\text{m} \times 1\text{m}$, mean camera residual

155 ≤ 0.35) using a motion analysis system with six infra-red cameras recording at 60 Hz

156 (VICON, Los Angeles, CA, USA) and Vicon Workstation software.

157

158 **Defining the local coordinate systems**

159 After the dynamic trials, soft tissues were removed to expose the bones. Bones were
160 disarticulated, and four anatomical markers were placed at standardized locations on
161 the joint surfaces (Fig 2): the first two markers were placed at the centre of the proximal
162 and distal articular surfaces; the third and fourth markers were located in the middle of
163 the lateral side and in the middle of the **caudal/palmar** side of the distal articular surface.
164 A static recording was made that captured the position of the triad markers with respect
165 to the anatomical markers.

166

167 An anatomically relevant, right-handed local coordinate system (LCS) was defined for
168 each forelimb segment using the anatomical markers (Fig 2). The origin of the LCS was
169 placed in the centre of the distal articular surface using the first anatomical marker
170 (Grood & Suntay, 1983). The proximodistal-axis ran through the long axis of the bone
171 and was positive in the proximal direction. The craniocaudal/dorsopalmar-axis pointed
172 from the origin towards the caudal side of the segment. The mediolateral-axis ran
173 transversely and was positive toward the lateral side.

174

175 **Pre-Analysis**

176 The virtual marker coordinates from the dynamic trial and the static recording were
177 tracked using the Vicon Workstation software. The coordinate data were filtered using
178 a fourth-order Butterworth filter with a cut-off frequency of 5 Hz. The orientation
179 matrices and displacement vectors between the tracking markers of the static file and
180 the dynamic trials were calculated using singular value decomposition (Söderkvist &
181 Wedin, 1993). Coordinates of the anatomical markers of both segments were

182 transformed to the global coordinate system (GCS) of the dynamic trials. Calibration of
183 the Vicon camera system was performed using a T-wand and an L-frame, the position of
184 the latter was used to define the GCS. The LCS of the proximal (P-LCS) and distal (D-LCS)
185 segments were determined using the transformed anatomical marker coordinates. For
186 each frame, the P-LCS was translated to the origin of the GCS and rotated to align with
187 the orientation of the axes of the GCS. The D-LCS was expressed relative to the P-LCS.

188

189 **Analysis: Joint angles and Range of Motion**

190 Kinematic analyses were performed using the MATLAB (The MathWorks Inc, Natick,
191 Massachusetts) based software package KineMat (Reinschmidt & van den Bogert, 1997),
192 based on the work of Grood & Suntay (1983). The orientations of the D-LCS with respect
193 to the P-LCS were used to calculate the joint Cardan angles, which were defined by
194 rotations that occurred about three axes: the first rotation was flexion-extension about
195 the mediolateral-axis of the proximal segment, the second rotation was abduction-
196 adduction about the floating axis, which was the result of the cross product between
197 the mediolateral-axis of the proximal segment and the proximodistal-axis of the distal
198 segment. The third rotation was internal-external rotation about the proximodistal-axis
199 of the distal segment (Grood & Suntay, 1983; Zatsiorsky, 1997).

200

201 Data from the dynamic trials, for each type of rotation, were used to determine the
202 minimal and maximal joint angle and the ROM for each rDOF. Joint angles were
203 calculated using the neutral positions of the forelimb joints as reported in Weller et al
204 2006, (Fig. 1). During the quality control, trials were removed from the data set when

205 for example bone pins appeared to be loose, misplacement of the anatomical markers
206 led to untenable joint angles or when the out of plane motion showed large deviations
207 when testing for a certain rDOF.

