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1 **Title:** Individuals with impaired lumbopelvic control demonstrate lumbar multifidus muscle
2 activation deficit using ultrasound imaging in conjunction with electrical stimulation: A cross-
3 sectional study.

4

5 Panakorn Sungnak, Sranya Songjaroen, Warin Krityakiarana, Hsing-Kuo Wang, Jim Richards,
6 Peemongkon Wattananon, PhD

7

8 **Abstract**

9 **Objective:** To determine lumbar multifidus muscle (LM) activation deficits in individuals with
10 impaired lumbopelvic control (iLPC) based on musculoskeletal ultrasound (MSKUS) in
11 conjunction with electrical stimulation approach, and the correlation between back extension force
12 and LM activation.

13 **Design:** A cross-sectional study design.

14 **Setting:** A university laboratory.

15 **Participants:** Fifty participants (25 iLPC and 25 NoLBP) were recruited from the university
16 physical therapy clinic and surrounding areas.

17 **Main Outcome Measures:** The MSKUS was used to measure LM thickness at rest, maximum
18 voluntary isometric contraction (MVIC), and electrical stimulation combined with MVIC, while a
19 hand-held dynamometer was used to record force during MVIC and electrical stimulation
20 combined with MVIC. These data were used to derive LM activation (LM_{ACT}) and percentage
21 force generation ($Force_{GEN}$).

1 **Results:** The iLPC group had significantly lower LM_{ACT} (17%) than the NoLBP group ($P<0.05$).

2 No significant difference was seen in Force_{GEN} between the NoLBP and iLPC groups ($P>0.05$).

3 No significant correlation was seen between LM_{ACT} and Force_{GEN} ($P>0.05$).

4 **Conclusion:** The findings support the utility of our protocol to determine LM activation deficits.

5 The lower LM activation in iLPC group suggests that individuals with iLPC were unable to fully

6 recruit the motor units available in LM. Force generation measurements may not be an appropriate

7 approach to determine such deficits in LM.

8

9 **Key words:** Impaired lumbopelvic control; Recurrent low back pain; Lumbar multifidus muscle;

10 Ultrasound imaging; Neuromuscular electrical stimulation; Muscle activation deficit

11

12 **List of abbreviation**

13 COMB Maximum voluntary isometric contraction combined with neuromuscular electrical

14 stimulation

15 ICC Intraclass correlation coefficient

16 iLPC Impaired lumbopelvic control

17 IQR Inter-quartile range

18 LBP Low back pain

19 LM Lumbar multifidus muscle

20 MDD₉₅ 95% confidence minimal detectable difference

21 MSKUS Musculoskeletal ultrasound

22 MVIC Maximum voluntary isometric contraction

23 NMES Neu

- 1 romuscular electrical stimulation
- 2 NoLBP Individuals without low back pain
- 3 rLBP Recurrent low back pain

1 INTRODUCTION

2 Mechanical low back pain (LBP) is one of the most common musculoskeletal conditions
3 worldwide.¹ Although clinical practice guideline recommends several interventions to improve
4 pain and disability, the recurrence rate is still high.^{1,2} This could be because mechanical LBP has
5 different underlying impairments.²⁻⁶

6 Clinical lumbar instability has been identified as a subgroup of LBP.^{7,8} This subgroup
7 demonstrates impaired lumbopelvic control (iLPC) represented by observed aberrant movement
8 patterns during functional movements.⁷⁻⁹ These repeated aberrant movements result in excessive
9 tissue stress and microtrauma which could further cause episodes of LBP.⁵ Furthermore, evidence
10 indicates the existence of these aberrant movements in individuals with both a history of LBP and
11 chronic LBP,^{4,9-11} indicating that iLPC may be responsible for recurrent LBP (rLBP).^{4,9-11}