208

209 **Analysis: Helical axis**

210 Data from the dynamic trials and the corresponding static recordings were used to
211 calculate the FHA. From each trial, one movement cycle was selected. The increasing
212 and decreasing joint angle phases of the movement cycle were divided into equal steps
213 of approximately 5 degrees. Using the KineMat software package (Reinschmidt & van
214 den Bogert, 1997), based on Spoor & Veldpaus (1980), FHA's were calculated for each
215 step (van den Bogert et al., 2008). A homogeneous transformation matrix (T) was
216 calculated between the transformed D-LCS of θ and $\theta \pm \sim 5$ degrees using singular value
217 decomposition. Properties of the FHA were extracted from T using the method of Spoor
218 and Veldpaus (1980) adapted by Reinschmidt and van den Bogert (1997).

219

220 Depending on the rDOF that was measured, the position of the FHA was calculated as
221 the distance between the origin and the intersection of the FHA with the plane
222 perpendicular to the FHA which was then projected on the associated P-LCS axis (Fig 3).
223 The planes used for the different trials were the sagittal plane for FE trials, the frontal
224 plane for AA trials and the transverse plane for IE trials (Fig 3A-B). The proximodistal
225 position of the FHA was the distance between the intersection of the FHA with
226 associated plane projected onto the proximodistal axis and the origin of the P-LCS (Fig
227 3AB). The distance between the intersection of the FHA with the associated plane

228 projected onto the craniocaudal/dorsopalmar axis and the origin of the P-LCS (Fig 3BC).
229 The medio-lateral distance was defined as the distance between the intersection of the
230 FHA and the associated plane projected onto the mediolateral axis and the origin of the
231 P-LCS (Fig 3AC).

232

233 The deviation angle of the FHA was the angle between the projection of the FHA onto
234 the transverse plane of the proximal segment and the mediolateral axis of the P-LCS (Fig
235 3D). The angle between the FHA projected onto the sagittal plane of the proximal
236 segment and the proximodistal axis was defined as the inclination angle (Fig 3E)
237 (Blankevoort et al., 1990).

238

239 **Statistical analysis**

240 The five variables describing the position and orientation of the FHA (inclination,
241 deviation, craniocaudal/dorsopalmar position, proximodistal position and mediolateral
242 position (Fig 3)) were analysed statistically using SPSS (SPSS version 24.0; SPSS Inc,
243 Chicago, Illinois). A multivariate analysis of covariance (MANCOVA) test was performed
244 on the set of five dependent variables, inclination angle, deviation angle,
245 craniocaudal/dorsopalmar position, proximodistal position and mediolateral position
246 for each joint with joint angle of the tested rDOF as a covariate and leg number
247 (individual) and direction of movement as fixed, independent factors. The latter was
248 added to test whether the FHA variables were influenced by the direction of movement.

249

250 To study the effect of joint angle on each of the FHA variables for each joint for the horse
251 as a species, leg number was removed from the MANCOVA and either a regression
252 equation or a mean value was calculated over all limbs for the FHA variable depending
253 on the outcome of the MANCOVA, Table 4-5. Mean values for FHA variables were
254 calculated over the entire ROM over all limbs when there only was a significant effect of
255 leg number (Table 4). If there was a significant effect of joint angle or for the interaction
256 effect between leg number and joint angle, i.e. the FHA changed with joint angle,
257 subsequent **reduced major axis regression analyses were performed using JMP (JMP®,**
258 **Version 13.2.1 SAS Institute Inc., Cary, NC, 1989-2019)** to determine the amount of
259 change of the variable over the entire ROM (Table 4-5).

260

261 **The interaction between rDOF can be a complex relationship in a three-dimensional**
262 **space, however for this study we only calculated the individual relationships between**
263 **the rDOF (Kambic et al., 2017).** Pearson correlation tests were performed on all three
264 rDOFs for each trial to test whether there was a coupling between the rDOFs, Table 6.
265 **An reduced major axis regression** was calculated if there was a significant correlation
266 between two rDOFs.

267

268 **Method validation**

269 Prior to conducting the experiments, the analysis script developed for this study and
270 experimental setup were validated using an artificial test joint with one rDOF (i.e. a door
271 hinge connecting two wooden segments). The location of the FHA should be stable and

272 running through the centre of the door hinge, indicated with extra markers, for the
273 experiment and analysis to be correct.