12 One possible cause that could be responsible for such iLPC is arthrogenic muscle inhibition
13 of the lumbar multifidus muscles (LM).^{12,13} This reflex inhibition may reduce the ability of LM to
14 generate sufficient force to stabilize the lumbar spine; thereby, increasing the shear forces on the
15 lumbar spine resulting in tissue stress and microtrauma.^{5,12,13} It has been reported that the LM
16 provides up to two-thirds of the force generation needed to stabilize the lumbar spine.¹⁴ In addition,
17 the LM does not spontaneously recover after an episode of LBP, suggesting the existence of
18 continued deficits in LM activation.^{12,13,15-17} Although decreases in LM activity in individuals with
19 rLBP have been reported using surface and intramuscular electromyography,^{18,19} no study has
20 previously investigated the amount of such LM activation deficits.

21 Force production with neuromuscular electrical stimulation (NMES) is one of the most
22 common techniques used to quantify muscle activation deficits.²⁰⁻²² Previous studies have reported
23 that when participants were asked to perform maximum voluntary isometric contractions (MVIC)

1 and NMES was then applied to superimpose MVIC, this offers a better representation of the
2 maximum force from all motor units available in the muscle, which can be used to determine the
3 amount of muscle activation deficit.²⁰⁻²² However, this technique was used to measure limb muscle
4 activation, which can selectively investigate the muscle of interest. Therefore, this technique might
5 not be suitable to measure the LM deficits as the back extension force is generated from all the
6 back muscles, and not just LM. Accordingly, the overall force generated may not be correlated
7 with the LM activation.

8 To overcome the challenge of force production using the NMES technique,
9 musculoskeletal ultrasound (MSKUS) can be used in conjunction with the NMES to specifically
10 investigate LM activation deficit.²³ Several studies and our pilot study demonstrated association
11 between LM thickness measured by the MSKUS and LM activity using surface electromyography
12 suggesting the validity of the MSKUS to measure LM activation.²⁴⁻²⁷ Therefore, the MSKUS could
13 be used to fulfill the gap in the knowledge regarding the degree of LM activation deficit.

14 The objectives of this study were to determine the existence of LM activation deficit in
15 individuals with iLPC, and the correlation between back extension force and LM activation. We
16 hypothesized that the iLPC group would demonstrate lower LM activation compared with
17 individuals with no LBP (NoLBP), and that there would be no correlation between back extension
18 force and LM activation.

19 **METHODS**

20 **Participants**

21 Fifty participants aged between 18 and 40 years (25 iLPC and 25 NoLBP) were recruited
22 to the study. The inclusion criteria for iLPC were; at least two episodes of LBP that interfered with
23 activities of daily living or required treatment, but were pain free at the time of data collection,

1 presence of aberrant movement during active standing trunk flexion, and an average passive
2 straight leg raise of greater than 91 degrees. These inclusion criteria are supported by previous
3 literature that demonstrated that an age of less than 40 years, presence of aberrant movement, and
4 a passive straight leg raise greater than 91 degrees were predictors for iLPC.^{7,28,29} Aberrant
5 movements were defined as a painful arc in flexion or on return, Gower sign “thigh climbing”,
6 instability catch or reversal of lumbopelvic rhythm.¹⁷ Participants with iLPC were selected during
7 remission of pain to ensure that LM activation deficit was as a result of unrecovered LM, not just
8 the response to the pain. The inclusion criteria for NoLBP were; no previous episodes of LBP,
9 absence of aberrant movement, and a passive straight leg raise of less than 91 degrees. The
10 participants were excluded if they had clinical signs of systemic disease, definitive neurologic
11 signs including pain, previous spinal surgery, severe spinal stenosis and/or inflammatory joint
12 disease, and BMI greater than 30 kg/m². An initial pilot study demonstrated that the iLPC group
13 had lower LM activation than NoLBP with moderate effect size (Cohen’s $d = 0.73$). We used this
14 effect size with a one-tailed level of confidence with an alpha of 5% with a power of 80% to
15 calculate the sample size. This determined that fifty participants, 25 in each group, were required
16 for this study. Data were collected between June 2019 and March 2020.

17 **Instruments and measures**

18 A musculoskeletal ultrasound (MSKUS; model CX50, Philips, NV, USA) with a
19 broadband linear array transducer (12-3 MHz) was used to measure bilateral LM thickness at the
20 L4-5 facet joints (2 cm lateral to the lower half of the L4 spinous process).²³ Gain, frequency,
21 depth, and focus were adjusted by the researcher for each participant. Several studies support the
22 validity and reliability of the MSKUS to measure LM thickness in both individuals with and
23 without LBP.²³⁻²⁷ In addition, a hand-held dynamometer (Power Track II Jtech Commander, USA)

1 was used to measure back extension force, which was positioned and attached using straps at the
2 thoracic spine (T3). Studies supported the validity and reliability of the hand-held
3 dynamometer,^{30,31} and the pilot data demonstrated an excellent test-retest reliability of back
4 extension force measurements ($ICC_{3,2} = 0.94$).

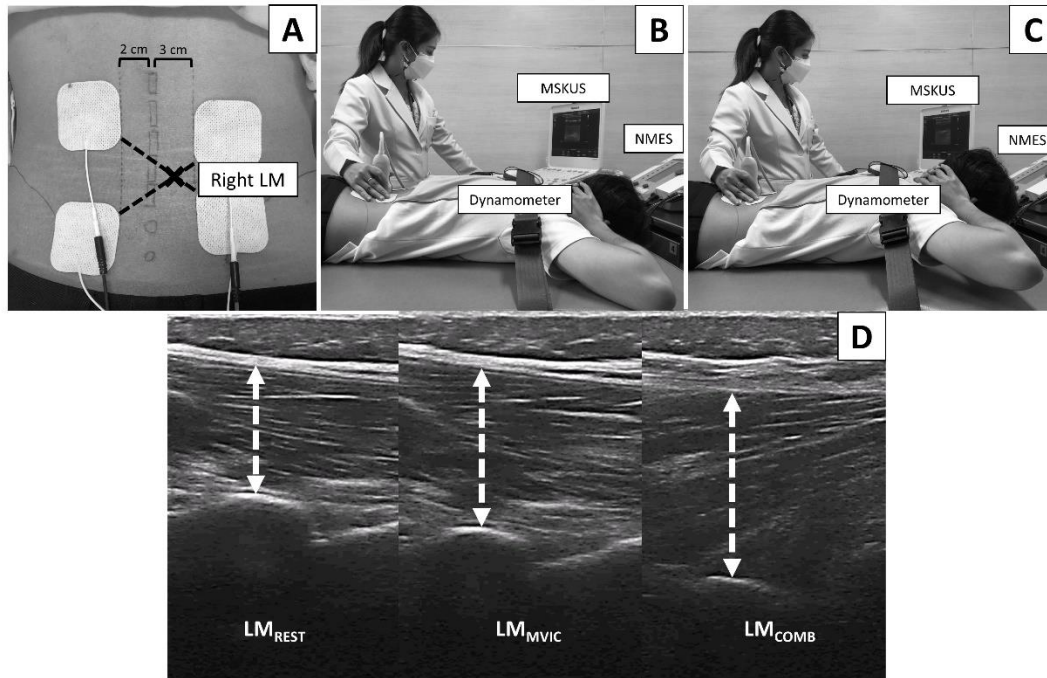
5 **Procedure**

6 This cross-sectional study was designed to explore the existence of LM activation deficits
7 in individuals with iLPC, and the correlation between back extension force and LM activation.
8 The study protocol was approved by the Mahidol University institutional review board (MU-CIRB
9 2018/215.0712). All participants provided written informed consent prior to data collection.
10 Demographic data from both groups and LBP behavior information from the iLPC group were
11 collected.