274

275 **Results**

276 **Joint angles and range of motion**

277 The results for the joint angles and ROM and the out of plane motion for each of the
278 forelimb joints are reported in Table 1-3. The variation in the ROM of the carpus, fetlock
279 and PIP-DIP joints were smaller compared to the variation of the shoulder and elbow. In
280 the distal joints, the endpoints were determined by bone to bone contact and in the
281 proximal joints the endpoints were influenced by the amount of muscle tissue
282 surrounding the joints. The ROM values reported in this study were aimed to be larger
283 than those reported in kinematic studies and exceeded the normal physiologic ranges
284 (Back et al., 1995).

285

286 **Finite helical axis**

287 The FHA for the artificial joint (i.e. a door hinge connecting two wooden segments) was
288 located, as expected, at the same location as the door hinge and did not show any
289 change in orientation or translation when changing the angle between the wooden
290 segments.

291

292 The FHA was calculated for all rDOFs with a ROM above 25 degrees in order to obtain a
293 clear progression of the FHA over the entire ROM. Results of the statistical tests
294 regarding leg number and joint angle are reported in Table 4-5. Locations of the FHA

295 were not calculated for the axis about which the rotation occurred. A significant effect
296 of the direction of movement was only found for the deviation angle when moving the
297 shoulder in flexion and extension ($P_{UD*JA} = 0.02$), the inclination angle for the internal
298 external rotation trials for the shoulder ($P_{UD*JA} = 0.02$), proximodistal position in the
299 flexion-extension trial of the carpus ($P_{UD*JA} = 0.00$, $P_{UD} = 0.00$) and for the inclination angle
300 while moving the fetlock in flexion-extension ($P_{UD*JA} = 0.03$) and for the dorsopalmar
301 position while moving the PIP-DIP in flexion-extension ($P_{UD*JA} = 0.03$).

302

303 Results of the orientation (inclination and deviation) and positional
304 (craniocaudal/dorsopalmar, proximodistal and mediolateral position) of the FHA over
305 the entire ROM are shown in Table 4. The position and orientation of the FHA remained
306 constant over the entire ROM when moving the shoulder through abduction-adduction
307 and internal-external rotation. FHA position and orientation reported below were
308 determined by calculating the difference between minimal and maximal joint angle
309 using the regression equations reported in Table 5. The FHA of the elbow and fetlock
310 showed a significant change in inclination (elbow = 46° , fetlock(up) = 71° fetlock(down)
311 = 56°) and deviation angle (fetlock = 6°), whereas the FHA position was constant for both
312 joints. When moving the shoulder in flexion-extension, only the orientation changed
313 significantly. Changes in proximodistal (up = 8mm, down = 26mm) and dorsopalmar (40
314 mm) position were found for the flexion-extension trials of the carpus, which also
315 showed change in inclination (-60°) and deviation (9°) angle. The PIP-DIP joints displayed
316 a change in inclination angle (-144°) and in proximodistal position (up = 3 mm, down =
317 8 mm)

318

319 No significant correlations were found between flexion-extension and internal-external
320 rotation in the shoulder, carpus and PIP-DIP joints and internal-external rotation and
321 flexion-extension for the shoulder, Table 6. Weak correlation (significant correlation
322 with a Pearsons correlation between 0.3-0.65) were found between both rDOF when
323 moving the shoulder in abduction-adduction and between flexion-extension and
324 abduction-adduction for the shoulder, elbow, carpus, fetlock and PIP-DIP joints, Table 6.
325 The fetlock also showed weak correlations between flexion-extension and internal-
326 external rotation. Internal-external rotation and abduction-adduction for the shoulder
327 and flexion-extension and internal-external rotation for the elbow showed a strong
328 correlation (significant correlation with a Pearsons correlation above 0.65). **Reduced**
329 **major axis regression equations describing the correlation** are reported in Table 6 which
330 can be used when modelling the forelimb joints in musculoskeletal models.

331

332 **Discussion**

333 In the current study, we describe the three-dimensional properties of the FHA over the
334 entire ROM for all forelimb joints as a function of joint angle. **As expected**, the shoulder
335 displayed substantial rotation in abduction-adduction and internal-external rotation **due**
336 **to its joint surface morphology**. For abduction-adduction the FHA of the shoulder
337 showed no orientational or positional changes, orientational changes were observed
338 when moving the shoulder in flexion-extension, orientational and position changes were
339 found for internal-external rotation. The FHA of the elbow and fetlock only showed
340 orientation changes. The carpus and PIP-DIP joint displayed both orientation and

341 position changes. Most of the joints also showed significant correlations between the
342 rDOFs except between flexion-extension and internal-external rotation for the shoulder.

343

344 Equine forelimb joints moved mainly in the parasagittal plane: only the shoulder allowed
345 substantial extra-sagittal motion. The shoulder is classified as an ellipsoidal ball and
346 socket joint (Budras, Sack, Röck, Horowitz, & Berg, 2012). Due to the elongated elliptic
347 shape of the glenoid cavity, the FHA translated significantly along the proximodistal axis
348 when moving the joint through flexion-extension. Translations of the FHA were not
349 found for the abduction-adduction trials. Previous models generally excluded the
350 shoulder (Brown, Pandy, Kawcak, & Mcilwraith, 2003; Michael D. Swanstrom, Zarucco,
351 Hubbard, Stover, & Hawkins, 2005), or only studied the motion of the shoulder within
352 the sagittal plane making it difficult to compare all our findings. Leach et al. (1988) did
353 found translations of the instant centre of rotation as a function of flexion-extension
354 angle, however these were in the craniocaudal direction.

355

356 In contrast with the shoulder, movements of the elbow are mainly restricted to the
357 parasagittal plane and are reflected in the morphology of their articular surfaces. In the
358 elbow, a groove running along the centre of the articular surface fits into a matching
359 ridge on the adjacent joint surface. These interlocking structures in combination with
360 collateral ligaments restrict the joints to parasagittal plane motion only (Ross & Dyson,
361 2011). The fetlock has a similar interlocking structure, however due to the shape of the
362 condyle of the fetlock, which is larger on the medial side compared to the lateral side,
363 there is more out-of-sagittal plane motion allowed compared to the elbow, which is also

364 shown in our results (Table 1-3). Both showed changes in inclination angle, the fetlock
365 also displayed a change in deviation angle when flexing the joint.

366

367 In the **PIP-DIP** joints, a saddle-like joint articulation morphology is found, allowing more
368 out of sagittal plane motion compared to the interlocking structures of the elbow
369 (Budras et al., 2012). This is also reflected in the larger out of sagittal plane ROM values
370 of the **PIP-DIP** joints compared to the elbow (Table1-3). When comparing our results
371 with three-dimensional kinematic studies, we found that the ROM values for the out of
372 sagittal plane motion for the **PIP-DIP** joints was larger than observed in *in vivo* gait
373 experiments (H. Chateau, Degueurce, & Denoix, 2006; Henry Chateau, Degueurce, &
374 Denoix, 2004; Clayton, Sha, Stick, & Robinson, 2007; Panagiotopoulou, Rankin, Gatesy,
375 & Hutchinson, 2016; Roach et al., 2014). However, these gait experiments were
376 performed on a relative flat surface and at relative low locomotion speeds. Even though
377 larger out of sagittal plane ROMs were measured, the FHA **PIP-DIP** joints did not show a
378 significant change in deviation angle and the proximodistal position.