12 The researcher who performed the data collection was blinded to the group allocation. The
13 researcher received 3 hours of training by a MSKUS-certified expert and 50 hours of practice prior
14 to data collection. The participants were asked to position themselves in prone lying on a treatment
15 table, and the thorax (T3 level) and pelvis (S2 level) were then securely fastened to the table. A
16 hand-held dynamometer was placed at the thorax to measure back extension force. Neuromuscular
17 electrical stimulation (NMES; model Sonopuls 490 combination therapy, Enraf-Nonius BV,
18 Netherlands) was used to deliver an electrical current to superimpose the MVIC. To measure right
19 LM activation, a pair of adhesive electrodes (5cmX5cm) were attached at 3 cm lateral to the L3
20 and L5 spinous process levels on the right side, while another pair of electrodes were placed on
21 the left side thus creating two diagonal lines intersecting at the right side LM (Fig 1A). Then, the
22 ultrasound transducer was positioned in the longitudinal view at zero inclination on the L4-5 facet
23 joint on the right side (Fig 1B).

1 The participant was asked to put both their hands behind their neck and relax in the prone
2 position, while the researcher took two recordings of the LM thickness at rest (Fig 1B). The
3 participant was then asked to perform 2 repetitions of a back extension MVIC against the hand-
4 held dynamometer for 5 seconds with 1-minute rest between repetitions (Fig 1C; NMES: off). The
5 instruction was “Please lift your head, trunk, and upper extremities using your lower back with
6 maximum effort against the hand-held dynamometer attached on the strap while both ultrasound
7 and force data were simultaneously collected”. One practice trial was provided to ensure that the
8 participant extended from the lower back and performed the task correctly according to the
9 protocol. The MVIC value represents the participant’s ability to recruit motor units in the muscle.
10 Then, the NMES was applied to recruit additional motor units during MVIC. The NMES was set
11 at interferential mode (6000 Hz, beat frequency 20-50 Hz, scanning effect).³²⁻³⁴ The researcher
12 increased NMES intensity until the point of maximum tolerance. The participant was asked to
13 perform another 2 repetitions of a 5-second MVIC with NMES (Fig 1C; NMES: on), while LM
14 thickness and peak force were concurrently collected. The LM thickness values from resting,
15 MVIC and combined MVIC with NMES (Fig 1D) were then used to calculate the percentage of
16 muscle activation, a value of less than 100 indicating a muscle activation deficit.²⁰⁻²² A one-minute
17 rest period was provided between repetitions to prevent muscle fatigue. For the measurements
18 combining MVIC with NMES, the LM thickness was recorded from a 5-second video file as the
19 frequency was modulated between 20-50 Hz using the scanning technique. The entire process was
20 performed to measure left LM activation, while the NMES electrodes were re-attached to create
21 two diagonal lines intersecting over the LM on the left side. The overall protocol was
22 approximately 15 minutes.

23



1
 2 **Fig 1.** (A) Electrode placement for neuromuscular electrical stimulation (NMES) to activate right
 3 lumbar multifidus muscle (LM). (B) Example of participant's setup to measure LM thickness
 4 during resting (LM_{REST}) and (C) during maximum voluntary isometric contraction (LM_{MVIC}) and
 5 combined NMES with MVIC (LM_{COMB}). (D) LM thickness during rest, MVIC, and combined
 6 NMES with MVIC.

7

8 **Data reduction**

9 Data analysis was performed using a custom Matlab program (version R2014a, The
 10 MatWorks Inc, MA, USA). The researcher blinded to the group allocation measured the LM
 11 thickness from the resting and MVIC images (LM_{REST} and LM_{MVIC}), respectively. For the video
 12 files, the researcher selected the frames taken during the relaxation period and measured the LM
 13 thickness, then selected a frame showing maximum LM thickness during combined MVIC with
 14 NMES to measure LM thickness (LM_{COMB}). Our reliability analysis was based on all presented