379

380 The **PIP-DIP** joints, the proximal and distal interphalangeal joint, were measured
381 simultaneously because most gait analysis consider the first and second phalanx are a
382 single segment (Back et al., 1995; Khumsap, Clayton, Lanovaz, & Bouchey, 2002) and the
383 musculoskeletal models for which the FHA results are reported will be driven by
384 kinematic data. Previous detailed studies on the distal joints showed that there is a
385 relative small amount of motion occurring at the proximal interphalangeal joint (Henry
386 Chateau et al., 2004; Clayton et al., 2007) and translations of the distal interphalangeal

387 joint were ~1mm (Michael David Swanstrom, 1998), for models that require such
388 detailed data on the individual joints, **we suggest undertaking an three-dimensional X-**
389 **ray study or using prior XROMM data from which the FHA can be determined in more**
390 **detail for both joints individually (Panagiotopoulou et al., 2016; Roach et al., 2014).** The
391 same is suggested for the individual carpal bones, due to the complex movements of the
392 individual carpal bones (Yalden, 1971).

393

394 The carpus was measured as one entity although it technically also consists of multiple
395 joints. The radio-ulna is connected to the metacarpus III via two rows of carpal bones
396 resulting in three joints, from proximal to distal, the antebrachiocarpal, middle carpal
397 and carpometacarpal joints (Budras et al., 2012). From flexion to extension, the FHA
398 translates simultaneously in both a distal and a dorsal direction which could be caused
399 by the conformation of the articular surfaces of the distal end of the radius and the
400 proximal rows of carpal bones and by the increased separation of the two proximal
401 carpal joints on their dorsal side in flexion. Studies have shown limited movement at the
402 carpometacarpal joint and the distal row of carpal bones has been attached to the
403 proximal metacarpus in previous musculoskeletal models (Brown et al. 2003;
404 Swanstrom et al. 2005).

405

406 Significant inter-limb variation was found in all forelimb joints, which most likely is
407 partially due to the manual placement of the anatomical markers, which determines the
408 position of the LCS. Small differences in the orientation of the LCS will lead to over or
409 under estimation of the joint angles (Clayton, Sha, Stick, & Mullineaux, 2004) but can

410 also lead to variation in the position and orientation of the FHA. To obtain an accurate
411 model of a specific horse, ideally the horse's own FHA data should be used. However,
412 when building a generic horse model, this inter-limb variation is relatively small and can
413 be neglected, the regression equations mentioned in Table 4 can be used to define the
414 FHA in equine musculoskeletal models.

415

416 When comparing our results to previous reported instant centre of rotations, similar
417 locations were found for the elbow and fetlock (Leach & Dyson, 1988). Leach et al. 1988
418 also found a dorsopalmar displacement of the instant centre of rotation for the carpus.
419 Comparing our results to previously reported data of musculoskeletal models proved
420 difficult. Brown et al. (2003) does not provide enough detail on the locations of the
421 coordinate systems to directly compare the location and focuses more on the muscle
422 geometry. Some of our data contrasts with Swanstrom M.D (1998) probably caused by
423 the differences in the number of subjects and the approach: we measured the carpus as
424 one entity and Swanstrom separated the carpus in two parts (Leach & Dyson, 1988).
425 They also reported small translations in the fetlock which we were not able to detect.
426 These small translations possibly disappeared in our dataset of five horses, whereas the
427 Swanstrom (1998) had data for one horse, which did not allow for a statistical analysis.
428 Differences were possibly also due to the use of different methods, MRI, CT and
429 radiographs, versus bone pins. We also used a different calculation method, the helical
430 axis method. A direct comparison of the absolute values of the centre of rotation was
431 not possible because we used such different methods.

432

433 In conclusion, in this study, we report the three-dimensional behaviour of the FHA,
434 relative to the proximal segment of the different forelimb joints, as a function of the
435 flexion-extension angle. The findings of this study should be taken into account when
436 constructing a musculoskeletal model for an equine forelimb, however differences in
437 the definition of the local coordinate systems between the model and this study should
438 be taken into account.

439

440 **Author contributions**

441 Sarah Jane Hobbs and Hilary Mary Clayton conceived the original idea for this study and
442 the methodology, they also proofread the manuscript prior to submission. Sarah Jane
443 Hobbs also contributed to part of the analysis. Peter Aerts and Sandra Nauwelaerts
444 supervised the project and were involved in proofreading the manuscript. Sandra
445 Nauwelaerts also helped with the experiments, parts of the analysis and helped with the
446 writing. Mariëlle Kaashoek carried out the experiments, performed the analysis and
447 wrote the manuscript.

448

449 **Acknowledgements**

450 We would like to thank Koen Chiers, Leen Van Brantegem, Michiel Moors and H el ene
451 Claeys from the pathology department at Ghent University for their kind help during the
452 experiments. Big thanks to Jan Scholliers, Kristiaan D'Ao ut and the FunMorphers that
453 helped us. We would like to thank the owners for making this study possible. And finally,
454 we would like to thank the Bijzondere Onderzoeksfonds (BOF DOC PRO ID-31518) and
455 the University of Antwerp for funding this project.

456

457 **References**

- 458 Back, W., Schamhardt, H. C., Savelberg, H. H. C. M., Van den Bogert, A. J., Bruin, G.,
459 Hartman, W., & Barenveld, A. (1995). How the horse moves: 1. Significance of
460 graphical representations of equine forelimb kinematics. *Equine Veterinary*
461 *Journal*, *27*, 31–38.
- 462 Blankevoort, L., Huiskes, R., & de Lange, A. (1990). Helical axes of passive knee joint
463 motions. *Journal of Biomechanics*, *23*, 1219–1229.
- 464 Brown, N. A. T., Pandy, M. G., Kawcak, C. E., & McIlwraith, W. C. (2003). Force- and
465 moment-generating capacities of muscles in the distal forelimb of the horse.
466 *Journal of Anatomy*, 101–113.
- 467 Bru, B., & Pasqui, V. (2010). Localisation of the Instantaneous Axis of Rotation in
468 Human Joints. In *Advances in Robot Kinematics: Motion in Man and Machine* (pp.
469 195–196).
- 470 Budras, K.-D., Sack, W. O., Röck, S., Horowitz, A., & Berg, R. (2012). Chapter 2: Thoracic
471 Limb. In *Anatomy of the Horse* (pp. 4–15). London: Schluetersche.
- 472 Camomilla, V., Cereatti, A., Cutti, A. G., Fantozzi, S., Stagni, R., & Vannozzi, G. (2017).
473 Methodological factors affecting joint moments estimation in clinical gait analysis:
474 A systematic review. *BioMedical Engineering Online*, *16*, 1–27.
- 475 Chateau, H., Degueurce, C., & Denoix, J. M. (2004). Evaluation of three-dimensional
476 kinematics of the distal portion of the forelimb in horses walking in a straight line.
477 *American Journal of Veterinary Research*, *65*, 447–455.
- 478 Chateau, H., Degueurce, C., & Denoix, J. M. (2006). Effects of egg-bar shoes on the 3-
479 dimensional kinematics of the distal forelimb in horses walking on a sand track.
480 *Equine Veterinary Journal*, *38*, 377–382.
- 481 Clayton, H. M., Sha, D. H., Stick, J. A., & Robinson, P. (2007). 3D kinematics of the
482 interphalangeal joints in the forelimb of walking and trotting horses. *Veterinary*
483 *and Comparative Orthopaedics and Traumatology*, *20*, 1–7.
- 484 Clayton, H. M., Sha, D., Stick, J. a., & Mullineaux, D. R. (2004). Three-dimensional carpal
485 kinematics of trotting horses. *Equine Veterinary Journal*, *36*, 671–676.
- 486 Delp, S. L., & Loan, J. P. (1995). A graphics-based software system to develop and
487 analyze models of musculoskeletal structures. *Computers in Biology and*
488 *Medicine*, *25*, 21–34.
- 489 Ehrig, R. M., & Heller, M. O. (2019). On intrinsic equivalences of the finite helical axis ,
490 the instantaneous helical axis , and the SARA approach . A mathematical
491 perspective. *Journal of Biomechanics*, *84*, 4–10.
- 492 Grood, E. S., & Suntay, W. J. (1983). A Joint Coordinate System for the Clinical
493 Description of Three-Dimensional Motions: Application to the Knee. *Journal of*
494 *Biomechanical Engineering*, *105*, 136.
- 495 Holden, J. P., & Stanhope, S. J. (1998). The effect of variation in knee center location
496 estimates on net knee joint moments. *Gait Posture*, *7*, 1–6.
- 497 Kambic, R. E., Roberts, T. J., & Gatesy, S. M. (2017). 3-D range of motion envelopes
498 reveal interacting degrees of freedom in avian hind limb joints. *Journal of*
499 *Anatomy*, *231*, 906–920.