1 data and showed excellent intra- and inter-rater reliability ($ICC_{3,1} = 0.97$ and $ICC_{2,2} = 0.95$,
 2 respectively) for LM thickness measurement, and the 95% confidence interval of the minimal
 3 detectable difference (MDD_{95}) was 0.08 cm. The averaged values from LM_{REST} , LM_{MVIC} , and
 4 LM_{COMB} across the two repetitions were used to calculate the percentage of LM activation (LM_{ACT})
 5 using the following formula.²³

$$6 \quad LM_{ACT} = \left(\frac{LM_{MVIC} - LM_{REST}}{LM_{COMB} - LM_{REST}} \right) X 100$$

7 A previous study has demonstrated that no significant side-to-side differences in LM
 8 morphology exist, even in the case of unilateral LBP.³⁵ In addition, our preliminary data analysis
 9 demonstrated no significant difference between right and left LM_{ACT} in the NoLBP group, the
 10 ipsilateral and contralateral sides in individuals presenting with unilateral pain, or between the
 11 right and left sides in individuals presenting with bilateral pain, therefore left and right averaged
 12 LM_{ACT} data were used for statistical analysis.

13 The back-extension force data were normalized to the participants body weight, and the
 14 averaged values across the two repetitions for MVIC and COMB ($Force_{MVIC}$ and $Force_{COMB}$), were
 15 taken respectively. These were then used to calculate the percentage force generation ($Force_{GEN}$)
 16 based on the formula below. The percentage force generation was used to compare between groups
 17 and determine the correlation with LM_{ACT} . Our pilot data demonstrated excellent test-retest
 18 reliability of the force measurements normalized to body weight during MVIC and COMB ($ICC_{3,1}$
 19 $= 0.95$ and 0.97), respectively, with MDD_{95} of 7.4 and 6.6% of body weight, respectively.

$$20 \quad Force_{GEN} = \left(\frac{Force_{MVIC}}{Force_{COMB}} \right) X 100$$

21 **Statistical analysis**

1 Statistical analysis was performed using SPSS version 21 (IBM Corp., NY, USA) in
2 conjunction with MDD₉₅. The distribution of the data was tested using Shapiro-Wilk tests, and the
3 LM thickness data were found to be normally distributed; therefore, a two-way mixed ANOVA
4 was used to determine the interaction between group and condition with planned post-hoc
5 comparisons between-group at rest, MVIC, and COMB, as well as within-group tests between rest
6 and MVIC, and between MVIC and COMB for each group. However, LM_{ACT} was found to be not
7 normally distributed; thus, a Mann-Whitney U test was performed to compare the LM activation
8 between groups. In addition, the normalized force data and percentage force generation were also
9 not normally distributed; therefore, Mann-Whitney U tests were performed to compare between
10 groups for MVIC and COMB conditions for the percentage force generation, and Wilcoxon tests
11 were used to compare within group differences between the MVIC and COMB data. Finally, a
12 Spearman's Rank correlation was used to determine the relationship between LM activation and
13 percentage force generation.

14 **RESULTS**

15 Demographic data are presented in Table 1. The two-way mixed ANOVA showed no
16 significant interactions between condition and group for the LM thickness, as well as no significant
17 main effect of group. However, a significant main effect was seen for condition ($P<0.05$). The
18 planned post-hoc comparisons (Table 2) revealed significant differences ($P<0.05$) in LM thickness
19 between LM_{REST} and LM_{MVIC}, as well as LM_{MVIC} and LM_{COMB} with differences greater than
20 MDD₉₅ (0.08 cm). LM_{REST} did not show a significant difference between groups ($P>0.05$).