- 500 Khumsap, S., Clayton, H. M., Lanovaz, J. L., & Bouchey, M. (2002). Effect of walking
501 velocity on forelimb kinematics and kinetics. *Equine Veterinary Journal*.
502 *Supplement, 34*, 325–329.
- 503 Leach, D. H., & Dyson, S. J. (1988). Instant centres of rotation of equine limb joints and
504 their relationship to standard skin marker locations. *Equine Veterinary Journal*,
505 *Supplement*, 113–9.
- 506 Panagiotopoulou, O., Rankin, J. W., Gatesy, S. M., & Hutchinson, J. R. (2016). A
507 preliminary case study of the effect of shoe-wearing on the biomechanics of a
508 horse 's foot. *PeerJ*, *4*, 1–25.
- 509 Reinschmidt, C., & van den Bogert, A. J. (1997). KineMat: A MATLAB toolbox for the
510 reconstruction of spatial marker positions and for the analysis of three-
511 dimensional joint movements. *International Society of Biomechanics*,
512 [/http://www.isbweb.org/software/movanal/kinemat/](http://www.isbweb.org/software/movanal/kinemat/).
- 513 Roach, J. M., Pfau, T., Bryars, J., Unt, V., Channon, S. B., & Weller, R. (2014). Sagittal
514 distal limb kinematics inside the hoof capsule captured using high-speed
515 fluoroscopy in walking and trotting horses. *Veterinary Journal*, *202*, 94–98.
- 516 Ross, M. W., & Dyson, S. J. (2011). Chapter 26: The biomechanics of the equine limb
517 and its effect on lameness. In *Diagnosis and Management of Lameness in the*
518 *Horse, 2nd Edition* (Revised, pp. 270–281). London: Elsevier Health Sciences.
- 519 Seth, A., Sherman, M., Reinbolt, J. A., & Delp, S. L. (2011). OpenSim: A musculoskeletal
520 modeling and simulation framework for in silico investigations and exchange.
521 *Procedia IUTAM*, *2*, 212–232.
- 522 Shiavi, R., Limbird, T., Frazer, M., Stivers, K., Strauss, A., & Abramovitz, J. (1987). Helical
523 motion analyses of the knee - I. Methodology for studying kinematics during
524 locomotion. *Journal of Biomechanics*, *20*, 459–469.
- 525 Söderkvist, I., & Wedin, P.-Å. (1993). Determining the movements of the skeleton using
526 well-configured markers. *Journal of Biomechanics*, *26*, 1473–1477.
- 527 Spoor, C. W., & Veldpaus, F. E. (1980). Rigid body motion calculated from spatial co-
528 ordinates of markers. *Journal Biomechanics*, *13*, 391–393.
- 529 Stagni, R., Leardini, A., Cappozzo, A., Benedetti, M. G., & Cappello, A. (2000). E ! ects of
530 hip joint centre mislocation on gait analysis results. *Journal of Biomechanics*, *33*,
531 1479–1487.
- 532 Swanstrom, M. D. (1998). Joint Kinematics and Inertial Properties of the Thoroughbred
533 Forelimb, M.S. thesis, University of California, Davis, CA.
- 534 Swanstrom, M. D., Zarucco, L., Hubbard, M., Stover, S. M., & Hawkins, D. A. (2005).
535 Musculoskeletal Modeling and Dynamic Simulation of the Thoroughbred Equine
536 Forelimb During Stance Phase of the Gallop. *Journal of Biomechanical*
537 *Engineering*, *127*, 318.
- 538 Umberger, B. R., & Caldwell, G. E. (2013). Musculoskeletal modeling. In *Research*
539 *methods in biomechanics, second edition* (pp. 247–250). Champaign: Human
540 Kinetics Publishers.
- 541 van den Bogert, A. J., Reinschmidt, C., & Lundberg, A. (2008). Helical axes of skeletal
542 knee joint motion during running. *Journal of Biomechanics*, *41*, 1632–1638.
- 543 Weller, R., Pfau, T., May, S. A., & Wilson, A. M. (2006). Variation in conformation in a
544 cohort of National Hunt racehorses. *Equine Veterinary Journal*, *38*, 616–621.

- 545 Yalden, D. W. (1971). The functional morphology of the carpus in ungulate mammals.
546 *Acta Anat*, 78, 461–487.
- 547 Zatsiorsky, V. M. (1997). 2.1.5. Joint Rotation Convention. In *Kinematics of Human*
548 *Motion* (pp. 98–101).
- 549
- 550

551 Figure Legends

552 Fig 1. Schematic overview of the left equine forelimb. A) Schematic overview of the
553 forelimb bones. B) Schematic overview of the experimental cadaver limbs. Segment
554 boundaries are indicated with dark grey lines. Dashed red lines indicate the
555 standardised locations of the mid-level cuts through the soft tissue. Bone pins were
556 placed in the different forelimb bones as indicated in the figure. C) Overview of the
557 joint angles at neutral position as reported in Weller et al. 2006. Grey circles indicate
558 the extension angles and white circles indicate flexion angles of the joint.

559

560

561

562

563

564

565 Fig 2. Schematic overview of the standardized anatomical marker locations, A) shows
566 the marker locations in more detail on the dorsal joint surface and B) the overall
567 marker placement. Grey spheres indicate the locations of the anatomical markers,
568 marker numbers are shown inside the grey spheres. The origin of the bone is defined
569 by the position of anatomical marker 1. The proximodistal-axis, represented by the
570 blue arrow, is positive in a proximal direction. The green arrow represents the
571 mediolateral-axis which is positive towards the lateral side and the red arrow indicates
572 the craniocaudal/dorsopalmar-axis which is positive in a caudal/palmar direction.

573

574 Fig 3. Overview of the four FHA properties. The distance is defined as the distance
575 between the intersection of the FHA (black dot) with the plane perpendicular to the
576 FHA (grey) projected onto the proximodistal axis (blue) relative to the origin of the
577 proximal segment (grey sphere). A) For FE, FHA intersection (black dot) with the
578 sagittal plane (grey) projected onto the proximodistal axis (blue) for proximodistal
579 distance and projected onto the mediolateral axis (green) for the mediolateral
580 distance. B) For AA, FHA intersection (black dot) with the frontal plane (grey) projected
581 onto the proximodistal axis (blue) for proximodistal distance and projected onto the
582 cranial-caudal/dorsopalmar axis (red) for the cranial-caudal/dorsopalmar distance. C)
583 For IE, FHA intersection (black dot) with the transverse plane (grey) projected onto the
584 mediolateral axis (green) for mediolateral distance and projected onto the cranial-
585 caudal/dorsopalmar axis (red) for the cranial-caudal/dorsopalmar distance. D)
586 Deviation angle, the angle between the projection of the FHA (dashed line) onto the
587 transverse plane (grey) of the proximal segment and the mediolateral-axis of the
588 proximal segment (green arrow). E) Inclination angle, the angle between the projection
589 of the FHA (dashed line) onto the sagittal plane of the proximal segment and the
590 proximodistal-axis (grey) of the proximal segment.