21

22

23

1 **Table 1.** Demographic data

Parameter	NoLBP	iLPC
Age (years)	22.2±2.3	22.8±2.6
BMI (kg/m ²)	22.0±1.6	21.4±2.3
Sex (%female)	60	60
Duration (months)	N/A	36.9±28.3
Frequency per year (number of episodes)	N/A	11.2±13.5
Time since last episode (days)	N/A	24.1±24.2
Duration for last episode (days)	N/A	2.8±2.8
Pain at last episode (0=no pain, 10=intolerable pain)	N/A	4.5±1.4
Disability at last episode (0=no disability, 10=total disability)	N/A	3.2±1.7

2 NoLBP = no history of low back pain in lifetime, iLPC = impaired lumbopelvic control, BMI =
3 body mass index.

4

5 **Table 2.** Planned post-hoc pairwise comparisons between groups (mean±SD) for lumbar
6 multifidus muscle thickness at rest (LM_{REST}), maximum voluntary isometric contraction
7 (LM_{MVIC}), and combined MVIC with neuromuscular electrical stimulation (LM_{COMB}), as well as
8 percentage lumbar multifidus muscle activation (LM_{ACT}) between groups (median (IQR))

Parameter	NoLBP	iLPC	Between-group mean/median diff (NoLBP vs iLPC)
LM _{REST} (cm)	2.64±0.46	2.58±0.55	0.06
LM _{MVIC} (cm)	3.39±0.56	3.28±0.64	0.11 [†]
LM _{COMB} (cm)	3.51±0.57	3.55±0.63	0.04
Within-group mean diff (LM _{REST} vs LM _{MVIC})	0.76* [†]	0.70* [†]	
Within-group mean diff (LM _{MVIC} vs LM _{COMB})	0.12* [†]	0.27* [†]	
LM _{ACT} (%activation)	91.6 (82.7,93.8)	75.9 (64.0,83.3)	15.7*

9 NoLBP = no history of low back pain group, iLPC = impaired lumbopelvic control group, SD =
10 standard deviation, IQR = interquartile range

11 * = significant difference ($P < 0.05$)

12 [†] = exceeded 95% confidence minimal detectable difference

13

14

15 For LM_{ACT}, the results demonstrated that the iLPC group had significantly lower values
16 than the NoLBP group ($P < 0.05$); with a median = 75.9, interquartile range; IQR = 64.0 and 83.3
17 for the iLPC group, and median = 91.6, IQR = 82.7 and 93.8 for the NoLBP group, respectively.

1 The non-parametric Mann-Whitney U and Wilcoxon tests did not show any significant
 2 differences ($P>0.05$) in normalized force generation between-group and within-group comparisons
 3 (Table 3), respectively, and the median differences did not exceed MDD_{95} . In addition, no
 4 significant difference was seen in $Force_{GEN}$ between the NoLBP and iLPC groups; with a median
 5 = 101.2, IQR = 88.4 and 109.4 for the NoLBP group, and median = 95.7, IQR = 87.5 and 102.8
 6 for the iLPC group. In addition, the Spearman's Rank correlation coefficient demonstrated no
 7 significant correlation between LM_{ACT} and $Force_{GEN}$ ($\rho=0.18$, $P>0.05$).

8
 9 **Table 3.** Comparisons between groups (median (IQR)) for normalized force generation at
 10 maximum voluntary isometric contraction (F_{MVIC}), and combined MVIC with peripheral electrical
 11 stimulation (F_{COMB}), as well as the percentage force generation (F_{GEN}) between groups (median
 12 (IQR))

Parameter	NoLBP	iLPC	Between-group median diff (NoLBP vs iLPC)
F_{MVIC} (%body weight)	20.5 (17.1, 26.9)	24.0 (17.4, 25.9)	3.5
F_{COMB} (%body weight)	20.7 (16.0, 26.2)	23.6 (16.9, 28.3)	2.9
Within-group median diff (F_{MVIC} vs F_{COMB})	0.2	-0.4	
F_{GEN} (%force generation)	101.2 (88.4, 109.4)	95.7 (87.5, 102.8)	5.5

13 NoLBP = no history of low back pain group, iLPC = impaired lumbopelvic control group, IQR =
 14 interquartile range

15
 16

17 DISCUSSION

18 The significant findings from LM thickness for within-group comparisons (LM_{REST} vs
 19 LM_{MVIC} , and LM_{MVIC} vs LM_{COMB}) exceeded the MDD_{95} , this suggests a true change between
 20 resting and MVIC, and NMES concurrently with MVIC which was shown to elicit a greater LM
 21 activation.²⁰⁻²² These findings support the utility of our protocol to determine LM activation.
 22 Although there were no significant differences in LM thickness between groups in any condition,
 23 a within-group difference demonstrated the pattern in which the NoLBP group showed a greater

1 increased LM thickness from rest to MVIC than the iLPC group, while the iLPC group had a
2 greater LM thickness change from MVIC to COMB than the NoLBP group. These findings suggest
3 that the NoLBP group could increase LM activation volitionally, while the iLPC group required
4 the NMES to facilitate LM activation.

5 The main findings from the LM activation supports our hypothesis in which individuals
6 with iLPC had lower LM activation than those without LBP. This indicates the perseverance of
7 LM activation deficit in individuals with iLPC.^{12,13,15-17} This lower LM activation in iLPC group
8 suggests that individuals with iLPC were unable to fully recruit all motor units available in the
9 LM. This muscle activation deficit in the LM could be as a result of the persistence of a reflex
10 inhibition mechanism.^{12,13} This reflex inhibition mechanism initially occurs after injury to the
11 tissue in the lumbar region,^{12,13} resulting in abnormal afferent signals from damaged tissues which
12 in turn reduce the motor commands to the LM.^{12,13} This mechanism inhibits the individual's ability
13 to fully recruit all motor units available in the LM.^{12,13} As a result, the LM might not be able to
14 provide sufficient force to stabilize the lumbar spine, thereby making the individual susceptible to
15 re-injury of the lumbar spine.^{5-8,12}

16 Fatty infiltration or muscle atrophy could be another potential factor that reduces the
17 contractility of the LM resulting in decreased LM activation in individuals with iLPC,^{15,36-40} which
18 in turn can reduce lumbar stability. In addition, the LM activation deficit may also be related to
19 changes in corticospinal mapping of the LM.⁴¹ This change at the cortical level may compromise
20 the ability to control and refine LM contractions,⁴¹ which may be represented as LM activation
21 deficits in our study. One study reported that NMES can send ascending activation to the
22 sensorimotor cortex in the brain during electrical stimulation.⁴² This mechanism has been shown
23 to enhance the motor unit recruitment during combined MVIC with NMES.⁴²

1 No study has previously used MSKUS in conjunction with the NMES to determine the LM
2 activation. Therefore, we do not have results from other studies to directly compare the LM
3 activation with ours. The finding in percentage LM activation is consistent with previous studies
4 using surface electromyography in which they have found that patients with chronic/recurrent LBP
5 had lower LM activation when compared with healthy individuals.^{18,19} This can be considered as
6 a confirmatory study for this point. Although using EMG amplitude can be interpreted as muscle
7 activation deficit,^{18,19} this approach failed to determine the extent of this deficit. Our approach uses
8 the superimposition technique in which the NMES passively helps the individual to recruit more
9 motor units in the LM.^{10,13,29} Therefore, the percentage LM activation derived from our approach
10 could be used to indirectly determine the extent of any deficit.²⁰⁻²² In this study, individuals with
11 iLPC had approximately 17% less LM activation when compared to individuals with NoLBP.

12 Although not significant the difference in LM thickness during MVIC exceeded the
13 MDD₉₅, this may be as a result of an underpowered statistical analysis. Our sample size calculation
14 was based on a large effect size from our pilot study which may have inflated the sample size.
15 Another potential explanation could be that LM fibers have transformed to type II fibers (fast-
16 twitch fibers) which have larger diameter causing greater thickness change during cross-
17 bridge.^{39,42} Studies using MRI and muscle biopsy demonstrated a lower proportion of type I fibers
18 and a greater proportion of type II fibers in patients with chronic LBP.^{39,42} This transformation
19 could be a coping strategy to compensate for insufficient force from type I fibers. Although this
20 strategy is beneficial to stabilize the lumbar spine, muscle fiber type II is more susceptible to
21 muscle fatigue.¹² This mechanism might be responsible for recurrence of LBP symptoms.
22 However, our study did not have the data to confirm our interpretation based on muscle fiber types.
23 Future studies to investigate the LM muscle type in individuals with iLPC is required.

1 As we expected, the results did not show a significantly lower force generation in the iLPC
2 group when compared with the NoLBP group. These non-significant results could be resulted from
3 the weakness of the approach using normalized force with NMES superimposition. Our results did
4 not show significant increase in normalized force during combined MVIC with NMES comparing
5 with MVIC in both groups even though we found a significant increase in LM thickness.

6 In addition, no correlation between LM activation and percentage force generation could
7 be another potential explanation for non-significant difference in force generation. Based on basic
8 anatomy, the LM has a short lever arm spanning only 1-2 vertebral segments.^{12,14} Therefore, the
9 LM might not generate greater force even though the thickness change was significant. In addition,
10 the LM is primarily responsible for providing lumbar stability, rather than generating a force to
11 move the spine into extension.^{12,14} Another potential explanation is that our force measurement
12 does not offer a representation of the LM function.²³ The force can be viewed as an output from
13 all back-extensor muscles, and not specifically from LM.

14 Our approach described in this study was reliable, and our findings support the validity for
15 LM activation measurement in which our approach had the ability to differentiate the amount of
16 LM activation between individuals with iLPC and no LBP. Our findings suggest the clinical
17 application to evaluate LBP patients in an iLPC subgroup. This approach could be used in future
18 studies to determine the ability of therapeutic exercises to restore LM activation.

19 **Study limitations**

20 Some limitations should be taken into consideration in this study. Firstly, we used an iLPC
21 subgroup of LBP, which limits the generalizability of these results to the wider LBP population.
22 Secondly, we did not control the characteristics (pain intensity, disability level, etc.) of the
23 participants with iLPC. These characteristics may affect our outcome measures, although these are

1 similar to those reported in the literature. Participants in this study did not have current pain,
2 therefore, we were unable to determine the role of pain during LM activation. In addition, we did
3 not collect fear avoidance behavior which may be associated with the memory of pain during
4 muscle contractions. Therefore, future studies should take pain and the memory of pain into
5 consideration. Another limitation is that we have assumed that the LM thickness can be used to
6 represent the LM motor unit activation. The LM thickness data can be confounded by the LM
7 physiological changes (e.g. fatty infiltration, muscle fiber type, etc.). We used a symmetrical task
8 for LM activation, however this might not be the best task to evaluate LM activation, and several
9 studies have reported using contralateral arm elevation against a force to activate LM function.
10 Therefore, future studies should consider using asymmetrical tasks when measuring LM thickness
11 as suggested by Sweeney et al.³⁸ In addition, this study did not include an investigation of the
12 motor unit behavior. Recent studies have shown the utility of decomposition EMG to directly
13 measure the motor unit behavior which may yield more information on any possible activation
14 deficits in the LM.⁴³

15 **CONCLUSION**

16 This is the first study using the NMES in conjunction with MSKUS and force measurement
17 to investigate LM activation and back extensor force generation in individuals with iLPC and
18 NoLBP and determine the correlation between LM activation and force generation. We found that
19 the iLPC group had lower LM activation than the NoLBP group indicating the perseverance of an
20 LM activation deficit. However, we did not find a difference in force generation between the
21 groups, nor a correlation between LM activation and force generation. The lack of a significant
22 correlation could be due to the fact that the force generation is an output from all back extensor

- 1 muscles, and not specifically from LM. These findings suggest that force generation measurements
- 2 may not be an appropriate approach to determine LM activation.

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3 the data collection space and equipment. We would also like to thank all participants in this study.

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